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[Continued on next page]

(54) Title: COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF BREAST CANCER

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1  [MSEFQGLQYQYIRFLYFPERFQAAGUGGCGUSSAGURSGFQAGASELAASGS]  H1002Cor1.PRO  iroquois.PRO
1  [LSNVLSVYIGAPYAAAAAAGCYGAFLPYATELRIFPQLQAUVELKRL]  H1002Cor1.PRO  iroquois.PRO
51  [SPGVQHPTAAAFPHFPAFEPYGVVQFQPSRPHAAFRKSTSTLRWLH]  H1002Cor1.PRO  iroquois.PRO
1  [GHEKNFYPTKGEKMLAETATHTLTQVSTMFANRRRLKKEKHTHAPKS]  H1002Cor1.PRO  iroquois.PRO
151  [RDERGHATGSEEREDEDEDEFEESKKELEHREERLAGRGCGGCGGAGGR]  H1002Cor1.PRO  iroquois.PRO
1  [RRRDEEIDLENLPSAAAAGSELTLAQAHHHGGDFGLHDSDEKTSLSUDEG]  H1002Cor1.PRO  iroquois.PRO
11  [ERGAGRO]SUGREEGARREREGU[ERHVVHOSLDTCAFAFA]SALOKP  H1002Cor1.PRO  iroquois.PRO
101  [GLEDRFLSVLSLAPFFPVARATASFTSPSSLDPCAFAPAFSSALOKP]  H1002Cor1.PRO  iroquois.PRO
61  [KINSLASTATSTDPNRRSPFPAAGSPFFGAAVA]SALDLSFAAAAAA  H1002Cor1.PRO  iroquois.PRO
151  [KINSLASTATSTDPNRRSPFPAAGSPFFGAAVA]PTLQLSPAAAAA  H1002Cor1.PRO  iroquois.PRO
109  [PLVSAPLGRFPANTNRFFPQF]PQFFPHPLSLHDSAPFHNLGLFGAAH  H1002Cor1.PRO  iroquois.PRO
401  [PLVSAPLGRFPANTNRFFPQF]PQFFPHPLSLHDSAPFHNLGLFGAAH  H1002Cor1.PRO  iroquois.PRO
159  [AAAAA]FARFAFELGQTDHCHALEVERKLELTAFQVVPFPPQD  H1002Cor1.PRO  iroquois.PRO
451  [AAAAA]YARFAFELGQTDHCHALEVERKLELTAFQVVPFPPQD  H1002Cor1.PRO  iroquois.PRO
209  [LSALSS]  H1002Cor1.PRO  iroquois.PRO
301  [LSALSS]  H1002Cor1.PRO  iroquois.PRO
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(57) Abstract: Compositions and methods for the therapy and diagnosis of cancer, such as breast cancer, are disclosed. Compositions may comprise one or more breast tumor proteins, immunogenic portions thereof, or polynucleotides that encode such portions. Alternatively, a therapeutic composition may comprise an antigen presenting cell that expresses a breast tumor protein, or a T cell that is specific for cells expressing such a protein. Such compositions may be used, for example, for the prevention and treatment of diseases such as breast cancer. Diagnostic methods based on detecting a breast tumor protein, or mRNA encoding such a protein, in a sample are also provided.

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COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF BREAST CANCER

TECHNICAL FIELD OF THE INVENTION

The present invention relates generally to therapy and diagnosis of
5 cancer, such as breast cancer. The invention is more specifically related to polypeptides
comprising at least a portion of a breast tumor protein, and to polynucleotides encoding
such polypeptides. Such polypeptides and polynucleotides may be used in vaccines and
pharmaceutical compositions for prevention and treatment of breast cancer, and for the
diagnosis and monitoring of such cancers.

10 BACKGROUND OF THE INVENTION

Breast cancer is a significant health problem for women in the United
States and throughout the world. Although advances have been made in detection and
treatment of the disease, breast cancer remains the second leading cause of cancer-
related deaths in women, affecting more than 180,000 women in the United States each
15 year. For women in North America, the life-time odds of getting breast cancer are now
one in eight.

No vaccine or other universally successful method for the prevention or
treatment of breast cancer is currently available. Management of the disease currently
relies on a combination of early diagnosis (through routine breast screening procedures)
20 and aggressive treatment, which may include one or more of a variety of treatments
such as surgery, radiotherapy, chemotherapy and hormone therapy. The course of
treatment for a particular breast cancer is often selected based on a variety of prognostic
parameters, including an analysis of specific tumor markers. *See, e.g., Porter-Jordan
and Lippman, Breast Cancer 8:73-100, 1994.* However, the use of established markers
25 often leads to a result that is difficult to interpret, and the high mortality observed in
breast cancer patients indicates that improvements are needed in the treatment,
diagnosis and prevention of the disease.

Immunotherapies have the potential to substantially improve breast
cancer treatment and survival. Such therapies may involve the generation or

enhancement of an immune response to a breast tumor antigen. However, to date, relatively few breast tumor antigens are known and the generation of an immune response against such antigens has not been shown to be therapeutically beneficial.

Accordingly, there is a need in the art for improved methods for
5 identifying breast tumor antigens and for using such antigens in the diagnosis and therapy of breast cancer. The present invention fulfills these needs and further provides other related advantages.

SUMMARY OF THE INVENTION

Briefly stated, the present invention provides compositions and methods
10 for the diagnosis and therapy of cancer, such as breast cancer. In one aspect, the present invention provides polypeptides comprising at least a portion of a breast tumor protein, or a variant thereof. Certain portions and other variants are immunogenic, such that the ability of the variant to react with antigen-specific antisera is not substantially diminished. Within certain embodiments, the polypeptide comprises a sequence that is
15 encoded by a polynucleotide sequence selected from the group consisting of sequences recited in SEQ ID NOs:1-125, variants of such sequences and complements of such sequences. One such polypeptide comprises a sequence recited in SEQ ID NO:126, or a variant thereof that is at least 90% identical to SEQ ID NO:126.

The present invention further provides polynucleotides that encode a
20 polypeptide as described above, or a portion thereof (such as a portion encoding at least 9, preferably at least 15, amino acid residues of a breast tumor protein), expression vectors comprising such polynucleotides and host cells transformed or transfected with such expression vectors.

Within other aspects, the present invention provides pharmaceutical
25 compositions comprising a polypeptide or polynucleotide as described above and a physiologically acceptable carrier.

Within a related aspect of the present invention, vaccines are provided. Such vaccines comprise a polypeptide or polynucleotide as described above and a non-specific immune response enhancer.

The present invention further provides pharmaceutical compositions that comprise: (a) an antibody or antigen-binding fragment thereof that specifically binds to a breast tumor protein; and (b) a physiologically acceptable carrier.

5 Within further aspects, the present invention provides pharmaceutical compositions comprising: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) a pharmaceutically acceptable carrier or excipient. Antigen presenting cells include dendritic cells, macrophages and B cells.

10 Within related aspects, vaccines are provided that comprise: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) a non-specific immune response enhancer.

The present invention further provides, in other aspects, fusion proteins that comprise at least one polypeptide as described above, as well as polynucleotides encoding such fusion proteins.

15 Within related aspects, pharmaceutical compositions comprising a fusion protein, or a polynucleotide encoding a fusion protein, in combination with a physiologically acceptable carrier are provided.

Vaccines are further provided, within other aspects, that comprise a fusion protein or a polynucleotide encoding a fusion protein in combination with a non-specific immune response enhancer.

20 Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as recited above.

25 The present invention further provides, within other aspects, methods for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with a breast tumor protein, wherein the step of contacting is performed under conditions and for a time sufficient to permit the removal of cells expressing the protein from the sample.

30 Within related aspects, methods are provided for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated as described above.

Methods are further provided, within other aspects, for stimulating and/or expanding T cells specific for a breast tumor protein, comprising contacting T cells with one or more of: (i) a polypeptide as described above; (ii) a polynucleotide encoding such a polypeptide; and/or (iii) an antigen presenting cell that expresses such a polypeptide; under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells. Isolated T cell populations comprising T cells prepared as described above are also provided.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a T cell population as described above.

The present invention further provides methods for inhibiting the development of a cancer in a patient, comprising the steps of: (a) incubating CD4⁺ and/or CD8⁺ T cells isolated from a patient with one or more of: (i) a polypeptide comprising at least an immunogenic portion of a breast tumor protein; (ii) a polynucleotide encoding such a polypeptide; and (iii) an antigen-presenting cell that expresses such a polypeptide; and (b) administering to the patient an effective amount of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient. Proliferated cells may, but need not, be cloned prior to administration to the patient.

Within further aspects, the present invention provides methods for determining the presence or absence of a cancer in a patient, comprising (a) contacting a biological sample obtained from a patient with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; and (c) comparing the amount of polypeptide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within preferred embodiments, the binding agent is an antibody, more preferably a monoclonal antibody. The cancer may be breast cancer.

The present invention also provides, within other aspects, methods for monitoring the progression of a cancer in a patient. Such methods comprise the steps of: (a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a polypeptide as recited above; (b) detecting in the

sample an amount of polypeptide that binds to the binding agent; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polypeptide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the
5 patient.

The present invention further provides, within other aspects, methods for determining the presence or absence of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein; (b) detecting in the
10 sample a level of a polynucleotide, preferably mRNA, that hybridizes to the oligonucleotide; and (c) comparing the level of polynucleotide that hybridizes to the oligonucleotide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within certain embodiments, the amount of mRNA is detected via polymerase chain reaction using, for example, at least one
15 oligonucleotide primer that hybridizes to a polynucleotide encoding a polypeptide as recited above, or a complement of such a polynucleotide. Within other embodiments, the amount of mRNA is detected using a hybridization technique, employing an oligonucleotide probe that hybridizes to a polynucleotide that encodes a polypeptide as recited above, or a complement of such a polynucleotide.

20 In related aspects, methods are provided for monitoring the progression of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein; (b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; (c) repeating steps (a) and (b)
25 using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polynucleotide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

Within further aspects, the present invention provides antibodies, such as
30 monoclonal antibodies, that bind to a polypeptide as described above, as well as

diagnostic kits comprising such antibodies. Diagnostic kits comprising one or more oligonucleotide probes or primers as described above are also provided.

These and other aspects of the present invention will become apparent upon reference to the following detailed description and attached drawings. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

BRIEF DESCRIPTION OF THE DRAWING

Figure 1 represents a sequence alignment between the representative breast tumor polypeptide B1002C (SEQ ID NO:126) and the mouse iroquois homeobox protein 3.

DETAILED DESCRIPTION OF THE INVENTION

As noted above, the present invention is generally directed to compositions and methods for using the compositions, for example in the therapy and diagnosis of cancer, such as breast cancer. Certain illustrative compositions described herein include breast tumor polypeptides, polynucleotides encoding such polypeptides, binding agents such as antibodies, antigen presenting cells (APCs) and/or immune system cells (*e.g.*, T cells). A "breast tumor protein," as the term is used herein, refers generally to a protein that is expressed in breast tumor cells at a level that is at least two fold, and preferably at least five fold, greater than the level of expression in a normal tissue, as determined using a representative assay provided herein. Certain breast tumor proteins are tumor proteins that react detectably (within an immunoassay, such as an ELISA or Western blot) with antisera of a patient afflicted with breast cancer.

Therefore, in accordance with the above, and as described further below, the present invention provides illustrative polynucleotide compositions having sequences set forth in SEQ ID NO:1-125, illustrative polypeptide compositions having amino acid sequences set forth in SEQ ID NO:126, antibody compositions capable of binding such polypeptides, and numerous additional embodiments employing such compositions, for example in the detection, diagnosis and/or therapy of human breast cancer.

POLYNUCLEOTIDE COMPOSITIONS

As used herein, the terms "DNA segment" and "polynucleotide" refer to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment encoding a polypeptide refers to a DNA segment
5 that contains one or more coding sequences yet is substantially isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment is obtained. Included within the terms "DNA segment" and "polynucleotide" are DNA segments and smaller fragments of such segments, and also recombinant vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, and the like.

10 As will be understood by those skilled in the art, the DNA segments of this invention can include genomic sequences, extra-genomic and plasmid-encoded sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally isolated, or modified synthetically by the hand of man.

15 "Isolated," as used herein, means that a polynucleotide is substantially away from other coding sequences, and that the DNA segment does not contain large portions of unrelated coding DNA, such as large chromosomal fragments or other functional genes or polypeptide coding regions. Of course, this refers to the DNA segment as originally isolated, and does not exclude genes or coding regions later added
20 to the segment by the hand of man.

As will be recognized by the skilled artisan, polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA molecules. RNA molecules include HnRNA molecules, which contain introns and correspond to a DNA molecule in a one-to-one manner, and
25 mRNA molecules, which do not contain introns. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

Polynucleotides may comprise a native sequence (*i.e.*, an endogenous
30 sequence that encodes a breast tumor protein or a portion thereof) or may comprise a variant, or a biological or antigenic functional equivalent of such a sequence.

Polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions, as further described below, preferably such that the immunogenicity of the encoded polypeptide is not diminished, relative to a native tumor protein. The effect on the immunogenicity of the encoded polypeptide may generally be assessed as
5 described herein. The term "variants" also encompasses homologous genes of xenogenic origin.

When comparing polynucleotide or polypeptide sequences, two sequences are said to be "identical" if the sequence of nucleotides or amino acids in the two sequences is the same when aligned for maximum correspondence, as described
10 below. Comparisons between two sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A "comparison window" as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to about 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions
15 after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment schemes described in the following references: Dayhoff, M.O. (1978) A
20 model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) Atlas of Protein Sequence and Structure, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenesis pp. 626-645 *Methods in Enzymology* vol. 183, Academic Press, Inc., San Diego, CA; Higgins, D.G. and Sharp, P.M. (1989)
25 *CABIOS* 5:151-153; Myers, E.W. and Muller W. (1988) *CABIOS* 4:11-17; Robinson, E.D. (1971) *Comb. Theor* 11:105; Santou, N. Nes, M. (1987) *Mol. Biol. Evol.* 4:406-425; Sneath, P.H.A. and Sokal, R.R. (1973) *Numerical Taxonomy – the Principles and Practice of Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and Lipman, D.J. (1983) *Proc. Natl. Acad., Sci. USA* 80:726-730.

30 Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL*.

Math 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity methods of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul *et al.* (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent sequence identity for the polynucleotides and polypeptides of the invention. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. In one illustrative example, cumulative scores can be calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). For amino acid sequences, a scoring matrix can be used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, and expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915) alignments, (B) of 50, expectation (E) of 10, M=5, N=-4 and a comparison of both strands.

Preferably, the "percentage of sequence identity" is determined by comparing two optimally aligned sequences over a window of comparison of at least 20 positions, wherein the portion of the polynucleotide or polypeptide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference

sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid bases or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the
5 total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

Therefore, the present invention encompasses polynucleotide and polypeptide sequences having substantial identity to the sequences disclosed herein, for example those comprising at least 50% sequence identity, preferably at least 55%, 60%,
10 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher, sequence identity compared to a polynucleotide or polypeptide sequence of this invention using the methods described herein, (*e.g.*, BLAST analysis using standard parameters, as described below). One skilled in this art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two
15 nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like.

In additional embodiments, the present invention provides isolated polynucleotides and polypeptides comprising various lengths of contiguous stretches of sequence identical to or complementary to one or more of the sequences disclosed
20 herein. For example, polynucleotides are provided by this invention that comprise at least about 15, 20, 30, 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more contiguous nucleotides of one or more of the sequences disclosed herein as well as all intermediate lengths there between. It will be readily understood that "intermediate lengths", in this context, means any length between the quoted values, such as 16, 17,
25 18, 19, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103, *etc.*; 150, 151, 152, 153, *etc.*; including all integers through 200-500; 500-1,000, and the like.

The polynucleotides of the present invention, or fragments thereof, regardless of the length of the coding sequence itself, may be combined with other
30 DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their

overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For example, illustrative DNA segments with total lengths of about 10,000, about 5000,
5 about 3000, about 2,000, about 1,000, about 500, about 200, about 100, about 50 base pairs in length, and the like, (including all intermediate lengths) are contemplated to be useful in many implementations of this invention.

In other embodiments, the present invention is directed to polynucleotides that are capable of hybridizing under moderately stringent conditions to
10 a polynucleotide sequence provided herein, or a fragment thereof, or a complementary sequence thereof. Hybridization techniques are well known in the art of molecular biology. For purposes of illustration, suitable moderately stringent conditions for testing the hybridization of a polynucleotide of this invention with other polynucleotides include prewashing in a solution of 5 X SSC, 0.5% SDS, 1.0 mM
15 EDTA (pH 8.0); hybridizing at 50°C-65°C, 5 X SSC, overnight; followed by washing twice at 65°C for 20 minutes with each of 2X, 0.5X and 0.2X SSC containing 0.1% SDS.

Moreover, it will be appreciated by those of ordinary skill in the art that, as a result of the degeneracy of the genetic code, there are many nucleotide sequences
20 that encode a polypeptide as described herein. Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention. Further, alleles of the genes comprising the polynucleotide sequences provided herein are within the scope of the present invention.
25 Alleles are endogenous genes that are altered as a result of one or more mutations, such as deletions, additions and/or substitutions of nucleotides. The resulting mRNA and protein may, but need not, have an altered structure or function. Alleles may be identified using standard techniques (such as hybridization, amplification and/or database sequence comparison).

PROBES AND PRIMERS

In other embodiments of the present invention, the polynucleotide sequences provided herein can be advantageously used as probes or primers for nucleic acid hybridization. As such, it is contemplated that nucleic acid segments that comprise
5 a sequence region of at least about 15 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 15 nucleotide long contiguous sequence disclosed herein will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000 (including all intermediate lengths) and even up to full length sequences will also be of
10 use in certain embodiments.

The ability of such nucleic acid probes to specifically hybridize to a sequence of interest will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are also envisioned, such as the use of the sequence information for the preparation of mutant species
15 primers, or primers for use in preparing other genetic constructions.

Polynucleotide molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so (including intermediate lengths as well), identical or complementary to a polynucleotide sequence disclosed herein, are particularly contemplated as
20 hybridization probes for use in, *e.g.*, Southern and Northern blotting. This would allow a gene product, or fragment thereof, to be analyzed, both in diverse cell types and also in various bacterial cells. The total size of fragment, as well as the size of the complementary stretch(es), will ultimately depend on the intended use or application of the particular nucleic acid segment. Smaller fragments will generally find use in
25 hybridization embodiments, wherein the length of the contiguous complementary region may be varied, such as between about 15 and about 100 nucleotides, but larger contiguous complementarity stretches may be used, according to the length complementary sequences one wishes to detect.

The use of a hybridization probe of about 15-25 nucleotides in length
30 allows the formation of a duplex molecule that is both stable and selective. Molecules having contiguous complementary sequences over stretches greater than 15 bases in

length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 15 to 25 contiguous nucleotides, or even longer where
5 desired.

Hybridization probes may be selected from any portion of any of the sequences disclosed herein. All that is required is to review the sequence set forth in SEQ ID NO:1-125, or to any continuous portion of the sequence, from about 15-25 nucleotides in length up to and including the full length sequence, that one wishes to
10 utilize as a probe or primer. The choice of probe and primer sequences may be governed by various factors. For example, one may wish to employ primers from towards the termini of the total sequence.

Small polynucleotide segments or fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, as is commonly
15 practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Patent 4,683,202 (incorporated herein by reference), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA techniques generally known to those of skill in the art of molecular
20 biology.

The nucleotide sequences of the invention may be used for their ability to selectively form duplex molecules with complementary stretches of the entire gene or gene fragments of interest. Depending on the application envisioned, one will typically desire to employ varying conditions of hybridization to achieve varying degrees of
25 selectivity of probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, *e.g.*, one will select relatively low salt and/or high temperature conditions, such as provided by a salt concentration of from about 0.02 M to about 0.15 M salt at temperatures of from about 50°C to about 70°C. Such selective conditions tolerate
30 little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating related sequences.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template, less stringent (reduced stringency) hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one
5 may desire to employ salt conditions such as those of from about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to
10 destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

POLYNUCLEOTIDE IDENTIFICATION AND CHARACTERIZATION

Polynucleotides may be identified, prepared and/or manipulated using
15 any of a variety of well established techniques. For example, a polynucleotide may be identified, as described in more detail below, by screening a microarray of cDNAs for tumor-associated expression (*i.e.*, expression that is at least two fold greater in a tumor than in normal tissue, as determined using a representative assay provided herein). Such screens may be performed, for example, using a Synteni microarray (Palo Alto,
20 CA) according to the manufacturer's instructions (and essentially as described by Schena *et al.*, *Proc. Natl. Acad. Sci. USA* 93:10614-10619, 1996 and Heller *et al.*, *Proc. Natl. Acad. Sci. USA* 94:2150-2155, 1997). Alternatively, polynucleotides may be amplified from cDNA prepared from cells expressing the proteins described herein, such as breast tumor cells. Such polynucleotides may be amplified via polymerase
25 chain reaction (PCR). For this approach, sequence-specific primers may be designed based on the sequences provided herein, and may be purchased or synthesized.

An amplified portion of a polynucleotide of the present invention may be used to isolate a full length gene from a suitable library (*e.g.*, a breast tumor cDNA library) using well known techniques. Within such techniques, a library (cDNA or
30 genomic) is screened using one or more polynucleotide probes or primers suitable for

amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

5 For hybridization techniques, a partial sequence may be labeled (*e.g.*, by nick-translation or end-labeling with ^{32}P) using well known techniques. A bacterial or bacteriophage library is then generally screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (*see* Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor
10 Laboratories, Cold Spring Harbor, NY, 1989). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. cDNA clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences may be generated to identify one or more overlapping
15 clones. The complete sequence may then be determined using standard techniques, which may involve generating a series of deletion clones. The resulting overlapping sequences can then assembled into a single contiguous sequence. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

Alternatively, there are numerous amplification techniques for obtaining
20 a full length coding sequence from a partial cDNA sequence. Within such techniques, amplification is generally performed via PCR. Any of a variety of commercially available kits may be used to perform the amplification step. Primers may be designed using, for example, software well known in the art. Primers are preferably 22-30 nucleotides in length, have a GC content of at least 50% and anneal to the target
25 sequence at temperatures of about 68°C to 72°C. The amplified region may be sequenced as described above, and overlapping sequences assembled into a contiguous sequence.

One such amplification technique is inverse PCR (*see* Triglia *et al.*, *Nucl. Acids Res.* 16:8186, 1988), which uses restriction enzymes to generate a fragment
30 in the known region of the gene. The fragment is then circularized by intramolecular ligation and used as a template for PCR with divergent primers derived from the known

region. Within an alternative approach, sequences adjacent to a partial sequence may be retrieved by amplification with a primer to a linker sequence and a primer specific to a known region. The amplified sequences are typically subjected to a second round of amplification with the same linker primer and a second primer specific to the known
5 region. A variation on this procedure, which employs two primers that initiate extension in opposite directions from the known sequence, is described in WO 96/38591. Another such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, which hybridizes to a polyA region or vector sequence, to identify sequences that are 5'
10 and 3' of a known sequence. Additional techniques include capture PCR (Lagerstrom *et al.*, *PCR Methods Applic.* 1:111-19, 1991) and walking PCR (Parker *et al.*, *Nucl. Acids. Res.* 19:3055-60, 1991). Other methods employing amplification may also be employed to obtain a full length cDNA sequence.

In certain instances, it is possible to obtain a full length cDNA sequence
15 by analysis of sequences provided in an expressed sequence tag (EST) database, such as that available from GenBank. Searches for overlapping ESTs may generally be performed using well known programs (*e.g.*, NCBI BLAST searches), and such ESTs may be used to generate a contiguous full length sequence. Full length DNA sequences may also be obtained by analysis of genomic fragments.

20 POLYNUCLEOTIDE EXPRESSION IN HOST CELLS

In other embodiments of the invention, polynucleotide sequences or fragments thereof which encode polypeptides of the invention, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct expression of a polypeptide in appropriate host cells. Due to the inherent degeneracy of
25 the genetic code, other DNA sequences that encode substantially the same or a functionally equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

As will be understood by those of skill in the art, it may be advantageous in some instances to produce polypeptide-encoding nucleotide sequences possessing
30 non-naturally occurring codons. For example, codons preferred by a particular

prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence.

5 Moreover, the polynucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter polypeptide encoding sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the gene product. For example, DNA shuffling by random fragmentation and PCR reassembly of gene
10 fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. In addition, site-directed mutagenesis may be used to insert new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, or introduce mutations, and so forth.

 In another embodiment of the invention, natural, modified, or
15 recombinant nucleic acid sequences may be ligated to a heterologous sequence to encode a fusion protein. For example, to screen peptide libraries for inhibitors of polypeptide activity, it may be useful to encode a chimeric protein that can be recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the polypeptide-encoding
20 sequence and the heterologous protein sequence, so that the polypeptide may be cleaved and purified away from the heterologous moiety.

 Sequences encoding a desired polypeptide may be synthesized, in whole or in part, using chemical methods well known in the art (see Caruthers, M. H. *et al.* (1980) *Nucl. Acids Res. Symp. Ser.* 215-223, Horn, T. *et al.* (1980) *Nucl. Acids Res. Symp. Ser.* 225-232). Alternatively, the protein itself may be produced using chemical
25 methods to synthesize the amino acid sequence of a polypeptide, or a portion thereof. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge, J. Y. *et al.* (1995) *Science* 269:202-204) and automated synthesis may be achieved, for example, using the ABI 431A Peptide Synthesizer (Perkin Elmer, Palo
30 Alto, CA).

A newly synthesized peptide may be substantially purified by preparative high performance liquid chromatography (*e.g.*, Creighton, T. (1983) *Proteins, Structures and Molecular Principles*, WH Freeman and Co., New York, N.Y.) or other comparable techniques available in the art. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (*e.g.*, the Edman degradation procedure). Additionally, the amino acid sequence of a polypeptide, or any part thereof, may be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

10 In order to express a desired polypeptide, the nucleotide sequences encoding the polypeptide, or functional equivalents, may be inserted into appropriate expression vector, *i.e.*, a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled in the art may be used to construct expression vectors containing
15 sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. Such techniques are described in Sambrook, J. *et al.* (1989) *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Plainview, N.Y., and Ausubel, F. M. *et al.* (1989) *Current*
20 *Protocols in Molecular Biology*, John Wiley & Sons, New York, N.Y.

A variety of expression vector/host systems may be utilized to contain and express polynucleotide sequences. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors;
25 insect cell systems infected with virus expression vectors (*e.g.*, baculovirus); plant cell systems transformed with virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (*e.g.*, Ti or pBR322 plasmids); or animal cell systems.

The "control elements" or "regulatory sequences" present in an
30 expression vector are those non-translated regions of the vector--enhancers, promoters, 5' and 3' untranslated regions--which interact with host cellular proteins to carry out

transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid
5 lacZ promoter of the PBLUESCRIPT phagemid (Stratagene, La Jolla, Calif.) or PSORT1 plasmid (Gibco BRL, Gaithersburg, MD) and the like may be used. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are generally preferred. If it is necessary to generate a cell line that contains multiple copies of the sequence encoding a polypeptide, vectors based on SV40 or EBV
10 may be advantageously used with an appropriate selectable marker.

In bacterial systems, a number of expression vectors may be selected depending upon the use intended for the expressed polypeptide. For example, when large quantities are needed, for example for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be used.
15 Such vectors include, but are not limited to, the multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene), in which the sequence encoding the polypeptide of interest may be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of β -galactosidase so that a hybrid protein is produced; pIN vectors (Van Heeke, G. and S. M. Schuster (1989) *J.*
20 *Biol. Chem.* 264:5503-5509); and the like. pGEX Vectors (Promega, Madison, Wis.) may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems may be designed to
25 include heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

In the yeast, *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH may be used. For reviews, see Ausubel *et al.* (supra) and Grant *et al.* (1987) *Methods*
30 *Enzymol.* 153:516-544.

In cases where plant expression vectors are used, the expression of sequences encoding polypeptides may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. 5 (1987) *EMBO J.* 6:307-311. Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. *et al.* (1984) *EMBO J.* 3:1671-1680; Broglie, R. *et al.* (1984) *Science* 224:838-843; and Winter, J. *et al.* (1991) *Results Probl. Cell Differ.* 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques 10 are described in a number of generally available reviews (see, for example, Hobbs, S. or Murry, L. E. in McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York, N.Y.; pp. 191-196).

An insect system may also be used to express a polypeptide of interest. For example, in one such system, *Autographa californica* nuclear polyhedrosis virus 15 (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia* larvae. The sequences encoding the polypeptide may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the polypeptide-encoding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat 20 protein. The recombinant viruses may then be used to infect, for example, *S. frugiperda* cells or *Trichoplusia* larvae in which the polypeptide of interest may be expressed (Engelhard, E. K. *et al.* (1994) *Proc. Natl. Acad. Sci.* 91:3224-3227).

In mammalian host cells, a number of viral-based expression systems are generally available. For example, in cases where an adenovirus is used as an expression 25 vector, sequences encoding a polypeptide of interest may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing the polypeptide in infected host cells (Logan, J. and Shenk, T. (1984) *Proc. Natl. Acad. Sci.* 81:3655-3659). In addition, 30 transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells.

Specific initiation signals may also be used to achieve more efficient translation of sequences encoding a polypeptide of interest. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding the polypeptide, its initiation codon, and upstream sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a portion thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers which are appropriate for the particular cell system which is used, such as those described in the literature (Scharf, D. *et al.* (1994) *Results Probl. Cell Differ.* 20:125-162).

In addition, a host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct insertion, folding and/or function. Different host cells such as CHO, HeLa, MDCK, HEK293, and WI38, which have specific cellular machinery and characteristic mechanisms for such post-translational activities, may be chosen to ensure the correct modification and processing of the foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is generally preferred. For example, cell lines which stably express a polynucleotide of interest may be transformed using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which

successfully express the introduced sequences. Resistant clones of stably transformed cells may be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler, M. *et al.* (1977) *Cell* 11:223-32) and adenine phosphoribosyltransferase (Lowy, I. *et al.* (1990) *Cell* 22:817-23) genes which can be employed in tk.sup.- or apt.sup.- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can be used as the basis for selection; for example, dhfr which confers resistance to methotrexate (Wigler, M. *et al.* (1980) *Proc. Natl. Acad. Sci.* 77:3567-70); npt, which confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin, F. *et al.* (1981) *J. Mol. Biol.* 150:1-14); and als or pat, which confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*). Additional selectable genes have been described, for example, trpB, which allows cells to utilize indole in place of tryptophan, or hisD, which allows cells to utilize histinol in place of histidine (Hartman, S. C. and R. C. Mulligan (1988) *Proc. Natl. Acad. Sci.* 85:8047-51). Recently, the use of visible markers has gained popularity with such markers as anthocyanins, beta-glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C. A. *et al.* (1995) *Methods Mol. Biol.* 55:121-131).

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, its presence and expression may need to be confirmed. For example, if the sequence encoding a polypeptide is inserted within a marker gene sequence, recombinant cells containing sequences can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a polypeptide-encoding sequence under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

Alternatively, host cells which contain and express a desired polynucleotide sequence may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-

RNA hybridizations and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein.

A variety of protocols for detecting and measuring the expression of polynucleotide-encoded products, using either polyclonal or monoclonal antibodies specific for the product are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on a given polypeptide may be preferred for some applications, but a competitive binding assay may also be employed. These and other assays are described, among other places, in Hampton, R. *et al.* (1990; Serological Methods, a Laboratory Manual, APS Press, St Paul, Minn.) and Maddox, D. E. *et al.* (1983; *J. Exp. Med.* 158:1211-1216).

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides include oligolabeling, nick translation, end-labeling or PCR amplification using a labeled nucleotide. Alternatively, the sequences, or any portions thereof may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits. Suitable reporter molecules or labels, which may be used include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with a polynucleotide sequence of interest may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides of the invention may be designed to contain signal sequences which direct secretion of the

encoded polypeptide through a prokaryotic or eukaryotic cell membrane. Other recombinant constructions may be used to join sequences encoding a polypeptide of interest to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Wash.). The inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen, San Diego, Calif.) between the purification domain and the encoded polypeptide may be used to facilitate purification. One such expression vector provides for expression of a fusion protein containing a polypeptide of interest and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography) as described in Porath, J. *et al.* (1992, *Prot. Exp. Purif.* 3:263-281) while the enterokinase cleavage site provides a means for purifying the desired polypeptide from the fusion protein. A discussion of vectors which contain fusion proteins is provided in Kroll, D. J. *et al.* (1993; *DNA Cell Biol.* 12:441-453).

In addition to recombinant production methods, polypeptides of the invention, and fragments thereof, may be produced by direct peptide synthesis using solid-phase techniques (Merrifield J. (1963) *J. Am. Chem. Soc.* 85:2149-2154). Protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Alternatively, various fragments may be chemically synthesized separately and combined using chemical methods to produce the full length molecule.

SITE-SPECIFIC MUTAGENESIS

Site-specific mutagenesis is a technique useful in the preparation of individual peptides, or biologically functional equivalent polypeptides, through specific mutagenesis of the underlying polynucleotides that encode them. The technique,

well-known to those of skill in the art, further provides a ready ability to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the DNA. Site-specific mutagenesis allows the production of mutants through the use of specific
5 oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides of the deletion junction being traversed. Mutations may be employed in a selected polynucleotide sequence to improve, alter, decrease, modify, or otherwise change the
10 properties of the polynucleotide itself, and/or alter the properties, activity, composition, stability, or primary sequence of the encoded polypeptide.

In certain embodiments of the present invention, the inventors contemplate the mutagenesis of the disclosed polynucleotide sequences to alter one or more properties of the encoded polypeptide, such as the antigenicity of a polypeptide
15 vaccine. The techniques of site-specific mutagenesis are well-known in the art, and are widely used to create variants of both polypeptides and polynucleotides. For example, site-specific mutagenesis is often used to alter a specific portion of a DNA molecule. In such embodiments, a primer comprising typically about 14 to about 25 nucleotides or so in length is employed, with about 5 to about 10 residues on both sides of the junction of
20 the sequence being altered.

As will be appreciated by those of skill in the art, site-specific mutagenesis techniques have often employed a phage vector that exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily
25 commercially-available and their use is generally well-known to those skilled in the art. Double-stranded plasmids are also routinely employed in site directed mutagenesis that eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of
30 a double-stranded vector that includes within its sequence a DNA sequence that encodes the desired peptide. An oligonucleotide primer bearing the desired mutated

sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original
5 non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

The preparation of sequence variants of the selected peptide-encoding
10 DNA segments using site-directed mutagenesis provides a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence
15 variants. Specific details regarding these methods and protocols are found in the teachings of Maloy *et al.*, 1994; Segal, 1976; Prokop and Bajpai, 1991; Kuby, 1994; and Maniatis *et al.*, 1982, each incorporated herein by reference, for that purpose.

As used herein, the term "oligonucleotide directed mutagenesis procedure" refers to template-dependent processes and vector-mediated propagation
20 which result in an increase in the concentration of a specific nucleic acid molecule relative to its initial concentration, or in an increase in the concentration of a detectable signal, such as amplification. As used herein, the term "oligonucleotide directed mutagenesis procedure" is intended to refer to a process that involves the template-dependent extension of a primer molecule. The term template dependent
25 process refers to nucleic acid synthesis of an RNA or a DNA molecule wherein the sequence of the newly synthesized strand of nucleic acid is dictated by the well-known rules of complementary base pairing (see, for example, Watson, 1987). Typically, vector mediated methodologies involve the introduction of the nucleic acid fragment into a DNA or RNA vector, the clonal amplification of the vector, and the recovery of
30 the amplified nucleic acid fragment. Examples of such methodologies are provided by U. S. Patent No. 4,237,224, specifically incorporated herein by reference in its entirety.

POLYNUCLEOTIDE AMPLIFICATION TECHNIQUES

A number of template dependent processes are available to amplify the target sequences of interest present in a sample. One of the best known amplification methods is the polymerase chain reaction (PCR™) which is described in detail in U.S. Patent Nos. 4,683,195, 4,683,202 and 4,800,159, each of which is incorporated herein by reference in its entirety. Briefly, in PCR™, two primer sequences are prepared which are complementary to regions on opposite complementary strands of the target sequence. An excess of deoxynucleoside triphosphates is added to a reaction mixture along with a DNA polymerase (*e.g.*, *Taq* polymerase). If the target sequence is present in a sample, the primers will bind to the target and the polymerase will cause the primers to be extended along the target sequence by adding on nucleotides. By raising and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target to form reaction products, excess primers will bind to the target and to the reaction product and the process is repeated. Preferably reverse transcription and PCR™ amplification procedure may be performed in order to quantify the amount of mRNA amplified. Polymerase chain reaction methodologies are well known in the art.

Another method for amplification is the ligase chain reaction (referred to as LCR), disclosed in Eur. Pat. Appl. Publ. No. 320,308 (specifically incorporated herein by reference in its entirety). In LCR, two complementary probe pairs are prepared, and in the presence of the target sequence, each pair will bind to opposite complementary strands of the target such that they abut. In the presence of a ligase, the two probe pairs will link to form a single unit. By temperature cycling, as in PCR™, bound ligated units dissociate from the target and then serve as "target sequences" for ligation of excess probe pairs. U.S. Patent No. 4,883,750, incorporated herein by reference in its entirety, describes an alternative method of amplification similar to LCR for binding probe pairs to a target sequence.

Qbeta Replicase, described in PCT Intl. Pat. Appl. Publ. No. PCT/US87/00880, incorporated herein by reference in its entirety, may also be used as still another amplification method in the present invention. In this method, a replicative sequence of RNA that has a region complementary to that of a target is added to a

sample in the presence of an RNA polymerase. The polymerase will copy the replicative sequence that can then be detected.

An isothermal amplification method, in which restriction endonucleases and ligases are used to achieve the amplification of target molecules that contain
5 nucleotide 5'-[α -thio]triphosphates in one strand of a restriction site (Walker *et al.*, 1992, incorporated herein by reference in its entirety), may also be useful in the amplification of nucleic acids in the present invention.

Strand Displacement Amplification (SDA) is another method of carrying out isothermal amplification of nucleic acids which involves multiple rounds of strand
10 displacement and synthesis, *i.e.* nick translation. A similar method, called Repair Chain Reaction (RCR) is another method of amplification which may be useful in the present invention and is involves annealing several probes throughout a region targeted for amplification, followed by a repair reaction in which only two of the four bases are present. The other two bases can be added as biotinylated derivatives for easy
15 detection. A similar approach is used in SDA.

Sequences can also be detected using a cyclic probe reaction (CPR). In CPR, a probe having a 3' and 5' sequences of non-target DNA and an internal or "middle" sequence of the target protein specific RNA is hybridized to DNA which is present in a sample. Upon hybridization, the reaction is treated with RNaseH, and the
20 products of the probe are identified as distinctive products by generating a signal that is released after digestion. The original template is annealed to another cycling probe and the reaction is repeated. Thus, CPR involves amplifying a signal generated by hybridization of a probe to a target gene specific expressed nucleic acid.

Still other amplification methods described in Great Britain Pat. Appl.
25 No. 2 202 328, and in PCT Intl. Pat. Appl. Publ. No. PCT/US89/01025, each of which is incorporated herein by reference in its entirety, may be used in accordance with the present invention. In the former application, "modified" primers are used in a PCR-like, template and enzyme dependent synthesis. The primers may be modified by labeling with a capture moiety (*e.g.*, biotin) and/or a detector moiety (*e.g.*, enzyme). In
30 the latter application, an excess of labeled probes is added to a sample. In the presence of the target sequence, the probe binds and is cleaved catalytically. After cleavage, the

target sequence is released intact to be bound by excess probe. Cleavage of the labeled probe signals the presence of the target sequence.

Other nucleic acid amplification procedures include transcription-based amplification systems (TAS) (Kwoh *et al.*, 1989; PCT Intl. Pat. Appl. Publ. No. WO 88/10315, incorporated herein by reference in its entirety), including nucleic acid
5 sequence based amplification (NASBA) and 3SR. In NASBA, the nucleic acids can be prepared for amplification by standard phenol/chloroform extraction, heat denaturation of a sample, treatment with lysis buffer and minispin columns for isolation of DNA and RNA or guanidinium chloride extraction of RNA. These amplification techniques
10 involve annealing a primer that has sequences specific to the target sequence. Following polymerization, DNA/RNA hybrids are digested with RNase H while double stranded DNA molecules are heat-denatured again. In either case the single stranded DNA is made fully double stranded by addition of second target-specific primer, followed by polymerization. The double stranded DNA molecules are then multiply
15 transcribed by a polymerase such as T7 or SP6. In an isothermal cyclic reaction, the RNAs are reverse transcribed into DNA, and transcribed once again with a polymerase such as T7 or SP6. The resulting products, whether truncated or complete, indicate target-specific sequences.

Eur. Pat. Appl. Publ. No. 329,822, incorporated herein by reference in its
20 entirety, disclose a nucleic acid amplification process involving cyclically synthesizing single-stranded RNA ("ssRNA"), ssDNA, and double-stranded DNA (dsDNA), which may be used in accordance with the present invention. The ssRNA is a first template for a first primer oligonucleotide, which is elongated by reverse transcriptase (RNA-dependent DNA polymerase). The RNA is then removed from resulting
25 DNA:RNA duplex by the action of ribonuclease H (RNase H, an RNase specific for RNA in a duplex with either DNA or RNA). The resultant ssDNA is a second template for a second primer, which also includes the sequences of an RNA polymerase promoter (exemplified by T7 RNA polymerase) 5' to its homology to its template. This primer is then extended by DNA polymerase (exemplified by the large "Klenow"
30 fragment of *E. coli* DNA polymerase I), resulting as a double-stranded DNA ("dsDNA") molecule, having a sequence identical to that of the original RNA between

the primers and having additionally, at one end, a promoter sequence. This promoter sequence can be used by the appropriate RNA polymerase to make many RNA copies of the DNA. These copies can then re-enter the cycle leading to very swift amplification. With proper choice of enzymes, this amplification can be done
5 isothermally without addition of enzymes at each cycle. Because of the cyclical nature of this process, the starting sequence can be chosen to be in the form of either DNA or RNA.

PCT Intl. Pat. Appl. Publ. No. WO 89/06700, incorporated herein by reference in its entirety, disclose a nucleic acid sequence amplification scheme based on
10 the hybridization of a promoter/primer sequence to a target single-stranded DNA ("ssDNA") followed by transcription of many RNA copies of the sequence. This scheme is not cyclic; *i.e.* new templates are not produced from the resultant RNA transcripts. Other amplification methods include "RACE" (Frohman, 1990), and "one-sided PCR" (Ohara, 1989) which are well-known to those of skill in the art.

15 Methods based on ligation of two (or more) oligonucleotides in the presence of nucleic acid having the sequence of the resulting "di-oligonucleotide", thereby amplifying the di-oligonucleotide (Wu and Dean, 1996, incorporated herein by reference in its entirety), may also be used in the amplification of DNA sequences of the present invention.

20 BIOLOGICAL FUNCTIONAL EQUIVALENTS

Modification and changes may be made in the structure of the polynucleotides and polypeptides of the present invention and still obtain a functional molecule that encodes a polypeptide with desirable characteristics. As mentioned above, it is often desirable to introduce one or more mutations into a specific
25 polynucleotide sequence. In certain circumstances, the resulting encoded polypeptide sequence is altered by this mutation, or in other cases, the sequence of the polypeptide is unchanged by one or more mutations in the encoding polynucleotide.

When it is desirable to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, second-generation molecule, the amino acid

changes may be achieved by changing one or more of the codons of the encoding DNA sequence, according to Table 1.

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated by the inventors that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

TABLE 1

Amino Acids			Codons					
Alanine	Ala	A	GCA	GCC	GCG	GCU		
Cysteine	Cys	C	UGC	UGU				
Aspartic acid	Asp	D	GAC	GAU				
Glutamic acid	Glu	E	GAA	GAG				
Phenylalanine	Phe	F	UUC	UUU				
Glycine	Gly	G	GGA	GGC	GGG	GGU		
Histidine	His	H	CAC	CAU				
Isoleucine	Ile	I	AUA	AUC	AUU			
Lysine	Lys	K	AAA	AAG				
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU
Methionine	Met	M	AUG					
Asparagine	Asn	N	AAC	AAU				
Proline	Pro	P	CCA	CCC	CCG	CCU		
Glutamine	Gln	Q	CAA	CAG				
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU
Threonine	Thr	T	ACA	ACC	ACG	ACU		
Valine	Val	V	GUA	GUC	GUG	GUU		
Tryptophan	Trp	W	UGG					
Tyrosine	Tyr	Y	UAC	UAU				

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics (Kyte and Doolittle, 1982). These values are:

isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5); glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, *i.e.* still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred. It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101 (specifically incorporated herein by reference in its entirety), states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0 \pm 1); glutamate (+3.0 \pm 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5 \pm 1); alanine (-0.5); histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); tryptophan (-3.4). It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those

of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

In addition, any polynucleotide may be further modified to increase stability *in vivo*. Possible modifications include, but are not limited to, the addition of
5 flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queosine and wybutosine, as well as acetyl-methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine and uridine.

10 IN VIVO POLYNUCLEOTIDE DELIVERY TECHNIQUES

In additional embodiments, genetic constructs comprising one or more of the polynucleotides of the invention are introduced into cells *in vivo*. This may be achieved using any of a variety of well known approaches, several of which are outlined below for the purpose of illustration.

15 1. ADENOVIRUS

One of the preferred methods for *in vivo* delivery of one or more nucleic acid sequences involves the use of an adenovirus expression vector. "Adenovirus expression vector" is meant to include those constructs containing adenovirus sequences sufficient to (a) support packaging of the construct and (b) to express a
20 polynucleotide that has been cloned therein in a sense or antisense orientation. Of course, in the context of an antisense construct, expression does not require that the gene product be synthesized.

The expression vector comprises a genetically engineered form of an adenovirus. Knowledge of the genetic organization of adenovirus, a 36 kb, linear,
25 double-stranded DNA virus, allows substitution of large pieces of adenoviral DNA with foreign sequences up to 7 kb (Grunhaus and Horwitz, 1992). In contrast to retrovirus, the adenoviral infection of host cells does not result in chromosomal integration because adenoviral DNA can replicate in an episomal manner without potential genotoxicity. Also, adenoviruses are structurally stable, and no genome rearrangement

has been detected after extensive amplification. Adenovirus can infect virtually all epithelial cells regardless of their cell cycle stage. So far, adenoviral infection appears to be linked only to mild disease such as acute respiratory disease in humans.

Adenovirus is particularly suitable for use as a gene transfer vector because of its mid-sized genome, ease of manipulation, high titer, wide target-cell range and high infectivity. Both ends of the viral genome contain 100-200 base pair inverted repeats (ITRs), which are *cis* elements necessary for viral DNA replication and packaging. The early (E) and late (L) regions of the genome contain different transcription units that are divided by the onset of viral DNA replication. The E1 region (E1A and E1B) encodes proteins responsible for the regulation of transcription of the viral genome and a few cellular genes. The expression of the E2 region (E2A and E2B) results in the synthesis of the proteins for viral DNA replication. These proteins are involved in DNA replication, late gene expression and host cell shut-off (Renan, 1990). The products of the late genes, including the majority of the viral capsid proteins, are expressed only after significant processing of a single primary transcript issued by the major late promoter (MLP). The MLP, (located at 16.8 m.u.) is particularly efficient during the late phase of infection, and all the mRNA's issued from this promoter possess a 5'-tripartite leader (TPL) sequence which makes them preferred mRNA's for translation.

In a current system, recombinant adenovirus is generated from homologous recombination between shuttle vector and provirus vector. Due to the possible recombination between two proviral vectors, wild-type adenovirus may be generated from this process. Therefore, it is critical to isolate a single clone of virus from an individual plaque and examine its genomic structure.

Generation and propagation of the current adenovirus vectors, which are replication deficient, depend on a unique helper cell line, designated 293, which was transformed from human embryonic kidney cells by Ad5 DNA fragments and constitutively expresses E1 proteins (Graham *et al.*, 1977). Since the E3 region is dispensable from the adenovirus genome (Jones and Shenk, 1978), the current adenovirus vectors, with the help of 293 cells, carry foreign DNA in either the E1, the D3 or both regions (Graham and Prevec, 1991). In nature, adenovirus can package

approximately 105% of the wild-type genome (Ghosh-Choudhury *et al.*, 1987), providing capacity for about 2 extra kB of DNA. Combined with the approximately 5.5 kB of DNA that is replaceable in the E1 and E3 regions, the maximum capacity of the current adenovirus vector is under 7.5 kB, or about 15% of the total length of the vector. More than 80% of the adenovirus viral genome remains in the vector backbone and is the source of vector-borne cytotoxicity. Also, the replication deficiency of the E1-deleted virus is incomplete. For example, leakage of viral gene expression has been observed with the currently available vectors at high multiplicities of infection (MOI) (Mulligan, 1993).

10 Helper cell lines may be derived from human cells such as human embryonic kidney cells, muscle cells, hematopoietic cells or other human embryonic mesenchymal or epithelial cells. Alternatively, the helper cells may be derived from the cells of other mammalian species that are permissive for human adenovirus. Such cells include, *e.g.*, Vero cells or other monkey embryonic mesenchymal or epithelial cells.

15 As stated above, the currently preferred helper cell line is 293.

 Recently, Racher *et al.* (1995) disclosed improved methods for culturing 293 cells and propagating adenovirus. In one format, natural cell aggregates are grown by inoculating individual cells into 1 liter siliconized spinner flasks (Techne, Cambridge, UK) containing 100-200 ml of medium. Following stirring at 40 rpm, the cell viability is estimated with trypan blue. In another format, Fibra-Cel microcarriers (Bibby Sterlin, Stone, UK) (5 g/l) is employed as follows. A cell inoculum, resuspended in 5 ml of medium, is added to the carrier (50 ml) in a 250 ml Erlenmeyer flask and left stationary, with occasional agitation, for 1 to 4 h. The medium is then replaced with 50 ml of fresh medium and shaking initiated. For virus production, cells are allowed to grow to about 80% confluence, after which time the medium is replaced (to 25% of the final volume) and adenovirus added at an MOI of 0.05. Cultures are left stationary overnight, following which the volume is increased to 100% and shaking commenced for another 72 h.

 Other than the requirement that the adenovirus vector be replication defective, or at least conditionally defective, the nature of the adenovirus vector is not believed to be crucial to the successful practice of the invention. The adenovirus may

be of any of the 42 different known serotypes or subgroups A-F. Adenovirus type 5 of subgroup C is the preferred starting material in order to obtain a conditional replication-defective adenovirus vector for use in the present invention, since Adenovirus type 5 is a human adenovirus about which a great deal of biochemical and genetic information is known, and it has historically been used for most constructions employing adenovirus as a vector.

As stated above, the typical vector according to the present invention is replication defective and will not have an adenovirus E1 region. Thus, it will be most convenient to introduce the polynucleotide encoding the gene of interest at the position from which the E1-coding sequences have been removed. However, the position of insertion of the construct within the adenovirus sequences is not critical to the invention. The polynucleotide encoding the gene of interest may also be inserted in lieu of the deleted E3 region in E3 replacement vectors as described by Karlsson *et al.* (1986) or in the E4 region where a helper cell line or helper virus complements the E4 defect.

Adenovirus is easy to grow and manipulate and exhibits broad host range *in vitro* and *in vivo*. This group of viruses can be obtained in high titers, *e.g.*, 10^9 - 10^{11} plaque-forming units per ml, and they are highly infective. The life cycle of adenovirus does not require integration into the host cell genome. The foreign genes delivered by adenovirus vectors are episomal and, therefore, have low genotoxicity to host cells. No side effects have been reported in studies of vaccination with wild-type adenovirus (Couch *et al.*, 1963; Top *et al.*, 1971), demonstrating their safety and therapeutic potential as *in vivo* gene transfer vectors.

Adenovirus vectors have been used in eukaryotic gene expression (Levrero *et al.*, 1991; Gomez-Foix *et al.*, 1992) and vaccine development (Grunhaus and Horwitz, 1992; Graham and Prevec, 1992). Recently, animal studies suggested that recombinant adenovirus could be used for gene therapy (Stratford-Perricaudet and Perricaudet, 1991; Stratford-Perricaudet *et al.*, 1990; Rich *et al.*, 1993). Studies in administering recombinant adenovirus to different tissues include trachea instillation (Rosenfeld *et al.*, 1991; Rosenfeld *et al.*, 1992), muscle injection (Ragot *et al.*, 1993),

peripheral intravenous injections (Herz and Gerard, 1993) and stereotactic inoculation into the brain (Le Gal La Salle *et al.*, 1993).

2. RETROVIRUSES

The retroviruses are a group of single-stranded RNA viruses characterized by an ability to convert their RNA to double-stranded DNA in infected cells by a process of reverse-transcription (Coffin, 1990). The resulting DNA then stably integrates into cellular chromosomes as a provirus and directs synthesis of viral proteins. The integration results in the retention of the viral gene sequences in the recipient cell and its descendants. The retroviral genome contains three genes, gag, pol, and env that code for capsid proteins, polymerase enzyme, and envelope components, respectively. A sequence found upstream from the gag gene contains a signal for packaging of the genome into virions. Two long terminal repeat (LTR) sequences are present at the 5' and 3' ends of the viral genome. These contain strong promoter and enhancer sequences and are also required for integration in the host cell genome (Coffin, 1990).

In order to construct a retroviral vector, a nucleic acid encoding one or more oligonucleotide or polynucleotide sequences of interest is inserted into the viral genome in the place of certain viral sequences to produce a virus that is replication-defective. In order to produce virions, a packaging cell line containing the gag, pol, and env genes but without the LTR and packaging components is constructed (Mann *et al.*, 1983). When a recombinant plasmid containing a cDNA, together with the retroviral LTR and packaging sequences is introduced into this cell line (by calcium phosphate precipitation for example), the packaging sequence allows the RNA transcript of the recombinant plasmid to be packaged into viral particles, which are then secreted into the culture media (Nicolas and Rubenstein, 1988; Temin, 1986; Mann *et al.*, 1983). The media containing the recombinant retroviruses is then collected, optionally concentrated, and used for gene transfer. Retroviral vectors are able to infect a broad variety of cell types. However, integration and stable expression require the division of host cells (Paskind *et al.*, 1975).

A novel approach designed to allow specific targeting of retrovirus vectors was recently developed based on the chemical modification of a retrovirus by the chemical addition of lactose residues to the viral envelope. This modification could permit the specific infection of hepatocytes *via* sialoglycoprotein receptors.

5 A different approach to targeting of recombinant retroviruses was designed in which biotinylated antibodies against a retroviral envelope protein and against a specific cell receptor were used. The antibodies were coupled *via* the biotin components by using streptavidin (Roux *et al.*, 1989). Using antibodies against major histocompatibility complex class I and class II antigens, they demonstrated the infection
10 of a variety of human cells that bore those surface antigens with an ecotropic virus *in vitro* (Roux *et al.*, 1989).

3. ADENO-ASSOCIATED VIRUSES

AAV (Ridgeway, 1988; Hermonat and Muzycska, 1984) is a parovirus, discovered as a contamination of adenoviral stocks. It is a ubiquitous virus (antibodies
15 are present in 85% of the US human population) that has not been linked to any disease. It is also classified as a dependovirus, because its replications is dependent on the presence of a helper virus, such as adenovirus. Five serotypes have been isolated, of which AAV-2 is the best characterized. AAV has a single-stranded linear DNA that is encapsidated into capsid proteins VP1, VP2 and VP3 to form an icosahedral virion of
20 20 to 24 nm in diameter (Muzyczka and McLaughlin, 1988).

The AAV DNA is approximately 4.7 kilobases long. It contains two open reading frames and is flanked by two ITRs (FIG. 2). There are two major genes in the AAV genome: *rep* and *cap*. The *rep* gene codes for proteins responsible for viral replications, whereas *cap* codes for capsid protein VP1-3. Each ITR forms a T-shaped
25 hairpin structure. These terminal repeats are the only essential *cis* components of the AAV for chromosomal integration. Therefore, the AAV can be used as a vector with all viral coding sequences removed and replaced by the cassette of genes for delivery. Three viral promoters have been identified and named p5, p19, and p40, according to their map position. Transcription from p5 and p19 results in production of rep proteins,

and transcription from p40 produces the capsid proteins (Hermonat and Muzyczka, 1984).

There are several factors that prompted researchers to study the possibility of using rAAV as an expression vector. One is that the requirements for
5 delivering a gene to integrate into the host chromosome are surprisingly few. It is necessary to have the 145-bp ITRs, which are only 6% of the AAV genome. This leaves room in the vector to assemble a 4.5-kb DNA insertion. While this carrying capacity may prevent the AAV from delivering large genes, it is amply suited for delivering the antisense constructs of the present invention.

10 AAV is also a good choice of delivery vehicles due to its safety. There is a relatively complicated rescue mechanism: not only wild type adenovirus but also AAV genes are required to mobilize rAAV. Likewise, AAV is not pathogenic and not associated with any disease. The removal of viral coding sequences minimizes immune reactions to viral gene expression, and therefore, rAAV does not evoke an inflammatory
15 response.

4. OTHER VIRAL VECTORS AS EXPRESSION CONSTRUCTS

Other viral vectors may be employed as expression constructs in the present invention for the delivery of oligonucleotide or polynucleotide sequences to a host cell. Vectors derived from viruses such as vaccinia virus (Ridgeway, 1988;
20 Coupar *et al.*, 1988), lentiviruses, polio viruses and herpes viruses may be employed. They offer several attractive features for various mammalian cells (Friedmann, 1989; Ridgeway, 1988; Coupar *et al.*, 1988; Horwich *et al.*, 1990).

With the recent recognition of defective hepatitis B viruses, new insight was gained into the structure-function relationship of different viral sequences. *In vitro*
25 studies showed that the virus could retain the ability for helper-dependent packaging and reverse transcription despite the deletion of up to 80% of its genome (Horwich *et al.*, 1990). This suggested that large portions of the genome could be replaced with foreign genetic material. The hepatotropism and persistence (integration) were particularly attractive properties for liver-directed gene transfer. Chang *et al.* (1991)
30 introduced the chloramphenicol acetyltransferase (CAT) gene into duck hepatitis B

virus genome in the place of the polymerase, surface, and pre-surface coding sequences. It was cotransfected with wild-type virus into an avian hepatoma cell line. Culture media containing high titers of the recombinant virus were used to infect primary duckling hepatocytes. Stable CAT gene expression was detected for at least 24 days
5 after transfection (Chang *et al.*, 1991).

5. NON-VIRAL VECTORS

In order to effect expression of the oligonucleotide or polynucleotide sequences of the present invention, the expression construct must be delivered into a cell. This delivery may be accomplished *in vitro*, as in laboratory procedures for
10 transforming cells lines, or *in vivo* or *ex vivo*, as in the treatment of certain disease states. As described above, one preferred mechanism for delivery is *via* viral infection where the expression construct is encapsulated in an infectious viral particle.

Once the expression construct has been delivered into the cell the nucleic acid encoding the desired oligonucleotide or polynucleotide sequences may be
15 positioned and expressed at different sites. In certain embodiments, the nucleic acid encoding the construct may be stably integrated into the genome of the cell. This integration may be in the specific location and orientation *via* homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the nucleic acid may be
20 stably maintained in the cell as a separate, episomal segment of DNA. Such nucleic acid segments or "episomes" encode sequences sufficient to permit maintenance and replication independent of or in synchronization with the host cell cycle. How the expression construct is delivered to a cell and where in the cell the nucleic acid remains is dependent on the type of expression construct employed.

25 In certain embodiments of the invention, the expression construct comprising one or more oligonucleotide or polynucleotide sequences may simply consist of naked recombinant DNA or plasmids. Transfer of the construct may be performed by any of the methods mentioned above which physically or chemically permeabilize the cell membrane. This is particularly applicable for transfer *in vitro* but
30 it may be applied to *in vivo* use as well. Dubensky *et al.* (1984) successfully injected

polyomavirus DNA in the form of calcium phosphate precipitates into liver and spleen of adult and newborn mice demonstrating active viral replication and acute infection. Benvenisty and Reshef (1986) also demonstrated that direct intraperitoneal injection of calcium phosphate-precipitated plasmids results in expression of the transfected genes.

- 5 It is envisioned that DNA encoding a gene of interest may also be transferred in a similar manner *in vivo* and express the gene product.

Another embodiment of the invention for transferring a naked DNA expression construct into cells may involve particle bombardment. This method depends on the ability to accelerate DNA-coated microprojectiles to a high velocity
10 allowing them to pierce cell membranes and enter cells without killing them (Klein *et al.*, 1987). Several devices for accelerating small particles have been developed. One such device relies on a high voltage discharge to generate an electrical current, which in turn provides the motive force (Yang *et al.*, 1990). The microprojectiles used have consisted of biologically inert substances such as tungsten or gold beads.

- 15 Selected organs including the liver, skin, and muscle tissue of rats and mice have been bombarded *in vivo* (Yang *et al.*, 1990; Zelenin *et al.*, 1991). This may require surgical exposure of the tissue or cells, to eliminate any intervening tissue between the gun and the target organ, *i.e.* *ex vivo* treatment. Again, DNA encoding a particular gene may be delivered *via* this method and still be incorporated by the present
20 invention.

ANTISENSE OLIGONUCLEOTIDES

- The end result of the flow of genetic information is the synthesis of protein. DNA is transcribed by polymerases into messenger RNA and translated on the ribosome to yield a folded, functional protein. Thus there are several steps along the
25 route where protein synthesis can be inhibited. The native DNA segment coding for a polypeptide described herein, as all such mammalian DNA strands, has two strands: a sense strand and an antisense strand held together by hydrogen bonding. The messenger RNA coding for polypeptide has the same nucleotide sequence as the sense DNA strand except that the DNA thymidine is replaced by uridine. Thus, synthetic

antisense nucleotide sequences will bind to a mRNA and inhibit expression of the protein encoded by that mRNA.

The targeting of antisense oligonucleotides to mRNA is thus one mechanism to shut down protein synthesis, and, consequently, represents a powerful and targeted therapeutic approach. For example, the synthesis of polygalacturonase and the muscarine type 2 acetylcholine receptor are inhibited by antisense oligonucleotides directed to their respective mRNA sequences (U. S. Patent 5,739,119 and U. S. Patent 5,759,829, each specifically incorporated herein by reference in its entirety). Further, examples of antisense inhibition have been demonstrated with the nuclear protein cyclin, the multiple drug resistance gene (MDG1), ICAM-1, E-selectin, STK-1, striatal GABA_A receptor and human EGF (Jaskulski *et al.*, 1988; Vasanthakumar and Ahmed, 1989; Peris *et al.*, 1998; U. S. Patent 5,801,154; U. S. Patent 5,789,573; U. S. Patent 5,718,709 and U. S. Patent 5,610,288, each specifically incorporated herein by reference in its entirety). Antisense constructs have also been described that inhibit and can be used to treat a variety of abnormal cellular proliferations, *e.g.* cancer (U. S. Patent 5,747,470; U. S. Patent 5,591,317 and U. S. Patent 5,783,683, each specifically incorporated herein by reference in its entirety).

Therefore, in exemplary embodiments, the invention provides oligonucleotide sequences that comprise all, or a portion of, any sequence that is capable of specifically binding to polynucleotide sequence described herein, or a complement thereof. In one embodiment, the antisense oligonucleotides comprise DNA or derivatives thereof. In another embodiment, the oligonucleotides comprise RNA or derivatives thereof. In a third embodiment, the oligonucleotides are modified DNAs comprising a phosphorothioated modified backbone. In a fourth embodiment, the oligonucleotide sequences comprise peptide nucleic acids or derivatives thereof. In each case, preferred compositions comprise a sequence region that is complementary, and more preferably substantially-complementary, and even more preferably, completely complementary to one or more portions of polynucleotides disclosed herein.

Selection of antisense compositions specific for a given gene sequence is based upon analysis of the chosen target sequence (*i.e.* in these illustrative examples the rat and human sequences) and determination of secondary structure, T_m , binding

energy, relative stability, and antisense compositions were selected based upon their relative inability to form dimers, hairpins, or other secondary structures that would reduce or prohibit specific binding to the target mRNA in a host cell.

Highly preferred target regions of the mRNA, are those which are at or
5 near the AUG translation initiation codon, and those sequences which were substantially complementary to 5' regions of the mRNA. These secondary structure analyses and target site selection considerations were performed using v.4 of the OLIGO primer analysis software (Rychlik, 1997) and the BLASTN 2.0.5 algorithm software (Altschul *et al.*, 1997).

10 The use of an antisense delivery method employing a short peptide vector, termed MPG (27 residues), is also contemplated. The MPG peptide contains a hydrophobic domain derived from the fusion sequence of HIV gp41 and a hydrophilic domain from the nuclear localization sequence of SV40 T-antigen (Morris *et al.*, 1997). It has been demonstrated that several molecules of the MPG peptide coat the antisense
15 oligonucleotides and can be delivered into cultured mammalian cells in less than 1 hour with relatively high efficiency (90%). Further, the interaction with MPG strongly increases both the stability of the oligonucleotide to nuclease and the ability to cross the plasma membrane (Morris *et al.*, 1997).

RIBOZYMES

20 Although proteins traditionally have been used for catalysis of nucleic acids, another class of macromolecules has emerged as useful in this endeavor. Ribozymes are RNA-protein complexes that cleave nucleic acids in a site-specific fashion. Ribozymes have specific catalytic domains that possess endonuclease activity (Kim and Cech, 1987; Gerlach *et al.*, 1987; Forster and Symons, 1987). For example, a
25 large number of ribozymes accelerate phosphoester transfer reactions with a high degree of specificity, often cleaving only one of several phosphoesters in an oligonucleotide substrate (Cech *et al.*, 1981; Michel and Westhof, 1990; Reinhold-Hurek and Shub, 1992). This specificity has been attributed to the requirement that the substrate bind via specific base-pairing interactions to the internal guide sequence
30 ("IGS") of the ribozyme prior to chemical reaction.

Ribozyme catalysis has primarily been observed as part of sequence-specific cleavage/ligation reactions involving nucleic acids (Joyce, 1989; Cech *et al.*, 1981). For example, U. S. Patent No. 5,354,855 (specifically incorporated herein by reference) reports that certain ribozymes can act as endonucleases with a sequence
5 specificity greater than that of known ribonucleases and approaching that of the DNA restriction enzymes. Thus, sequence-specific ribozyme-mediated inhibition of gene expression may be particularly suited to therapeutic applications (Scanlon *et al.*, 1991; Sarver *et al.*, 1990). Recently, it was reported that ribozymes elicited genetic changes in some cells lines to which they were applied; the altered genes included the oncogenes
10 H-ras, c-fos and genes of HIV. Most of this work involved the modification of a target mRNA, based on a specific mutant codon that is cleaved by a specific ribozyme.

Six basic varieties of naturally-occurring enzymatic RNAs are known presently. Each can catalyze the hydrolysis of RNA phosphodiester bonds *in trans* (and thus can cleave other RNA molecules) under physiological conditions. In general,
15 enzymatic nucleic acids act by first binding to a target RNA. Such binding occurs through the target binding portion of a enzymatic nucleic acid which is held in close proximity to an enzymatic portion of the molecule that acts to cleave the target RNA. Thus, the enzymatic nucleic acid first recognizes and then binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to
20 cut the target RNA. Strategic cleavage of such a target RNA will destroy its ability to direct synthesis of an encoded protein. After an enzymatic nucleic acid has bound and cleaved its RNA target, it is released from that RNA to search for another target and can repeatedly bind and cleave new targets.

The enzymatic nature of a ribozyme is advantageous over many
25 technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its translation) since the concentration of ribozyme necessary to affect a therapeutic treatment is lower than that of an antisense oligonucleotide. This advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of
30 target RNA. In addition, the ribozyme is a highly specific inhibitor, with the specificity of inhibition depending not only on the base pairing mechanism of binding to the target

RNA, but also on the mechanism of target RNA cleavage. Single mismatches, or base-substitutions, near the site of cleavage can completely eliminate catalytic activity of a ribozyme. Similar mismatches in antisense molecules do not prevent their action (Woolf *et al.*, 1992). Thus, the specificity of action of a ribozyme is greater than that of
5 an antisense oligonucleotide binding the same RNA site.

The enzymatic nucleic acid molecule may be formed in a hammerhead, hairpin, a hepatitis δ virus, group I intron or RNaseP RNA (in association with an RNA guide sequence) or Neurospora VS RNA motif. Examples of hammerhead motifs are described by Rossi *et al.* (1992). Examples of hairpin motifs are described by Hampel
10 *et al.* (Eur. Pat. Appl. Publ. No. EP 0360257), Hampel and Tritz (1989), Hampel *et al.* (1990) and U. S. Patent 5,631,359 (specifically incorporated herein by reference). An example of the hepatitis δ virus motif is described by Perrotta and Been (1992); an example of the RNaseP motif is described by Guerrier-Takada *et al.* (1983); Neurospora VS RNA ribozyme motif is described by Collins (Saville and Collins,
15 1990; Saville and Collins, 1991; Collins and Olive, 1993); and an example of the Group I intron is described in (U. S. Patent 4,987,071, specifically incorporated herein by reference). All that is important in an enzymatic nucleic acid molecule of this invention is that it has a specific substrate binding site which is complementary to one or more of the target gene RNA regions, and that it have nucleotide sequences within or
20 surrounding that substrate binding site which impart an RNA cleaving activity to the molecule. Thus the ribozyme constructs need not be limited to specific motifs mentioned herein.

In certain embodiments, it may be important to produce enzymatic cleaving agents which exhibit a high degree of specificity for the RNA of a desired
25 target, such as one of the sequences disclosed herein. The enzymatic nucleic acid molecule is preferably targeted to a highly conserved sequence region of a target mRNA. Such enzymatic nucleic acid molecules can be delivered exogenously to specific cells as required. Alternatively, the ribozymes can be expressed from DNA or RNA vectors that are delivered to specific cells.

30 Small enzymatic nucleic acid motifs (*e.g.*, of the hammerhead or the hairpin structure) may also be used for exogenous delivery. The simple structure of

these molecules increases the ability of the enzymatic nucleic acid to invade targeted regions of the mRNA structure. Alternatively, catalytic RNA molecules can be expressed within cells from eukaryotic promoters (*e.g.*, Scanlon *et al.*, 1991; Kashani-Sabet *et al.*, 1992; Dropulic *et al.*, 1992; Weerasinghe *et al.*, 1991; Ojwang *et al.*, 1992; 5 Chen *et al.*, 1992; Sarver *et al.*, 1990). Those skilled in the art realize that any ribozyme can be expressed in eukaryotic cells from the appropriate DNA vector. The activity of such ribozymes can be augmented by their release from the primary transcript by a second ribozyme (Int. Pat. Appl. Publ. No. WO 93/23569, and Int. Pat. Appl. Publ. No. WO 94/02595, both hereby incorporated by reference; Ohkawa *et al.*, 10 1992; Taira *et al.*, 1991; and Ventura *et al.*, 1993).

Ribozymes may be added directly, or can be complexed with cationic lipids, lipid complexes, packaged within liposomes, or otherwise delivered to target cells. The RNA or RNA complexes can be locally administered to relevant tissues *ex vivo*, or *in vivo* through injection, aerosol inhalation, infusion pump or stent, with or 15 without their incorporation in biopolymers.

Ribozymes may be designed as described in Int. Pat. Appl. Publ. No. WO 93/23569 and Int. Pat. Appl. Publ. No. WO 94/02595, each specifically incorporated herein by reference) and synthesized to be tested *in vitro* and *in vivo*, as described. Such ribozymes can also be optimized for delivery. While specific 20 examples are provided, those in the art will recognize that equivalent RNA targets in other species can be utilized when necessary.

Hammerhead or hairpin ribozymes may be individually analyzed by computer folding (Jaeger *et al.*, 1989) to assess whether the ribozyme sequences fold into the appropriate secondary structure. Those ribozymes with unfavorable 25 intramolecular interactions between the binding arms and the catalytic core are eliminated from consideration. Varying binding arm lengths can be chosen to optimize activity. Generally, at least 5 or so bases on each arm are able to bind to, or otherwise interact with, the target RNA.

Ribozymes of the hammerhead or hairpin motif may be designed to 30 anneal to various sites in the mRNA message, and can be chemically synthesized. The method of synthesis used follows the procedure for normal RNA synthesis as described

in Usman *et al.* (1987) and in Scaringe *et al.* (1990) and makes use of common nucleic acid protecting and coupling groups, such as dimethoxytrityl at the 5'-end, and phosphoramidites at the 3'-end. Average stepwise coupling yields are typically >98%. Hairpin ribozymes may be synthesized in two parts and annealed to reconstruct an active ribozyme (Chowrira and Burke, 1992). Ribozymes may be modified extensively to enhance stability by modification with nuclease resistant groups, for example, 2'-amino, 2'-C-allyl, 2'-fluoro, 2'-o-methyl, 2'-H (for a review see *e.g.*, Usman and Cedergren, 1992). Ribozymes may be purified by gel electrophoresis using general methods or by high pressure liquid chromatography and resuspended in water.

10 Ribozyme activity can be optimized by altering the length of the ribozyme binding arms, or chemically synthesizing ribozymes with modifications that prevent their degradation by serum ribonucleases (see *e.g.*, Int. Pat. Appl. Publ. No. WO 92/07065; Perrault *et al.*, 1990; Pieken *et al.*, 1991; Usman and Cedergren, 1992; Int. Pat. Appl. Publ. No. WO 93/15187; Int. Pat. Appl. Publ. No. WO 91/03162; Eur. Pat. Appl. Publ. No. 92110298.4; U. S. Patent 5,334,711; and Int. Pat. Appl. Publ. No. WO 94/13688, which describe various chemical modifications that can be made to the sugar moieties of enzymatic RNA molecules), modifications which enhance their efficacy in cells, and removal of stem II bases to shorten RNA synthesis times and reduce chemical requirements.

20 Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) describes the general methods for delivery of enzymatic RNA molecules. Ribozymes may be administered to cells by a variety of methods known to those familiar to the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by incorporation into other vehicles, such as hydrogels, cyclodextrins, biodegradable nanocapsules, and bioadhesive microspheres. For some indications, ribozymes may be directly delivered *ex vivo* to cells or tissues with or without the aforementioned vehicles. Alternatively, the RNA/vehicle combination may be locally delivered by direct inhalation, by direct injection or by use of a catheter, infusion pump or stent. Other routes of delivery include, but are not limited to, intravascular, intramuscular, 25 subcutaneous or joint injection, aerosol inhalation, oral (tablet or pill form), topical, systemic, ocular, intraperitoneal and/or intrathecal delivery. More detailed descriptions

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of ribozyme delivery and administration are provided in Int. Pat. Appl. Publ. No. WO 94/02595 and Int. Pat. Appl. Publ. No. WO 93/23569, each specifically incorporated herein by reference.

Another means of accumulating high concentrations of a ribozyme(s) within cells is to incorporate the ribozyme-encoding sequences into a DNA expression vector. Transcription of the ribozyme sequences are driven from a promoter for eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters will be expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type will depend on the nature of the gene regulatory sequences (enhancers, silencers, *etc.*) present nearby. Prokaryotic RNA polymerase promoters may also be used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells (Elroy-Stein and Moss, 1990; Gao and Huang, 1993; Lieber *et al.*, 1993; Zhou *et al.*, 1990). Ribozymes expressed from such promoters can function in mammalian cells (*e.g.* Kashani-Saber *et al.*, 1992; Ojwang *et al.*, 1992; Chen *et al.*, 1992; Yu *et al.*, 1993; L'Huillier *et al.*, 1992; Lisiewicz *et al.*, 1993). Such transcription units can be incorporated into a variety of vectors for introduction into mammalian cells, including but not restricted to, plasmid DNA vectors, viral DNA vectors (such as adenovirus or adeno-associated vectors), or viral RNA vectors (such as retroviral, semliki forest virus, sindbis virus vectors).

Ribozymes may be used as diagnostic tools to examine genetic drift and mutations within diseased cells. They can also be used to assess levels of the target RNA molecule. The close relationship between ribozyme activity and the structure of the target RNA allows the detection of mutations in any region of the molecule which alters the base-pairing and three-dimensional structure of the target RNA. By using multiple ribozymes, one may map nucleotide changes which are important to RNA structure and function *in vitro*, as well as in cells and tissues. Cleavage of target RNAs with ribozymes may be used to inhibit gene expression and define the role (essentially) of specified gene products in the progression of disease. In this manner, other genetic targets may be defined as important mediators of the disease. These studies will lead to better treatment of the disease progression by affording the possibility of combinational

therapies (e.g., multiple ribozymes targeted to different genes, ribozymes coupled with known small molecule inhibitors, or intermittent treatment with combinations of ribozymes and/or other chemical or biological molecules). Other *in vitro* uses of ribozymes are well known in the art, and include detection of the presence of mRNA
5 associated with an IL-5 related condition. Such RNA is detected by determining the presence of a cleavage product after treatment with a ribozyme using standard methodology.

PEPTIDE NUCLEIC ACIDS

In certain embodiments, the inventors contemplate the use of peptide
10 nucleic acids (PNAs) in the practice of the methods of the invention. PNA is a DNA mimic in which the nucleobases are attached to a pseudopeptide backbone (Good and Nielsen, 1997). PNA is able to be utilized in a number of methods that traditionally have used RNA or DNA. Often PNA sequences perform better in techniques than the corresponding RNA or DNA sequences and have utilities that are not inherent to RNA
15 or DNA. A review of PNA including methods of making, characteristics of, and methods of using, is provided by Corey (1997) and is incorporated herein by reference. As such, in certain embodiments, one may prepare PNA sequences that are complementary to one or more portions of the ACE mRNA sequence, and such PNA compositions may be used to regulate, alter, decrease, or reduce the translation of ACE-
20 specific mRNA, and thereby alter the level of ACE activity in a host cell to which such PNA compositions have been administered.

PNAs have 2-aminoethyl-glycine linkages replacing the normal phosphodiester backbone of DNA (Nielsen *et al.*, 1991; Hanvey *et al.*, 1992; Hyrup and Nielsen, 1996; Nielsen, 1996). This chemistry has three important consequences:
25 firstly, in contrast to DNA or phosphorothioate oligonucleotides, PNAs are neutral molecules; secondly, PNAs are achiral, which avoids the need to develop a stereoselective synthesis; and thirdly, PNA synthesis uses standard Boc (Dueholm *et al.*, 1994) or Fmoc (Thomson *et al.*, 1995) protocols for solid-phase peptide synthesis, although other methods, including a modified Merrifield method, have been used
30 (Christensen *et al.*, 1995).

PNA monomers or ready-made oligomers are commercially available from PerSeptive Biosystems (Framingham, MA). PNA syntheses by either Boc or Fmoc protocols are straightforward using manual or automated protocols (Norton *et al.*, 1995). The manual protocol lends itself to the production of chemically modified PNAs or the simultaneous synthesis of families of closely related PNAs.

As with peptide synthesis, the success of a particular PNA synthesis will depend on the properties of the chosen sequence. For example, while in theory PNAs can incorporate any combination of nucleotide bases, the presence of adjacent purines can lead to deletions of one or more residues in the product. In expectation of this difficulty, it is suggested that, in producing PNAs with adjacent purines, one should repeat the coupling of residues likely to be added inefficiently. This should be followed by the purification of PNAs by reverse-phase high-pressure liquid chromatography (Norton *et al.*, 1995) providing yields and purity of product similar to those observed during the synthesis of peptides.

Modifications of PNAs for a given application may be accomplished by coupling amino acids during solid-phase synthesis or by attaching compounds that contain a carboxylic acid group to the exposed N-terminal amine. Alternatively, PNAs can be modified after synthesis by coupling to an introduced lysine or cysteine. The ease with which PNAs can be modified facilitates optimization for better solubility or for specific functional requirements. Once synthesized, the identity of PNAs and their derivatives can be confirmed by mass spectrometry. Several studies have made and utilized modifications of PNAs (Norton *et al.*, 1995; Haaima *et al.*, 1996; Stetsenko *et al.*, 1996; Petersen *et al.*, 1995; Ulmann *et al.*, 1996; Koch *et al.*, 1995; Orum *et al.*, 1995; Footer *et al.*, 1996; Griffith *et al.*, 1995; Kremsky *et al.*, 1996; Pardridge *et al.*, 1995; Boffa *et al.*, 1995; Landsdorp *et al.*, 1996; Gambacorti-Passerini *et al.*, 1996; Armitage *et al.*, 1997; Seeger *et al.*, 1997; Ruskowski *et al.*, 1997). U.S. Patent No. 5,700,922 discusses PNA-DNA-PNA chimeric molecules and their uses in diagnostics, modulating protein in organisms, and treatment of conditions susceptible to therapeutics.

In contrast to DNA and RNA, which contain negatively charged linkages, the PNA backbone is neutral. In spite of this dramatic alteration, PNAs

recognize complementary DNA and RNA by Watson-Crick pairing (Egholm *et al.*, 1993), validating the initial modeling by Nielsen *et al.* (1991). PNAs lack 3' to 5' polarity and can bind in either parallel or antiparallel fashion, with the antiparallel mode being preferred (Egholm *et al.*, 1993).

5 Hybridization of DNA oligonucleotides to DNA and RNA is destabilized by electrostatic repulsion between the negatively charged phosphate backbones of the complementary strands. By contrast, the absence of charge repulsion in PNA-DNA or PNA-RNA duplexes increases the melting temperature (T_m) and reduces the dependence of T_m on the concentration of mono- or divalent cations
10 (Nielsen *et al.*, 1991). The enhanced rate and affinity of hybridization are significant because they are responsible for the surprising ability of PNAs to perform strand invasion of complementary sequences within relaxed double-stranded DNA. In addition, the efficient hybridization at inverted repeats suggests that PNAs can recognize secondary structure effectively within double-stranded DNA. Enhanced
15 recognition also occurs with PNAs immobilized on surfaces, and Wang *et al.* have shown that support-bound PNAs can be used to detect hybridization events (Wang *et al.*, 1996).

One might expect that tight binding of PNAs to complementary sequences would also increase binding to similar (but not identical) sequences, reducing
20 the sequence specificity of PNA recognition. As with DNA hybridization, however, selective recognition can be achieved by balancing oligomer length and incubation temperature. Moreover, selective hybridization of PNAs is encouraged by PNA-DNA hybridization being less tolerant of base mismatches than DNA-DNA hybridization. For example, a single mismatch within a 16 bp PNA-DNA duplex can reduce the T_m by
25 up to 15°C (Egholm *et al.*, 1993). This high level of discrimination has allowed the development of several PNA-based strategies for the analysis of point mutations (Wang *et al.*, 1996; Carlsson *et al.*, 1996; Thiede *et al.*, 1996; Webb and Hurskainen, 1996; Perry-O'Keefe *et al.*, 1996).

High-affinity binding provides clear advantages for molecular
30 recognition and the development of new applications for PNAs. For example, 11-13 nucleotide PNAs inhibit the activity of telomerase, a ribonucleo-protein that extends

telomere ends using an essential RNA template, while the analogous DNA oligomers do not (Norton *et al.*, 1996).

Neutral PNAs are more hydrophobic than analogous DNA oligomers, and this can lead to difficulty solubilizing them at neutral pH, especially if the PNAs
5 have a high purine content or if they have the potential to form secondary structures. Their solubility can be enhanced by attaching one or more positive charges to the PNA termini (Nielsen *et al.*, 1991).

Findings by Allfrey and colleagues suggest that strand invasion will occur spontaneously at sequences within chromosomal DNA (Boffa *et al.*, 1995; Boffa
10 *et al.*, 1996). These studies targeted PNAs to triplet repeats of the nucleotides CAG and used this recognition to purify transcriptionally active DNA (Boffa *et al.*, 1995) and to inhibit transcription (Boffa *et al.*, 1996). This result suggests that if PNAs can be delivered within cells then they will have the potential to be general sequence-specific regulators of gene expression. Studies and reviews concerning the use of PNAs as
15 antisense and anti-gene agents include Nielsen *et al.* (1993b), Hanvey *et al.* (1992), and Good and Nielsen (1997). Koppelhus *et al.* (1997) have used PNAs to inhibit HIV-1 inverse transcription, showing that PNAs may be used for antiviral therapies.

Methods of characterizing the antisense binding properties of PNAs are discussed in Rose (1993) and Jensen *et al.* (1997). Rose uses capillary gel
20 electrophoresis to determine binding of PNAs to their complementary oligonucleotide, measuring the relative binding kinetics and stoichiometry. Similar types of measurements were made by Jensen *et al.* using BIAcore™ technology.

Other applications of PNAs include use in DNA strand invasion (Nielsen *et al.*, 1991), antisense inhibition (Hanvey *et al.*, 1992), mutational analysis (Orum *et al.*,
25 *et al.*, 1993), enhancers of transcription (Mollegaard *et al.*, 1994), nucleic acid purification (Orum *et al.*, 1995), isolation of transcriptionally active genes (Boffa *et al.*, 1995), blocking of transcription factor binding (Vickers *et al.*, 1995), genome cleavage (Veselkov *et al.*, 1996), biosensors (Wang *et al.*, 1996), *in situ* hybridization (Thisted *et al.*, 1996), and in a alternative to Southern blotting (Perry-O'Keefe, 1996).

POLYPEPTIDE COMPOSITIONS

The present invention, in other aspects, provides polypeptide compositions. Generally, a polypeptide of the invention will be an isolated polypeptide (or an epitope, variant, or active fragment thereof) derived from a mammalian species.

5 Preferably, the polypeptide is encoded by a polynucleotide sequence disclosed herein or a sequence which hybridizes under moderately stringent conditions to a polynucleotide sequence disclosed herein. Alternatively, the polypeptide may be defined as a polypeptide which comprises a contiguous amino acid sequence from an amino acid sequence disclosed herein, or which polypeptide comprises an entire amino acid

10 sequence disclosed herein.

In the present invention, a polypeptide composition is also understood to comprise one or more polypeptides that are immunologically reactive with antibodies generated against a polypeptide of the invention, particularly a polypeptide having the amino acid sequence disclosed in SEQ ID NO:126, or to active fragments, or to variants

15 or biological functional equivalents thereof.

Likewise, a polypeptide composition of the present invention is understood to comprise one or more polypeptides that are capable of eliciting antibodies that are immunologically reactive with one or more polypeptides encoded by one or more contiguous nucleic acid sequences contained in SEQ ID NO:1-125, or to active

20 fragments, or to variants thereof, or to one or more nucleic acid sequences which hybridize to one or more of these sequences under conditions of moderate to high stringency. Particularly illustrative polypeptides include the amino acid sequence disclosed in SEQ ID NO:126.

As used herein, an active fragment of a polypeptide includes a whole or

25 a portion of a polypeptide which is modified by conventional techniques, *e.g.*, mutagenesis, or by addition, deletion, or substitution, but which active fragment exhibits substantially the same structure function, antigenicity, etc., as a polypeptide as described herein.

In certain illustrative embodiments, the polypeptides of the invention

30 will comprise at least an immunogenic portion of a breast tumor protein or a variant thereof, as described herein. As noted above, a "breast tumor protein" is a protein that

is expressed by breast tumor cells. Proteins that are breast tumor proteins also react detectably within an immunoassay (such as an ELISA) with antisera from a patient with breast cancer. Polypeptides as described herein may be of any length. Additional sequences derived from the native protein and/or heterologous sequences may be present, and such sequences may (but need not) possess further immunogenic or antigenic properties.

An "immunogenic portion," as used herein is a portion of a protein that is recognized (*i.e.*, specifically bound) by a B-cell and/or T-cell surface antigen receptor. Such immunogenic portions generally comprise at least 5 amino acid residues, more preferably at least 10, and still more preferably at least 20 amino acid residues of a breast tumor protein or a variant thereof. Certain preferred immunogenic portions include peptides in which an N-terminal leader sequence and/or transmembrane domain have been deleted. Other preferred immunogenic portions may contain a small N- and/or C-terminal deletion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids), relative to the mature protein.

Immunogenic portions may generally be identified using well known techniques, such as those summarized in Paul, *Fundamental Immunology*, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides for the ability to react with antigen-specific antibodies, antisera and/or T-cell lines or clones. As used herein, antisera and antibodies are "antigen-specific" if they specifically bind to an antigen (*i.e.*, they react with the protein in an ELISA or other immunoassay, and do not react detectably with unrelated proteins). Such antisera and antibodies may be prepared as described herein, and using well known techniques. An immunogenic portion of a native breast tumor protein is a portion that reacts with such antisera and/or T-cells at a level that is not substantially less than the reactivity of the full length polypeptide (*e.g.*, in an ELISA and/or T-cell reactivity assay). Such immunogenic portions may react within such assays at a level that is similar to or greater than the reactivity of the full length polypeptide. Such screens may generally be performed using methods well known to those of ordinary skill in the art, such as those described in Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. For example, a polypeptide may be

immobilized on a solid support and contacted with patient sera to allow binding of antibodies within the sera to the immobilized polypeptide. Unbound sera may then be removed and bound antibodies detected using, for example, ¹²⁵I-labeled Protein A.

As noted above, a composition may comprise a variant of a native breast
5 tumor protein. A polypeptide "variant," as used herein, is a polypeptide that differs from a native breast tumor protein in one or more substitutions, deletions, additions and/or insertions, such that the immunogenicity of the polypeptide is not substantially diminished. In other words, the ability of a variant to react with antigen-specific antisera may be enhanced or unchanged, relative to the native protein, or may be
10 diminished by less than 50%, and preferably less than 20%, relative to the native protein. Such variants may generally be identified by modifying one of the above polypeptide sequences and evaluating the reactivity of the modified polypeptide with antigen-specific antibodies or antisera as described herein. Preferred variants include those in which one or more portions, such as an N-terminal leader sequence or
15 transmembrane domain, have been removed. Other preferred variants include variants in which a small portion (e.g., 1-30 amino acids, preferably 5-15 amino acids) has been removed from the N- and/or C-terminal of the mature protein.

Polypeptide variants encompassed by the present invention include those exhibiting at least about 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%,
20 97%, 98%, or 99% or more identity (determined as described above) to the polypeptides disclosed herein.

Preferably, a variant contains conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide
25 chemistry would expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged. Amino acid substitutions may generally be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino
30 acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and

alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine. Other groups of amino acids that may represent conservative changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his. A variant may also, or alternatively, contain
5 nonconservative changes. In a preferred embodiment, variant polypeptides differ from a native sequence by substitution, deletion or addition of five amino acids or fewer. Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenicity, secondary structure and hydrophobic nature of the polypeptide.

10 As noted above, polypeptides may comprise a signal (or leader) sequence at the N-terminal end of the protein, which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the polypeptide (e.g., poly-His), or to enhance binding of the polypeptide to a solid support.
15 For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

Polypeptides may be prepared using any of a variety of well known techniques. Recombinant polypeptides encoded by DNA sequences as described above may be readily prepared from the DNA sequences using any of a variety of expression vectors known to those of ordinary skill in the art. Expression may be achieved in any
20 appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast, and higher eukaryotic cells, such as mammalian cells and plant cells. Preferably, the host cells employed are *E. coli*, yeast or a mammalian cell line such as COS or CHO. Supernatants from suitable host/vector systems which
25 secrete recombinant protein or polypeptide into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion exchange resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant polypeptide.

30 Portions and other variants having less than about 100 amino acids, and generally less than about 50 amino acids, may also be generated by synthetic means,

using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. See Merrifield, *J. Am. Chem. Soc.* 5 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, CA), and may be operated according to the manufacturer's instructions.

Within certain specific embodiments, a polypeptide may be a fusion protein that comprises multiple polypeptides as described herein, or that comprises at 10 least one polypeptide as described herein and an unrelated sequence, such as a known tumor protein. A fusion partner may, for example, assist in providing T helper epitopes (an immunological fusion partner), preferably T helper epitopes recognized by humans, or may assist in expressing the protein (an expression enhancer) at higher yields than 15 the native recombinant protein. Certain preferred fusion partners are both immunological and expression enhancing fusion partners. Other fusion partners may be selected so as to increase the solubility of the protein or to enable the protein to be targeted to desired intracellular compartments. Still further fusion partners include affinity tags, which facilitate purification of the protein.

20 Fusion proteins may generally be prepared using standard techniques, including chemical conjugation. Preferably, a fusion protein is expressed as a recombinant protein, allowing the production of increased levels, relative to a non-fused protein, in an expression system. Briefly, DNA sequences encoding the polypeptide components may be assembled separately, and ligated into an appropriate expression 25 vector. The 3' end of the DNA sequence encoding one polypeptide component is ligated, with or without a peptide linker, to the 5' end of a DNA sequence encoding the second polypeptide component so that the reading frames of the sequences are in phase. This permits translation into a single fusion protein that retains the biological activity of both component polypeptides.

30 A peptide linker sequence may be employed to separate the first and second polypeptide components by a distance sufficient to ensure that each polypeptide

5 folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion protein using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes. Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other near neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be usefully employed as linkers include those disclosed in Maratea *et al.*, *Gene* 40:39-46, 1985; Murphy *et al.*, *Proc. Natl. Acad. Sci. USA* 83:8258-8262, 1986; U.S. Patent No. 4,935,233 and U.S. Patent No. 4,751,180. The linker sequence may generally be from 1 to about 50 amino acids in length. Linker sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the functional domains and prevent steric interference.

15 The ligated DNA sequences are operably linked to suitable transcriptional or translational regulatory elements. The regulatory elements responsible for expression of DNA are located only 5' to the DNA sequence encoding the first polypeptides. Similarly, stop codons required to end translation and transcription termination signals are only present 3' to the DNA sequence encoding the second polypeptide.

25 Fusion proteins are also provided. Such proteins comprise a polypeptide as described herein together with an unrelated immunogenic protein. Preferably the immunogenic protein is capable of eliciting a recall response. Examples of such proteins include tetanus, tuberculosis and hepatitis proteins (*see*, for example, Stoute *et al. New Engl. J. Med.*, 336:86-91, 1997).

30 Within preferred embodiments, an immunological fusion partner is derived from protein D, a surface protein of the gram-negative bacterium *Haemophilus influenza B* (WO 91/18926). Preferably, a protein D derivative comprises approximately the first third of the protein (*e.g.*, the first N-terminal 100-110 amino acids), and a protein D derivative may be lipidated. Within certain preferred

embodiments, the first 109 residues of a Lipoprotein D fusion partner is included on the N-terminus to provide the polypeptide with additional exogenous T-cell epitopes and to increase the expression level in *E. coli* (thus functioning as an expression enhancer). The lipid tail ensures optimal presentation of the antigen to antigen presenting cells.

5 Other fusion partners include the non-structural protein from influenzae virus, NS1 (hemagglutinin). Typically, the N-terminal 81 amino acids are used, although different fragments that include T-helper epitopes may be used.

In another embodiment, the immunological fusion partner is the protein known as LYTA, or a portion thereof (preferably a C-terminal portion). LYTA is

10 derived from *Streptococcus pneumoniae*, which synthesizes an N-acetyl-L-alanine amidase known as amidase LYTA (encoded by the *LytA* gene; *Gene* 43:265-292, 1986). LYTA is an autolysin that specifically degrades certain bonds in the peptidoglycan backbone. The C-terminal domain of the LYTA protein is responsible for the affinity to the choline or to some choline analogues such as DEAE. This

15 property has been exploited for the development of *E. coli* C-LYTA expressing plasmids useful for expression of fusion proteins. Purification of hybrid proteins containing the C-LYTA fragment at the amino terminus has been described (see *Biotechnology* 10:795-798, 1992). Within a preferred embodiment, a repeat portion of LYTA may be incorporated into a fusion protein. A repeat portion is found in the C-

20 terminal region starting at residue 178. A particularly preferred repeat portion incorporates residues 188-305.

In general, polypeptides (including fusion proteins) and polynucleotides as described herein are isolated. An "isolated" polypeptide or polynucleotide is one that is removed from its original environment. For example, a naturally-occurring protein is

25 isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably, such polypeptides are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure. A polynucleotide is considered to be isolated if, for example, it is cloned into a vector that is not a part of the natural environment.

BINDING AGENTS

The present invention further provides agents, such as antibodies and antigen-binding fragments thereof, that specifically bind to a breast tumor protein. As used herein, an antibody, or antigen-binding fragment thereof, is said to "specifically bind" to a breast tumor protein if it reacts at a detectable level (within, for example, an ELISA) with a breast tumor protein, and does not react detectably with unrelated proteins under similar conditions. As used herein, "binding" refers to a noncovalent association between two separate molecules such that a complex is formed. The ability to bind may be evaluated by, for example, determining a binding constant for the formation of the complex. The binding constant is the value obtained when the concentration of the complex is divided by the product of the component concentrations. In general, two compounds are said to "bind," in the context of the present invention, when the binding constant for complex formation exceeds about 10^3 L/mol. The binding constant may be determined using methods well known in the art.

Binding agents may be further capable of differentiating between patients with and without a cancer, such as breast cancer, using the representative assays provided herein. In other words, antibodies or other binding agents that bind to a breast tumor protein will generate a signal indicating the presence of a cancer in at least about 20% of patients with the disease, and will generate a negative signal indicating the absence of the disease in at least about 90% of individuals without the cancer. To determine whether a binding agent satisfies this requirement, biological samples (*e.g.*, blood, sera, sputum, urine and/or tumor biopsies) from patients with and without a cancer (as determined using standard clinical tests) may be assayed as described herein for the presence of polypeptides that bind to the binding agent. It will be apparent that a statistically significant number of samples with and without the disease should be assayed. Each binding agent should satisfy the above criteria; however, those of ordinary skill in the art will recognize that binding agents may be used in combination to improve sensitivity.

Any agent that satisfies the above requirements may be a binding agent. For example, a binding agent may be a ribosome, with or without a peptide component, an RNA molecule or a polypeptide. In a preferred embodiment, a binding agent is an

antibody or an antigen-binding fragment thereof. Antibodies may be prepared by any of a variety of techniques known to those of ordinary skill in the art. *See, e.g.,* Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, antibodies can be produced by cell culture techniques, including the generation
5 of monoclonal antibodies as described herein, or via transfection of antibody genes into suitable bacterial or mammalian cell hosts, in order to allow for the production of recombinant antibodies. In one technique, an immunogen comprising the polypeptide is initially injected into any of a wide variety of mammals (*e.g.,* mice, rats, rabbits, sheep or goats). In this step, the polypeptides of this invention may serve as the
10 immunogen without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the animal host, preferably according to a predetermined schedule incorporating one or more booster immunizations, and the animals are bled
15 periodically. Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

Monoclonal antibodies specific for an antigenic polypeptide of interest may be prepared, for example, using the technique of Kohler and Milstein, *Eur. J.*
20 *Immunol.* 6:511-519, 1976, and improvements thereto. Briefly, these methods involve the preparation of immortal cell lines capable of producing antibodies having the desired specificity (*i.e.,* reactivity with the polypeptide of interest). Such cell lines may be produced, for example, from spleen cells obtained from an animal immunized as described above. The spleen cells are then immortalized by, for example, fusion with a
25 myeloma cell fusion partner, preferably one that is syngeneic with the immunized animal. A variety of fusion techniques may be employed. For example, the spleen cells and myeloma cells may be combined with a nonionic detergent for a few minutes and then plated at low density on a selective medium that supports the growth of hybrid cells, but not myeloma cells. A preferred selection technique uses HAT (hypoxanthine,
30 aminopterin, thymidine) selection. After a sufficient time, usually about 1 to 2 weeks, colonies of hybrids are observed. Single colonies are selected and their culture

supernatants tested for binding activity against the polypeptide. Hybridomas having high reactivity and specificity are preferred.

Monoclonal antibodies may be isolated from the supernatants of growing hybridoma colonies. In addition, various techniques may be employed to enhance the yield, such as injection of the hybridoma cell line into the peritoneal cavity of a suitable vertebrate host, such as a mouse. Monoclonal antibodies may then be harvested from the ascites fluid or the blood. Contaminants may be removed from the antibodies by conventional techniques, such as chromatography, gel filtration, precipitation, and extraction. The polypeptides of this invention may be used in the purification process in, for example, an affinity chromatography step.

Within certain embodiments, the use of antigen-binding fragments of antibodies may be preferred. Such fragments include Fab fragments, which may be prepared using standard techniques. Briefly, immunoglobulins may be purified from rabbit serum by affinity chromatography on Protein A bead columns (Harlow and Lane, 15 *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988) and digested by papain to yield Fab and Fc fragments. The Fab and Fc fragments may be separated by affinity chromatography on protein A bead columns.

Monoclonal antibodies of the present invention may be coupled to one or more therapeutic agents. Suitable agents in this regard include radionuclides, differentiation inducers, drugs, toxins, and derivatives thereof. Preferred radionuclides include ^{90}Y , ^{123}I , ^{125}I , ^{131}I , ^{186}Re , ^{188}Re , ^{211}At , and ^{212}Bi . Preferred drugs include methotrexate, and pyrimidine and purine analogs. Preferred differentiation inducers include phorbol esters and butyric acid. Preferred toxins include ricin, abrin, diphtheria toxin, cholera toxin, gelonin, *Pseudomonas* exotoxin, *Shigella* toxin, and pokeweed 25 antiviral protein.

A therapeutic agent may be coupled (e.g., covalently bonded) to a suitable monoclonal antibody either directly or indirectly (e.g., via a linker group). A direct reaction between an agent and an antibody is possible when each possesses a substituent capable of reacting with the other. For example, a nucleophilic group, such as an amino or sulfhydryl group, on one may be capable of reacting with a carbonyl- 30

containing group, such as an anhydride or an acid halide, or with an alkyl group containing a good leaving group (*e.g.*, a halide) on the other.

Alternatively, it may be desirable to couple a therapeutic agent and an antibody via a linker group. A linker group can function as a spacer to distance an
5 antibody from an agent in order to avoid interference with binding capabilities. A linker group can also serve to increase the chemical reactivity of a substituent on an agent or an antibody, and thus increase the coupling efficiency. An increase in chemical reactivity may also facilitate the use of agents, or functional groups on agents, which otherwise would not be possible.

10 It will be evident to those skilled in the art that a variety of bifunctional or polyfunctional reagents, both homo- and hetero-functional (such as those described in the catalog of the Pierce Chemical Co., Rockford, IL), may be employed as the linker group. Coupling may be effected, for example, through amino groups, carboxyl groups, sulfhydryl groups or oxidized carbohydrate residues. There are numerous references
15 describing such methodology, *e.g.*, U.S. Patent No. 4,671,958, to Rodwell *et al.*

Where a therapeutic agent is more potent when free from the antibody portion of the immunoconjugates of the present invention, it may be desirable to use a linker group which is cleavable during or upon internalization into a cell. A number of different cleavable linker groups have been described. The mechanisms for the
20 intracellular release of an agent from these linker groups include cleavage by reduction of a disulfide bond (*e.g.*, U.S. Patent No. 4,489,710, to Spitler), by irradiation of a photolabile bond (*e.g.*, U.S. Patent No. 4,625,014, to Senter *et al.*), by hydrolysis of derivatized amino acid side chains (*e.g.*, U.S. Patent No. 4,638,045, to Kohn *et al.*), by serum complement-mediated hydrolysis (*e.g.*, U.S. Patent No. 4,671,958, to Rodwell *et al.*), and acid-catalyzed hydrolysis (*e.g.*, U.S. Patent No. 4,569,789, to Blattler *et al.*).
25

It may be desirable to couple more than one agent to an antibody. In one embodiment, multiple molecules of an agent are coupled to one antibody molecule. In another embodiment, more than one type of agent may be coupled to one antibody. Regardless of the particular embodiment, immunoconjugates with more than one agent
30 may be prepared in a variety of ways. For example, more than one agent may be

coupled directly to an antibody molecule, or linkers that provide multiple sites for attachment can be used. Alternatively, a carrier can be used.

A carrier may bear the agents in a variety of ways, including covalent bonding either directly or via a linker group. Suitable carriers include proteins such as albumins (*e.g.*, U.S. Patent No. 4,507,234, to Kato *et al.*), peptides and polysaccharides such as aminodextran (*e.g.*, U.S. Patent No. 4,699,784, to Shih *et al.*). A carrier may also bear an agent by noncovalent bonding or by encapsulation, such as within a liposome vesicle (*e.g.*, U.S. Patent Nos. 4,429,008 and 4,873,088). Carriers specific for radionuclide agents include radiohalogenated small molecules and chelating compounds. For example, U.S. Patent No. 4,735,792 discloses representative radiohalogenated small molecules and their synthesis. A radionuclide chelate may be formed from chelating compounds that include those containing nitrogen and sulfur atoms as the donor atoms for binding the metal, or metal oxide, radionuclide. For example, U.S. Patent No. 4,673,562, to Davison *et al.* discloses representative chelating compounds and their synthesis.

A variety of routes of administration for the antibodies and immunoconjugates may be used. Typically, administration will be intravenous, intramuscular, subcutaneous or in the bed of a resected tumor. It will be evident that the precise dose of the antibody/immunoconjugate will vary depending upon the antibody used, the antigen density on the tumor, and the rate of clearance of the antibody.

T CELLS

Immunotherapeutic compositions may also, or alternatively, comprise T cells specific for a breast tumor protein. Such cells may generally be prepared *in vitro* or *ex vivo*, using standard procedures. For example, T cells may be isolated from bone marrow, peripheral blood, or a fraction of bone marrow or peripheral blood of a patient, using a commercially available cell separation system, such as the Isolex™ System, available from Nexell Therapeutics, Inc. (Irvine, CA; see also U.S. Patent No. 5,240,856; U.S. Patent No. 5,215,926; WO 89/06280; WO 91/16116 and WO

92/07243). Alternatively, T cells may be derived from related or unrelated humans, non-human mammals, cell lines or cultures.

T cells may be stimulated with a breast tumor polypeptide, polynucleotide encoding a breast tumor polypeptide and/or an antigen presenting cell (APC) that expresses such a polypeptide. Such stimulation is performed under conditions and for a time sufficient to permit the generation of T cells that are specific for the polypeptide. Preferably, a breast tumor polypeptide or polynucleotide is present within a delivery vehicle, such as a microsphere, to facilitate the generation of specific T cells.

10 T cells are considered to be specific for a breast tumor polypeptide if the T cells specifically proliferate, secrete cytokines or kill target cells coated with the polypeptide or expressing a gene encoding the polypeptide. T cell specificity may be evaluated using any of a variety of standard techniques. For example, within a chromium release assay or proliferation assay, a stimulation index of more than two
15 fold increase in lysis and/or proliferation, compared to negative controls, indicates T cell specificity. Such assays may be performed, for example, as described in Chen *et al.*, *Cancer Res.* 54:1065-1070, 1994. Alternatively, detection of the proliferation of T cells may be accomplished by a variety of known techniques. For example, T cell proliferation can be detected by measuring an increased rate of DNA synthesis (*e.g.*, by
20 pulse-labeling cultures of T cells with tritiated thymidine and measuring the amount of tritiated thymidine incorporated into DNA). Contact with a breast tumor polypeptide (100 ng/ml - 100 µg/ml, preferably 200 ng/ml - 25 µg/ml) for 3 - 7 days should result in at least a two fold increase in proliferation of the T cells. Contact as described above for 2-3 hours should result in activation of the T cells, as measured using standard
25 cytokine assays in which a two fold increase in the level of cytokine release (*e.g.*, TNF or IFN-γ) is indicative of T cell activation (*see* Coligan *et al.*, *Current Protocols in Immunology*, vol. 1, Wiley Interscience (Greene 1998)). T cells that have been activated in response to a breast tumor polypeptide, polynucleotide or polypeptide-expressing APC may be CD4⁺ and/or CD8⁺. Breast tumor protein-specific T cells may
30 be expanded using standard techniques. Within preferred embodiments, the T cells are

derived from a patient, a related donor or an unrelated donor, and are administered to the patient following stimulation and expansion.

For therapeutic purposes, CD4⁺ or CD8⁺ T cells that proliferate in response to a breast tumor polypeptide, polynucleotide or APC can be expanded in number either *in vitro* or *in vivo*. Proliferation of such T cells *in vitro* may be accomplished in a variety of ways. For example, the T cells can be re-exposed to a breast tumor polypeptide, or a short peptide corresponding to an immunogenic portion of such a polypeptide, with or without the addition of T cell growth factors, such as interleukin-2, and/or stimulator cells that synthesize a breast tumor polypeptide. Alternatively, one or more T cells that proliferate in the presence of a breast tumor protein can be expanded in number by cloning. Methods for cloning cells are well known in the art, and include limiting dilution.

PHARMACEUTICAL COMPOSITIONS

In additional embodiments, the present invention concerns formulation of one or more of the polynucleotide, polypeptide, T-cell and/or antibody compositions disclosed herein in pharmaceutically-acceptable solutions for administration to a cell or an animal, either alone, or in combination with one or more other modalities of therapy.

It will also be understood that, if desired, the nucleic acid segment, RNA, DNA or PNA compositions that express a polypeptide as disclosed herein may be administered in combination with other agents as well, such as, *e.g.*, other proteins or polypeptides or various pharmaceutically-active agents. In fact, there is virtually no limit to other components that may also be included, given that the additional agents do not cause a significant adverse effect upon contact with the target cells or host tissues. The compositions may thus be delivered along with various other agents as required in the particular instance. Such compositions may be purified from host cells or other biological sources, or alternatively may be chemically synthesized as described herein. Likewise, such compositions may further comprise substituted or derivatized RNA or DNA compositions.

Formulation of pharmaceutically-acceptable excipients and carrier solutions is well-known to those of skill in the art, as is the development of suitable

dosing and treatment regimens for using the particular compositions described herein in a variety of treatment regimens, including *e.g.*, oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation.

1. ORAL DELIVERY

5 In certain applications, the pharmaceutical compositions disclosed herein may be delivered *via* oral administration to an animal. As such, these compositions may be formulated with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

10 The active compounds may even be incorporated with excipients and used in the form of ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, syrups, wafers, and the like (Mathiowitz *et al.*, 1997; Hwang *et al.*, 1998; U. S. Patent 5,641,515; U. S. Patent 5,580,579 and U. S. Patent 5,792,451, each specifically incorporated herein by reference in its entirety). The tablets, troches, pills,
15 capsules and the like may also contain the following: a binder, as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such as dicalcium phosphate; a disintegrating agent, such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavoring agent, such as peppermint, oil of wintergreen, or cherry
20 flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar, or both. A syrup of elixir may contain the active compound sucrose as a sweetening agent methyl and
25 propylparabens as preservatives, a dye and flavoring, such as cherry or orange flavor. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compounds may be incorporated into sustained-release preparation and formulations.

Typically, these formulations may contain at least about 0.1% of the active compound or more, although the percentage of the active ingredient(s) may, of course, be varied and may conveniently be between about 1 or 2% and about 60% or 70% or more of the weight or volume of the total formulation. Naturally, the amount of active compound(s) in each therapeutically useful composition may be prepared in such a way that a suitable dosage will be obtained in any given unit dose of the compound. Factors such as solubility, bioavailability, biological half-life, route of administration, product shelf life, as well as other pharmacological considerations will be contemplated by one skilled in the art of preparing such pharmaceutical formulations, and as such, a variety of dosages and treatment regimens may be desirable.

For oral administration the compositions of the present invention may alternatively be incorporated with one or more excipients in the form of a mouthwash, dentifrice, buccal tablet, oral spray, or sublingual orally-administered formulation. For example, a mouthwash may be prepared incorporating the active ingredient in the required amount in an appropriate solvent, such as a sodium borate solution (Dobell's Solution). Alternatively, the active ingredient may be incorporated into an oral solution such as one containing sodium borate, glycerin and potassium bicarbonate, or dispersed in a dentifrice, or added in a therapeutically-effective amount to a composition that may include water, binders, abrasives, flavoring agents, foaming agents, and humectants. Alternatively the compositions may be fashioned into a tablet or solution form that may be placed under the tongue or otherwise dissolved in the mouth.

2. INJECTABLE DELIVERY

In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, or even intraperitoneally as described in U. S. Patent 5,543,158; U. S. Patent 5,641,515 and U. S. Patent 5,399,363 (each specifically incorporated herein by reference in its entirety). Solutions of the active compounds as free base or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of

storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (U. S. Patent 5,466,468, specifically incorporated herein by reference in its entirety). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (*e.g.*, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be facilitated by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile aqueous medium that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, "Remington's Pharmaceutical Sciences" 15th Edition, pages 1035-1038 and 1570-1580). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. The person responsible for administration will, in any event,

determine the appropriate dose for the individual subject. Moreover, for human administration, preparations should meet sterility, pyrogenicity, and the general safety and purity standards as required by FDA Office of Biologics standards.

Sterile injectable solutions are prepared by incorporating the active
5 compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the
10 preparation of sterile injectable solutions, the preferred methods of preparation are vacuum-drying and freeze-drying techniques which yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

The compositions disclosed herein may be formulated in a neutral or salt
15 form. Pharmaceutically-acceptable salts, include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium,
20 ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective. The formulations are easily administered in a variety of dosage forms such as injectable solutions, drug-release capsules, and the like.

25 As used herein, "carrier" includes any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active
30 ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when administered to a human. The preparation of an aqueous composition that contains a protein as an active ingredient is well understood in the art. Typically, such
5 compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection can also be prepared. The preparation can also be emulsified.

3. NASAL DELIVERY

In certain embodiments, the pharmaceutical compositions may be
10 delivered by intranasal sprays, inhalation, and/or other aerosol delivery vehicles. Methods for delivering genes, nucleic acids, and peptide compositions directly to the lungs *via* nasal aerosol sprays has been described *e.g.*, in U. S. Patent 5,756,353 and U. S. Patent 5,804,212 (each specifically incorporated herein by reference in its entirety). Likewise, the delivery of drugs using intranasal microparticle resins (Takenaga *et al.*,
15 1998) and lysophosphatidyl-glycerol compounds (U. S. Patent 5,725,871, specifically incorporated herein by reference in its entirety) are also well-known in the pharmaceutical arts. Likewise, transmucosal drug delivery in the form of a polytetrafluoroethylene support matrix is described in U. S. Patent 5,780,045 (specifically incorporated herein by reference in its entirety).

20 4. LIPOSOME-, NANOCAPSULE-, AND MICROPARTICLE-MEDIATED DELIVERY

In certain embodiments, the inventors contemplate the use of liposomes, nanocapsules, microparticles, microspheres, lipid particles, vesicles, and the like, for the introduction of the compositions of the present invention into suitable host cells. In particular, the compositions of the present invention may be formulated for delivery
25 either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like.

Such formulations may be preferred for the introduction of pharmaceutically-acceptable formulations of the nucleic acids or constructs disclosed herein. The formation and use of liposomes is generally known to those of skill in the

art (see for example, Couvreur *et al.*, 1977; Couvreur, 1988; Lasic, 1998; which describes the use of liposomes and nanocapsules in the targeted antibiotic therapy for intracellular bacterial infections and diseases). Recently, liposomes were developed with improved serum stability and circulation half-times (Gabizon and
5 Papahadjopoulos, 1988; Allen and Choun, 1987; U. S. Patent 5,741,516, specifically incorporated herein by reference in its entirety). Further, various methods of liposome and liposome like preparations as potential drug carriers have been reviewed (Takakura, 1998; Chandran *et al.*, 1997; Margalit, 1995; U. S. Patent 5,567,434; U. S. Patent 5,552,157; U. S. Patent 5,565,213; U. S. Patent 5,738,868 and U. S. Patent 5,795,587,
10 each specifically incorporated herein by reference in its entirety).

Liposomes have been used successfully with a number of cell types that are normally resistant to transfection by other procedures including T cell suspensions, primary hepatocyte cultures and PC 12 cells (Renneisen *et al.*, 1990; Muller *et al.*, 1990). In addition, liposomes are free of the DNA length constraints that are typical of
15 viral-based delivery systems. Liposomes have been used effectively to introduce genes, drugs (Heath and Martin, 1986; Heath *et al.*, 1986; Balazsovits *et al.*, 1989; Fresta and Puglisi, 1996), radiotherapeutic agents (Pikul *et al.*, 1987), enzymes (Imaizumi *et al.*, 1990a; Imaizumi *et al.*, 1990b), viruses (Faller and Baltimore, 1984), transcription factors and allosteric effectors (Nicolau and Gersonde, 1979) into a variety of cultured
20 cell lines and animals. In addition, several successful clinical trials examining the effectiveness of liposome-mediated drug delivery have been completed (Lopez-Berestein *et al.*, 1985a; 1985b; Coune, 1988; Sculier *et al.*, 1988). Furthermore, several studies suggest that the use of liposomes is not associated with autoimmune responses, toxicity or gonadal localization after systemic delivery (Mori and Fukatsu, 1992).

25 Liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs). MLVs generally have diameters of from 25 nm to 4 μ m. Sonication of MLVs results in the formation of small unilamellar vesicles (SUVs) with diameters in the range of 200 to 500 Å, containing an aqueous
30 solution in the core.

Liposomes bear resemblance to cellular membranes and are contemplated for use in connection with the present invention as carriers for the peptide compositions. They are widely suitable as both water- and lipid-soluble substances can be entrapped, *i.e.* in the aqueous spaces and within the bilayer itself, respectively. It is possible that the drug-bearing liposomes may even be employed for site-specific delivery of active agents by selectively modifying the liposomal formulation.

In addition to the teachings of Couvreur *et al.* (1977; 1988), the following information may be utilized in generating liposomal formulations. Phospholipids can form a variety of structures other than liposomes when dispersed in water, depending on the molar ratio of lipid to water. At low ratios the liposome is the preferred structure. The physical characteristics of liposomes depend on pH, ionic strength and the presence of divalent cations. Liposomes can show low permeability to ionic and polar substances, but at elevated temperatures undergo a phase transition which markedly alters their permeability. The phase transition involves a change from a closely packed, ordered structure, known as the gel state, to a loosely packed, less-ordered structure, known as the fluid state. This occurs at a characteristic phase-transition temperature and results in an increase in permeability to ions, sugars and drugs.

In addition to temperature, exposure to proteins can alter the permeability of liposomes. Certain soluble proteins, such as cytochrome c, bind, deform and penetrate the bilayer, thereby causing changes in permeability. Cholesterol inhibits this penetration of proteins, apparently by packing the phospholipids more tightly. It is contemplated that the most useful liposome formations for antibiotic and inhibitor delivery will contain cholesterol.

The ability to trap solutes varies between different types of liposomes. For example, MLVs are moderately efficient at trapping solutes, but SUVs are extremely inefficient. SUVs offer the advantage of homogeneity and reproducibility in size distribution, however, and a compromise between size and trapping efficiency is offered by large unilamellar vesicles (LUVs). These are prepared by ether evaporation and are three to four times more efficient at solute entrapment than MLVs.

In addition to liposome characteristics, an important determinant in entrapping compounds is the physicochemical properties of the compound itself. Polar compounds are trapped in the aqueous spaces and nonpolar compounds bind to the lipid bilayer of the vesicle. Polar compounds are released through permeation or when the bilayer is broken, but nonpolar compounds remain affiliated with the bilayer unless it is disrupted by temperature or exposure to lipoproteins. Both types show maximum efflux rates at the phase transition temperature.

Liposomes interact with cells *via* four different mechanisms: endocytosis by phagocytic cells of the reticuloendothelial system such as macrophages and neutrophils; adsorption to the cell surface, either by nonspecific weak hydrophobic or electrostatic forces, or by specific interactions with cell-surface components; fusion with the plasma cell membrane by insertion of the lipid bilayer of the liposome into the plasma membrane, with simultaneous release of liposomal contents into the cytoplasm; and by transfer of liposomal lipids to cellular or subcellular membranes, or vice versa, without any association of the liposome contents. It often is difficult to determine which mechanism is operative and more than one may operate at the same time.

The fate and disposition of intravenously injected liposomes depend on their physical properties, such as size, fluidity, and surface charge. They may persist in tissues for h or days, depending on their composition, and half lives in the blood range from min to several h. Larger liposomes, such as MLVs and LUVs, are taken up rapidly by phagocytic cells of the reticuloendothelial system, but physiology of the circulatory system restrains the exit of such large species at most sites. They can exit only in places where large openings or pores exist in the capillary endothelium, such as the sinusoids of the liver or spleen. Thus, these organs are the predominate site of uptake. On the other hand, SUVs show a broader tissue distribution but still are sequestered highly in the liver and spleen. In general, this *in vivo* behavior limits the potential targeting of liposomes to only those organs and tissues accessible to their large size. These include the blood, liver, spleen, bone marrow, and lymphoid organs.

Targeting is generally not a limitation in terms of the present invention. However, should specific targeting be desired, methods are available for this to be accomplished. Antibodies may be used to bind to the liposome surface and to direct the

antibody and its drug contents to specific antigenic receptors located on a particular cell-type surface. Carbohydrate determinants (glycoprotein or glycolipid cell-surface components that play a role in cell-cell recognition, interaction and adhesion) may also be used as recognition sites as they have potential in directing liposomes to particular
5 cell types. Mostly, it is contemplated that intravenous injection of liposomal preparations would be used, but other routes of administration are also conceivable.

Alternatively, the invention provides for pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (Henry-Michelland
10 *et al.*, 1987; Quintanar-Guerrero *et al.*, 1998; Douglas *et al.*, 1987). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1 μm) should be designed using polymers able to be degraded *in vivo*. Biodegradable polyalkyl-cyanoacrylate nanoparticles that meet these requirements are contemplated for use in the present invention. Such particles may be easily made, as described
15 (Couvreur *et al.*, 1980; 1988; zur Muhlen *et al.*, 1998; Zambaux *et al.* 1998; Pinto-Alphandry *et al.*, 1995 and U. S. Patent 5,145,684, specifically incorporated herein by reference in its entirety).

VACCINES

In certain preferred embodiments of the present invention, vaccines are
20 provided. The vaccines will generally comprise one or more pharmaceutical compositions, such as those discussed above, in combination with an immunostimulant. An immunostimulant may be any substance that enhances or potentiates an immune response (antibody and/or cell-mediated) to an exogenous antigen. Examples of immunostimulants include adjuvants, biodegradable microspheres (*e.g.*, polylactic galactide) and liposomes (into which the compound is incorporated; *see e.g.*, Fullerton,
25 U.S. Patent No. 4,235,877). Vaccine preparation is generally described in, for example, M.F. Powell and M.J. Newman, eds., "Vaccine Design (the subunit and adjuvant approach)," Plenum Press (NY, 1995). Pharmaceutical compositions and vaccines within the scope of the present invention may also contain other compounds, which
30 may be biologically active or inactive. For example, one or more immunogenic

portions of other tumor antigens may be present, either incorporated into a fusion polypeptide or as a separate compound, within the composition or vaccine.

Illustrative vaccines may contain DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. As noted above, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid expression systems, bacteria and viral expression systems. Numerous gene delivery techniques are well known in the art, such as those described by Rolland, *Crit. Rev. Therap. Drug Carrier Systems* 15:143-198, 1998, and references cited therein. Appropriate nucleic acid expression systems contain the necessary DNA sequences for expression in the patient (such as a suitable promoter and terminating signal). Bacterial delivery systems involve the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the polypeptide on its cell surface or secretes such an epitope. In a preferred embodiment, the DNA may be introduced using a viral expression system (*e.g.*, vaccinia or other pox virus, retrovirus, or adenovirus), which may involve the use of a non-pathogenic (defective), replication competent virus. Suitable systems are disclosed, for example, in Fisher-Hoch *et al.*, *Proc. Natl. Acad. Sci. USA* 86:317-321, 1989; Flexner *et al.*, *Ann. N.Y. Acad. Sci.* 569:86-103, 1989; Flexner *et al.*, *Vaccine* 8:17-21, 1990; U.S. Patent Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Patent No. 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; Berkner, *Biotechniques* 6:616-627, 1988; Rosenfeld *et al.*, *Science* 252:431-434, 1991; Kolls *et al.*, *Proc. Natl. Acad. Sci. USA* 91:215-219, 1994; Kass-Eisler *et al.*, *Proc. Natl. Acad. Sci. USA* 90:11498-11502, 1993; Guzman *et al.*, *Circulation* 88:2838-2848, 1993; and Guzman *et al.*, *Cir. Res.* 73:1202-1207, 1993. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer *et al.*, *Science* 259:1745-1749, 1993 and reviewed by Cohen, *Science* 259:1691-1692, 1993. The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells. It will be apparent that a vaccine may comprise both a polynucleotide and a polypeptide component. Such vaccines may provide for an enhanced immune response.

It will be apparent that a vaccine may contain pharmaceutically acceptable salts of the polynucleotides and polypeptides provided herein. Such salts may be prepared from pharmaceutically acceptable non-toxic bases, including organic bases (*e.g.*, salts of primary, secondary and tertiary amines and basic amino acids) and
5 inorganic bases (*e.g.*, sodium, potassium, lithium, ammonium, calcium and magnesium salts).

While any suitable carrier known to those of ordinary skill in the art may be employed in the vaccine compositions of this invention, the type of carrier will vary depending on the mode of administration. Compositions of the present invention may
10 be formulated for any appropriate manner of administration, including for example, topical, oral, nasal, intravenous, intracranial, intraperitoneal, subcutaneous or intramuscular administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol,
15 lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (*e.g.*, polylactate polyglycolate) may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268; 5,075,109; 5,928,647;
20 5,811,128; 5,820,883; 5,853,763; 5,814,344 and 5,942,252. One may also employ a carrier comprising the particulate-protein complexes described in U.S. Patent No. 5,928,647, which are capable of inducing a class I-restricted cytotoxic T lymphocyte responses in a host.

Such compositions may also comprise buffers (*e.g.*, neutral buffered
25 saline or phosphate buffered saline), carbohydrates (*e.g.*, glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents such as EDTA or glutathione, adjuvants (*e.g.*, aluminum hydroxide), solutes that render the formulation isotonic, hypotonic or weakly hypertonic with the blood of a recipient, suspending agents, thickening agents and/or preservatives.
30 Alternatively, compositions of the present invention may be formulated as a

lyophilizate. Compounds may also be encapsulated within liposomes using well known technology.

Any of a variety of immunostimulants may be employed in the vaccines of this invention. For example, an adjuvant may be included. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a stimulator of immune responses, such as lipid A, *Bordetella pertussis* or *Mycobacterium tuberculosis* derived proteins. Suitable adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI); Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ); AS-2 (SmithKline Beecham, Philadelphia, PA); aluminum salts such as aluminum hydroxide gel (alum) or aluminum phosphate; salts of calcium, iron or zinc; an insoluble suspension of acylated tyrosine; acylated sugars; cationically or anionically derivatized polysaccharides; polyphosphazenes; biodegradable microspheres; monophosphoryl lipid A and quil A. Cytokines, such as GM-CSF or interleukin-2, -7, or -12, may also be used as adjuvants.

Within the vaccines provided herein, the adjuvant composition is preferably designed to induce an immune response predominantly of the Th1 type. High levels of Th1-type cytokines (*e.g.*, IFN- γ , TNF α , IL-2 and IL-12) tend to favor the induction of cell mediated immune responses to an administered antigen. In contrast, high levels of Th2-type cytokines (*e.g.*, IL-4, IL-5, IL-6 and IL-10) tend to favor the induction of humoral immune responses. Following application of a vaccine as provided herein, a patient will support an immune response that includes Th1- and Th2-type responses. Within a preferred embodiment, in which a response is predominantly Th1-type, the level of Th1-type cytokines will increase to a greater extent than the level of Th2-type cytokines. The levels of these cytokines may be readily assessed using standard assays. For a review of the families of cytokines, see Mosmann and Coffman, *Ann. Rev. Immunol.* 7:145-173, 1989.

Preferred adjuvants for use in eliciting a predominantly Th1-type response include, for example, a combination of monophosphoryl lipid A, preferably 3-de-O-acylated monophosphoryl lipid A (3D-MPL), together with an aluminum salt. MPL adjuvants are available from Corixa Corporation (Seattle, WA; *see* US Patent

Nos. 4,436,727; 4,877,611; 4,866,034 and 4,912,094). CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1 response. Such oligonucleotides are well known and are described, for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and
5 5,856,462. Immunostimulatory DNA sequences are also described, for example, by Sato *et al.*, *Science* 273:352, 1996. Another preferred adjuvant is a saponin, preferably QS21 (Aquila Biopharmaceuticals Inc., Framingham, MA), which may be used alone or in combination with other adjuvants. For example, an enhanced system involves the combination of a monophosphoryl lipid A and saponin derivative, such as the
10 combination of QS21 and 3D-MPL as described in WO 94/00153, or a less reactogenic composition where the QS21 is quenched with cholesterol, as described in WO 96/33739. Other preferred formulations comprise an oil-in-water emulsion and tocopherol. A particularly potent adjuvant formulation involving QS21, 3D-MPL and tocopherol in an oil-in-water emulsion is described in WO 95/17210.

15 Other preferred adjuvants include Montanide ISA 720 (Seppic, France), SAF (Chiron, California, United States), ISCOMS (CSL), MF-59 (Chiron), the SBAS series of adjuvants (*e.g.*, SBAS-2 or SBAS-4, available from SmithKline Beecham, Rixensart, Belgium), Detox (Corixa, Hamilton, MT), RC-529 (Corixa, Hamilton, MT) and other aminoalkyl glucosaminide 4-phosphates (AGPs), such as those described in
20 pending U.S. Patent Application Serial Nos. 08/853,826 and 09/074,720, the disclosures of which are incorporated herein by reference in their entireties.

Any vaccine provided herein may be prepared using well known methods that result in a combination of antigen, immune response enhancer and a suitable carrier or excipient. The compositions described herein may be administered
25 as part of a sustained release formulation (*i.e.*, a formulation such as a capsule, sponge or gel (composed of polysaccharides, for example) that effects a slow release of compound following administration). Such formulations may generally be prepared using well known technology (*see, e.g.*, Coombes *et al.*, *Vaccine* 14:1429-1438, 1996) and administered by, for example, oral, rectal or subcutaneous implantation, or by
30 implantation at the desired target site. Sustained-release formulations may contain a

polypeptide, polynucleotide or antibody dispersed in a carrier matrix and/or contained within a reservoir surrounded by a rate controlling membrane.

Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of
5 active component release. Such carriers include microparticles of poly(lactide-co-glycolide), polyacrylate, latex, starch, cellulose, dextran and the like. Other delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (*e.g.*, a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer comprising an amphiphilic compound, such as a
10 phospholipid (*see e.g.*, U.S. Patent No. 5,151,254 and PCT applications WO 94/20078, WO/94/23701 and WO 96/06638). The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

Any of a variety of delivery vehicles may be employed within
15 pharmaceutical compositions and vaccines to facilitate production of an antigen-specific immune response that targets tumor cells. Delivery vehicles include antigen presenting cells (APCs), such as dendritic cells, macrophages, B cells, monocytes and other cells that may be engineered to be efficient APCs. Such cells may, but need not, be genetically modified to increase the capacity for presenting the antigen, to improve
20 activation and/or maintenance of the T cell response, to have anti-tumor effects *per se* and/or to be immunologically compatible with the receiver (*i.e.*, matched HLA haplotype). APCs may generally be isolated from any of a variety of biological fluids and organs, including tumor and peritumoral tissues, and may be autologous, allogeneic, syngeneic or xenogeneic cells.

25 Certain preferred embodiments of the present invention use dendritic cells or progenitors thereof as antigen-presenting cells. Dendritic cells are highly potent APCs (Banchereau and Steinman, *Nature* 392:245-251, 1998) and have been shown to be effective as a physiological adjuvant for eliciting prophylactic or therapeutic antitumor immunity (*see* Timmerman and Levy, *Ann. Rev. Med.* 50:507-529, 1999). In
30 general, dendritic cells may be identified based on their typical shape (stellate *in situ*, with marked cytoplasmic processes (dendrites) visible *in vitro*), their ability to take up,

process and present antigens with high efficiency and their ability to activate naïve T cell responses. Dendritic cells may, of course, be engineered to express specific cell-surface receptors or ligands that are not commonly found on dendritic cells *in vivo* or *ex vivo*, and such modified dendritic cells are contemplated by the present invention. As
5 an alternative to dendritic cells, secreted vesicles antigen-loaded dendritic cells (called exosomes) may be used within a vaccine (*see Zitvogel et al., Nature Med. 4:594-600, 1998*).

Dendritic cells and progenitors may be obtained from peripheral blood, bone marrow, tumor-infiltrating cells, peritumoral tissues-infiltrating cells, lymph
10 nodes, spleen, skin, umbilical cord blood or any other suitable tissue or fluid. For example, dendritic cells may be differentiated *ex vivo* by adding a combination of cytokines such as GM-CSF, IL-4, IL-13 and/or TNF α to cultures of monocytes harvested from peripheral blood. Alternatively, CD34 positive cells harvested from peripheral blood, umbilical cord blood or bone marrow may be differentiated into
15 dendritic cells by adding to the culture medium combinations of GM-CSF, IL-3, TNF α , CD40 ligand, LPS, flt3 ligand and/or other compound(s) that induce differentiation, maturation and proliferation of dendritic cells.

Dendritic cells are conveniently categorized as "immature" and "mature" cells, which allows a simple way to discriminate between two well characterized
20 phenotypes. However, this nomenclature should not be construed to exclude all possible intermediate stages of differentiation. Immature dendritic cells are characterized as APC with a high capacity for antigen uptake and processing, which correlates with the high expression of Fc γ receptor and mannose receptor. The mature phenotype is typically characterized by a lower expression of these markers, but a high
25 expression of cell surface molecules responsible for T cell activation such as class I and class II MHC, adhesion molecules (*e.g.*, CD54 and CD11) and costimulatory molecules (*e.g.*, CD40, CD80, CD86 and 4-1BB).

APCs may generally be transfected with a polynucleotide encoding a breast tumor protein (or portion or other variant thereof) such that the breast tumor
30 polypeptide, or an immunogenic portion thereof, is expressed on the cell surface. Such transfection may take place *ex vivo*, and a composition or vaccine comprising such

transfected cells may then be used for therapeutic purposes, as described herein. Alternatively, a gene delivery vehicle that targets a dendritic or other antigen presenting cell may be administered to a patient, resulting in transfection that occurs *in vivo*. *In vivo* and *ex vivo* transfection of dendritic cells, for example, may generally be performed using any methods known in the art, such as those described in WO 97/24447, or the gene gun approach described by Mahvi *et al.*, *Immunology and cell Biology* 75:456-460, 1997. Antigen loading of dendritic cells may be achieved by incubating dendritic cells or progenitor cells with the breast tumor polypeptide, DNA (naked or within a plasmid vector) or RNA; or with antigen-expressing recombinant bacterium or viruses (*e.g.*, vaccinia, fowlpox, adenovirus or lentivirus vectors). Prior to loading, the polypeptide may be covalently conjugated to an immunological partner that provides T cell help (*e.g.*, a carrier molecule). Alternatively, a dendritic cell may be pulsed with a non-conjugated immunological partner, separately or in the presence of the polypeptide.

Vaccines and pharmaceutical compositions may be presented in unit-dose or multi-dose containers, such as sealed ampoules or vials. Such containers are preferably hermetically sealed to preserve sterility of the formulation until use. In general, formulations may be stored as suspensions, solutions or emulsions in oily or aqueous vehicles. Alternatively, a vaccine or pharmaceutical composition may be stored in a freeze-dried condition requiring only the addition of a sterile liquid carrier immediately prior to use.

CANCER THERAPY

In further aspects of the present invention, the compositions described herein may be used for immunotherapy of cancer, such as breast cancer. Within such methods, pharmaceutical compositions and vaccines are typically administered to a patient. As used herein, a "patient" refers to any warm-blooded animal, preferably a human. A patient may or may not be afflicted with cancer. Accordingly, the above pharmaceutical compositions and vaccines may be used to prevent the development of a cancer or to treat a patient afflicted with a cancer. A cancer may be diagnosed using criteria generally accepted in the art, including the presence of a malignant tumor.

Pharmaceutical compositions and vaccines may be administered either prior to or following surgical removal of primary tumors and/or treatment such as administration of radiotherapy or conventional chemotherapeutic drugs. Administration may be by any suitable method, including administration by intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal, intradermal, anal, vaginal, topical and oral routes.

Within certain embodiments, immunotherapy may be active immunotherapy, in which treatment relies on the *in vivo* stimulation of the endogenous host immune system to react against tumors with the administration of immune response-modifying agents (such as polypeptides and polynucleotides as provided herein).

Within other embodiments, immunotherapy may be passive immunotherapy, in which treatment involves the delivery of agents with established tumor-immune reactivity (such as effector cells or antibodies) that can directly or indirectly mediate antitumor effects and does not necessarily depend on an intact host immune system. Examples of effector cells include T cells as discussed above, T lymphocytes (such as CD8⁺ cytotoxic T lymphocytes and CD4⁺ T-helper tumor-infiltrating lymphocytes), killer cells (such as Natural Killer cells and lymphokine-activated killer cells), B cells and antigen-presenting cells (such as dendritic cells and macrophages) expressing a polypeptide provided herein. T cell receptors and antibody receptors specific for the polypeptides recited herein may be cloned, expressed and transferred into other vectors or effector cells for adoptive immunotherapy. The polypeptides provided herein may also be used to generate antibodies or anti-idiotypic antibodies (as described above and in U.S. Patent No. 4,918,164) for passive immunotherapy.

Effector cells may generally be obtained in sufficient quantities for adoptive immunotherapy by growth *in vitro*, as described herein. Culture conditions for expanding single antigen-specific effector cells to several billion in number with retention of antigen recognition *in vivo* are well known in the art. Such *in vitro* culture conditions typically use intermittent stimulation with antigen, often in the presence of cytokines (such as IL-2) and non-dividing feeder cells. As noted above,

immunoreactive polypeptides as provided herein may be used to rapidly expand antigen-specific T cell cultures in order to generate a sufficient number of cells for immunotherapy. In particular, antigen-presenting cells, such as dendritic, macrophage, monocyte, fibroblast and/or B cells, may be pulsed with immunoreactive polypeptides or transfected with one or more polynucleotides using standard techniques well known in the art. For example, antigen-presenting cells can be transfected with a polynucleotide having a promoter appropriate for increasing expression in a recombinant virus or other expression system. Cultured effector cells for use in therapy must be able to grow and distribute widely, and to survive long term *in vivo*. Studies have shown that cultured effector cells can be induced to grow *in vivo* and to survive long term in substantial numbers by repeated stimulation with antigen supplemented with IL-2 (*see, for example, Cheever et al., Immunological Reviews 157:177, 1997*).

Alternatively, a vector expressing a polypeptide recited herein may be introduced into antigen presenting cells taken from a patient and clonally propagated *ex vivo* for transplant back into the same patient. Transfected cells may be reintroduced into the patient using any means known in the art, preferably in sterile form by intravenous, intracavitary, intraperitoneal or intratumor administration.

Routes and frequency of administration of the therapeutic compositions described herein, as well as dosage, will vary from individual to individual, and may be readily established using standard techniques. In general, the pharmaceutical compositions and vaccines may be administered by injection (*e.g.*, intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (*e.g.*, by aspiration) or orally. Preferably, between 1 and 10 doses may be administered over a 52 week period. Preferably, 6 doses are administered, at intervals of 1 month, and booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of a compound that, when administered as described above, is capable of promoting an anti-tumor immune response, and is at least 10-50% above the basal (*i.e.*, untreated) level. Such response can be monitored by measuring the anti-tumor antibodies in a patient or by vaccine-dependent generation of cytolytic effector cells capable of killing the patient's tumor cells *in vitro*. Such vaccines should also be capable of causing an immune response that

leads to an improved clinical outcome (e.g., more frequent remissions, complete or partial or longer disease-free survival) in vaccinated patients as compared to non-vaccinated patients. In general, for pharmaceutical compositions and vaccines comprising one or more polypeptides, the amount of each polypeptide present in a dose
5 ranges from about 25 µg to 5 mg per kg of host. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In general, an appropriate dosage and treatment regimen provides the active compound(s) in an amount sufficient to provide therapeutic and/or prophylactic benefit. Such a response can be monitored by establishing an improved clinical
10 outcome (e.g., more frequent remissions, complete or partial, or longer disease-free survival) in treated patients as compared to non-treated patients. Increases in preexisting immune responses to a breast tumor protein generally correlate with an improved clinical outcome. Such immune responses may generally be evaluated using standard proliferation, cytotoxicity or cytokine assays, which may be performed using
15 samples obtained from a patient before and after treatment.

CANCER DETECTION AND DIAGNOSIS

In general, a cancer may be detected in a patient based on the presence of one or more breast tumor proteins and/or polynucleotides encoding such proteins in a biological sample (for example, blood, sera, sputum urine and/or tumor biopsies)
20 obtained from the patient. In other words, such proteins may be used as markers to indicate the presence or absence of a cancer such as breast cancer. In addition, such proteins may be useful for the detection of other cancers. The binding agents provided herein generally permit detection of the level of antigen that binds to the agent in the biological sample. Polynucleotide primers and probes may be used to detect the level
25 of mRNA encoding a tumor protein, which is also indicative of the presence or absence of a cancer. In general, a breast tumor sequence should be present at a level that is at least three fold higher in tumor tissue than in normal tissue

There are a variety of assay formats known to those of ordinary skill in the art for using a binding agent to detect polypeptide markers in a sample. *See, e.g.,*
30 Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory,

1988. In general, the presence or absence of a cancer in a patient may be determined by (a) contacting a biological sample obtained from a patient with a binding agent; (b) detecting in the sample a level of polypeptide that binds to the binding agent; and (c) comparing the level of polypeptide with a predetermined cut-off value.

5 In a preferred embodiment, the assay involves the use of binding agent immobilized on a solid support to bind to and remove the polypeptide from the remainder of the sample. The bound polypeptide may then be detected using a detection reagent that contains a reporter group and specifically binds to the binding agent/polypeptide complex. Such detection reagents may comprise, for example, a
10 binding agent that specifically binds to the polypeptide or an antibody or other agent that specifically binds to the binding agent, such as an anti-immunoglobulin, protein G, protein A or a lectin. Alternatively, a competitive assay may be utilized, in which a polypeptide is labeled with a reporter group and allowed to bind to the immobilized binding agent after incubation of the binding agent with the sample. The extent to
15 which components of the sample inhibit the binding of the labeled polypeptide to the binding agent is indicative of the reactivity of the sample with the immobilized binding agent. Suitable polypeptides for use within such assays include full length breast tumor proteins and portions thereof to which the binding agent binds, as described above.

The solid support may be any material known to those of ordinary skill
20 in the art to which the tumor protein may be attached. For example, the solid support may be a test well in a microtiter plate or a nitrocellulose or other suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic material such as polystyrene or polyvinylchloride. The support may also be a magnetic particle or a fiber optic sensor, such as those disclosed, for example, in U.S.
25 Patent No. 5,359,681. The binding agent may be immobilized on the solid support using a variety of techniques known to those of skill in the art, which are amply described in the patent and scientific literature. In the context of the present invention, the term "immobilization" refers to both noncovalent association, such as adsorption, and covalent attachment (which may be a direct linkage between the agent and
30 functional groups on the support or may be a linkage by way of a cross-linking agent). Immobilization by adsorption to a well in a microtiter plate or to a membrane is

preferred. In such cases, adsorption may be achieved by contacting the binding agent, in a suitable buffer, with the solid support for a suitable amount of time. The contact time varies with temperature, but is typically between about 1 hour and about 1 day. In general, contacting a well of a plastic microtiter plate (such as polystyrene or polyvinylchloride) with an amount of binding agent ranging from about 10 ng to about 10 μ g, and preferably about 100 ng to about 1 μ g, is sufficient to immobilize an adequate amount of binding agent.

Covalent attachment of binding agent to a solid support may generally be achieved by first reacting the support with a bifunctional reagent that will react with both the support and a functional group, such as a hydroxyl or amino group, on the binding agent. For example, the binding agent may be covalently attached to supports having an appropriate polymer coating using benzoquinone or by condensation of an aldehyde group on the support with an amine and an active hydrogen on the binding partner (*see, e.g.,* Pierce Immunotechnology Catalog and Handbook, 1991, at A12-A13).

In certain embodiments, the assay is a two-antibody sandwich assay. This assay may be performed by first contacting an antibody that has been immobilized on a solid support, commonly the well of a microtiter plate, with the sample, such that polypeptides within the sample are allowed to bind to the immobilized antibody. Unbound sample is then removed from the immobilized polypeptide-antibody complexes and a detection reagent (preferably a second antibody capable of binding to a different site on the polypeptide) containing a reporter group is added. The amount of detection reagent that remains bound to the solid support is then determined using a method appropriate for the specific reporter group.

More specifically, once the antibody is immobilized on the support as described above, the remaining protein binding sites on the support are typically blocked. Any suitable blocking agent known to those of ordinary skill in the art, such as bovine serum albumin or Tween 20TM (Sigma Chemical Co., St. Louis, MO). The immobilized antibody is then incubated with the sample, and polypeptide is allowed to bind to the antibody. The sample may be diluted with a suitable diluent, such as phosphate-buffered saline (PBS) prior to incubation. In general, an appropriate contact

time (*i.e.*, incubation time) is a period of time that is sufficient to detect the presence of polypeptide within a sample obtained from an individual with breast cancer. Preferably, the contact time is sufficient to achieve a level of binding that is at least about 95% of that achieved at equilibrium between bound and unbound polypeptide.

- 5 Those of ordinary skill in the art will recognize that the time necessary to achieve equilibrium may be readily determined by assaying the level of binding that occurs over a period of time. At room temperature, an incubation time of about 30 minutes is generally sufficient.

Unbound sample may then be removed by washing the solid support
10 with an appropriate buffer, such as PBS containing 0.1% Tween 20™. The second antibody, which contains a reporter group, may then be added to the solid support. Preferred reporter groups include those groups recited above.

- The detection reagent is then incubated with the immobilized antibody-polypeptide complex for an amount of time sufficient to detect the bound polypeptide.
15 An appropriate amount of time may generally be determined by assaying the level of binding that occurs over a period of time. Unbound detection reagent is then removed and bound detection reagent is detected using the reporter group. The method employed for detecting the reporter group depends upon the nature of the reporter group. For radioactive groups, scintillation counting or autoradiographic methods are
20 generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups and fluorescent groups. Biotin may be detected using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme). Enzyme reporter groups may generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of
25 the reaction products.

To determine the presence or absence of a cancer, such as breast cancer, the signal detected from the reporter group that remains bound to the solid support is generally compared to a signal that corresponds to a predetermined cut-off value. In one preferred embodiment, the cut-off value for the detection of a cancer is the average
30 mean signal obtained when the immobilized antibody is incubated with samples from patients without the cancer. In general, a sample generating a signal that is three

standard deviations above the predetermined cut-off value is considered positive for the cancer. In an alternate preferred embodiment, the cut-off value is determined using a Receiver Operator Curve, according to the method of Sackett *et al.*, *Clinical Epidemiology: A Basic Science for Clinical Medicine*, Little Brown and Co., 1985, 5 p. 106-7. Briefly, in this embodiment, the cut-off value may be determined from a plot of pairs of true positive rates (*i.e.*, sensitivity) and false positive rates (100%-specificity) that correspond to each possible cut-off value for the diagnostic test result. The cut-off value on the plot that is the closest to the upper left-hand corner (*i.e.*, the value that encloses the largest area) is the most accurate cut-off value, and a sample 10 generating a signal that is higher than the cut-off value determined by this method may be considered positive. Alternatively, the cut-off value may be shifted to the left along the plot, to minimize the false positive rate, or to the right, to minimize the false negative rate. In general, a sample generating a signal that is higher than the cut-off value determined by this method is considered positive for a cancer.

15 In a related embodiment, the assay is performed in a flow-through or strip test format, wherein the binding agent is immobilized on a membrane, such as nitrocellulose. In the flow-through test, polypeptides within the sample bind to the immobilized binding agent as the sample passes through the membrane. A second, labeled binding agent then binds to the binding agent-polypeptide complex as a solution 20 containing the second binding agent flows through the membrane. The detection of bound second binding agent may then be performed as described above. In the strip test format, one end of the membrane to which binding agent is bound is immersed in a solution containing the sample. The sample migrates along the membrane through a region containing second binding agent and to the area of immobilized binding agent. 25 Concentration of second binding agent at the area of immobilized antibody indicates the presence of a cancer. Typically, the concentration of second binding agent at that site generates a pattern, such as a line, that can be read visually. The absence of such a pattern indicates a negative result. In general, the amount of binding agent immobilized on the membrane is selected to generate a visually discernible pattern when the 30 biological sample contains a level of polypeptide that would be sufficient to generate a positive signal in the two-antibody sandwich assay, in the format discussed above.

Preferred binding agents for use in such assays are antibodies and antigen-binding fragments thereof. Preferably, the amount of antibody immobilized on the membrane ranges from about 25 ng to about 1 μ g, and more preferably from about 50 ng to about 500 ng. Such tests can typically be performed with a very small amount of biological sample.

Of course, numerous other assay protocols exist that are suitable for use with the tumor proteins or binding agents of the present invention. The above descriptions are intended to be exemplary only. For example, it will be apparent to those of ordinary skill in the art that the above protocols may be readily modified to use breast tumor polypeptides to detect antibodies that bind to such polypeptides in a biological sample. The detection of such breast tumor protein specific antibodies may correlate with the presence of a cancer.

A cancer may also, or alternatively, be detected based on the presence of T cells that specifically react with a breast tumor protein in a biological sample. Within certain methods, a biological sample comprising CD4⁺ and/or CD8⁺ T cells isolated from a patient is incubated with a breast tumor polypeptide, a polynucleotide encoding such a polypeptide and/or an APC that expresses at least an immunogenic portion of such a polypeptide, and the presence or absence of specific activation of the T cells is detected. Suitable biological samples include, but are not limited to, isolated T cells. For example, T cells may be isolated from a patient by routine techniques (such as by Ficoll/Hypaque density gradient centrifugation of peripheral blood lymphocytes). T cells may be incubated *in vitro* for 2-9 days (typically 4 days) at 37°C with polypeptide (*e.g.*, 5 - 25 μ g/ml). It may be desirable to incubate another aliquot of a T cell sample in the absence of breast tumor polypeptide to serve as a control. For CD4⁺ T cells, activation is preferably detected by evaluating proliferation of the T cells. For CD8⁺ T cells, activation is preferably detected by evaluating cytolytic activity. A level of proliferation that is at least two fold greater and/or a level of cytolytic activity that is at least 20% greater than in disease-free patients indicates the presence of a cancer in the patient.

As noted above, a cancer may also, or alternatively, be detected based on the level of mRNA encoding a breast tumor protein in a biological sample. For

example, at least two oligonucleotide primers may be employed in a polymerase chain reaction (PCR) based assay to amplify a portion of a breast tumor cDNA derived from a biological sample, wherein at least one of the oligonucleotide primers is specific for (*i.e.*, hybridizes to) a polynucleotide encoding the breast tumor protein. The amplified
5 cDNA is then separated and detected using techniques well known in the art, such as gel electrophoresis. Similarly, oligonucleotide probes that specifically hybridize to a polynucleotide encoding a breast tumor protein may be used in a hybridization assay to detect the presence of polynucleotide encoding the tumor protein in a biological sample.

To permit hybridization under assay conditions, oligonucleotide primers
10 and probes should comprise an oligonucleotide sequence that has at least about 60%, preferably at least about 75% and more preferably at least about 90%, identity to a portion of a polynucleotide encoding a breast tumor protein that is at least 10 nucleotides, and preferably at least 20 nucleotides, in length. Preferably, oligonucleotide primers and/or probes hybridize to a polynucleotide encoding a
15 polypeptide described herein under moderately stringent conditions, as defined above. Oligonucleotide primers and/or probes which may be usefully employed in the diagnostic methods described herein preferably are at least 10-40 nucleotides in length. In a preferred embodiment, the oligonucleotide primers comprise at least 10 contiguous nucleotides, more preferably at least 15 contiguous nucleotides, of a DNA molecule
20 having a sequence recited in SEQ ID NO:1-____. Techniques for both PCR based assays and hybridization assays are well known in the art (*see*, for example, Mullis *et al.*, *Cold Spring Harbor Symp. Quant. Biol.*, 51:263, 1987; Erlich ed., *PCR Technology*, Stockton Press, NY, 1989).

One preferred assay employs RT-PCR, in which PCR is applied in
25 conjunction with reverse transcription. Typically, RNA is extracted from a biological sample, such as biopsy tissue, and is reverse transcribed to produce cDNA molecules. PCR amplification using at least one specific primer generates a cDNA molecule, which may be separated and visualized using, for example, gel electrophoresis. Amplification may be performed on biological samples taken from a test patient and
30 from an individual who is not afflicted with a cancer. The amplification reaction may be performed on several dilutions of cDNA spanning two orders of magnitude. A two-

fold or greater increase in expression in several dilutions of the test patient sample as compared to the same dilutions of the non-cancerous sample is typically considered positive.

In another embodiment, the compositions described herein may be used
5 as markers for the progression of cancer. In this embodiment, assays as described above for the diagnosis of a cancer may be performed over time, and the change in the level of reactive polypeptide(s) or polynucleotide(s) evaluated. For example, the assays may be performed every 24-72 hours for a period of 6 months to 1 year, and thereafter performed as needed. In general, a cancer is progressing in those patients in whom the
10 level of polypeptide or polynucleotide detected increases over time. In contrast, the cancer is not progressing when the level of reactive polypeptide or polynucleotide either remains constant or decreases with time.

Certain *in vivo* diagnostic assays may be performed directly on a tumor. One such assay involves contacting tumor cells with a binding agent. The bound
15 binding agent may then be detected directly or indirectly via a reporter group. Such binding agents may also be used in histological applications. Alternatively, polynucleotide probes may be used within such applications.

As noted above, to improve sensitivity, multiple breast tumor protein markers may be assayed within a given sample. It will be apparent that binding agents
20 specific for different proteins provided herein may be combined within a single assay. Further, multiple primers or probes may be used concurrently. The selection of tumor protein markers may be based on routine experiments to determine combinations that results in optimal sensitivity. In addition, or alternatively, assays for tumor proteins provided herein may be combined with assays for other known tumor antigens.

25 DIAGNOSTIC KITS

The present invention further provides kits for use within any of the above diagnostic methods. Such kits typically comprise two or more components necessary for performing a diagnostic assay. Components may be compounds, reagents, containers and/or equipment. For example, one container within a kit may
30 contain a monoclonal antibody or fragment thereof that specifically binds to a breast

tumor protein. Such antibodies or fragments may be provided attached to a support material, as described above. One or more additional containers may enclose elements, such as reagents or buffers, to be used in the assay. Such kits may also, or alternatively, contain a detection reagent as described above that contains a reporter group suitable
5 for direct or indirect detection of antibody binding.

Alternatively, a kit may be designed to detect the level of mRNA encoding a breast tumor protein in a biological sample. Such kits generally comprise at least one oligonucleotide probe or primer, as described above, that hybridizes to a polynucleotide encoding a breast tumor protein. Such an oligonucleotide may be used,
10 for example, within a PCR or hybridization assay. Additional components that may be present within such kits include a second oligonucleotide and/or a diagnostic reagent or container to facilitate the detection of a polynucleotide encoding a breast tumor protein.

The following Examples are offered by way of illustration and not by way of limitation.

EXAMPLE 1

IDENTIFICATION OF BREAST TUMOR PROTEIN CDNAS

5 This Example illustrates the identification of cDNA molecules encoding breast tumor proteins.

Potential breast-specific genes present in the GenBank human EST database were identified by electronic subtraction (similar to that described by Vasmatizis et al., *Proc. Natl. Acad. Sci. USA* 95:300-304, 1998). The sequences of EST
10 clones (26,074) derived from various human breast cDNA libraries were obtained from the GenBank public human EST database. Each breast EST sequence was used as a query sequence in a BLASTN (National Center for Biotechnology Information) search against the human EST database. All matches considered identical (BLAST score > 40, length of matching sequence >100 base pairs, density of identical matches over this
15 region > 80%) were grouped (aligned) together in a cluster. Clusters containing more than 200 ESTs were discarded since they probably represented repetitive elements or highly expressed genes such as those for ribosomal proteins. If two or more clusters shared common ESTs, those clusters were grouped together into a "supercluster," resulting in 9,974 breast superclusters.

20 Records for the 479 human cDNA libraries represented in the GenBank release were downloaded to create a database of these cDNA library records. These 479 cDNA libraries were grouped into three groups, Plus (normal breast and breast tumor libraries, and breast cell lines, in which expression was desired), Minus (libraries from other normal adult tissues, in which expression was not desirable), and Other
25 (fetal tissue, infant tissue, ovary, pregnant uterus, male-specific tissues, non-breast tumors and cell lines other than breast cell lines, in which expression was considered to be irrelevant). A summary of these library groups is presented in Table I.

TABLE I**BREAST CDNA LIBRARIES**

Library	# of Libraries
Plus	11
Normal	6
Tumor	4
Cell lines	1
Minus	171
Other	297

Each supercluster was analyzed in terms of the ESTs within the supercluster. The tissue source of each EST clone was noted and used to classify the superclusters into four groups: Type 1 - EST clones derived from the Plus group libraries only; no EST clones derived from Minus or Other group libraries; Type 2 - EST clones derived from the Plus and Other group libraries only; no EST clones derived from the Minus group; Type 3 - EST clones derived from the Plus, Minus and Other group libraries, but the number of EST clones derived from the Plus group is higher than from either the Minus or Other groups; and Type 4 - EST clones derived from Plus, Minus and Other group libraries, but the number of clones derived from the Plus group is higher than the number from the Minus group. For each type, subcategories were generated for clusters containing one or two clones. Types 1a, 2a, 3a and 4a represent Type 1, 2, 3 and 4 clusters where the number of clones in the Plus group is 2. Types 1b, 2b, 3b and 4b represent Type 1, 2, 3 and 4 clusters where the number of clones in the Plus group is 1. This analysis identified 3230 breast clusters. From these clusters, 2501 EST clones were ordered from Research Genetics, Inc. (Huntsville, AL), and were received as frozen glycerol stocks in 96-well plates. A summary of the clusters generated and clones ordered is shown in Table II.

TABLE II**BREAST CLUSTER SUMMARY**

Type	# of Superclusters	# of ESTs Ordered
1	50	49
1a (2/0/0)	130	130
1b (1/0/0)	65	65
2	167	162
2a (2/0/n)	312	321
2a (1/0/n)	1875	1752
3	56	22
3a	32	0
4	333	0
4a	210	0
Total	3230	2501

The EST clone inserts were PCR-amplified for Synteni microarray analysis using amino-linked PCR primers. When more than one PCR product was obtained for a particular clone, that clone was not used for expression analysis. In total, 1896 clones from the electronic subtraction method were analyzed by microarray analysis to identify electronic subtraction breast clones that had high tumor vs. normal tissue mRNA. Such screens were performed using a Synteni (Palo Alto, CA) microarray, according to the manufacturer's instructions (and essentially as described by Schena et al., *Proc. Natl. Acad. Sci. USA* 93:10614-10619, 1996 and Heller et al., *Proc. Natl. Acad. Sci. USA* 94:2150-2155, 1997). Within these analyses, the clones were arrayed on the chip, which was then probed with fluorescent probes generated from normal and tumor breast cDNA, as well as various other normal tissues. The slides were scanned and the fluorescence intensity was measured.

Clones with an expression ratio greater than 2 (i.e., the level in breast tumor cDNA was at least twice the level in normal breast cDNA) were identified as

breast tumor-specific sequences (Table III). The sequences of these clones are provided in SEQ ID NOs:1-124.

TABLE III

5

BREAST-TUMOR SPECIFIC CLONES

SEQ ID NO.	Sequence Designation
1	19702
2	19703
3	B1003C
4	B1002C
5	19708
6	19709
7	19710
8	19711
9	B1006C
10	B1007C
11	19714
12	19715
13	19716
14	19717
15	19718
16	19719
17	19720
18	19721
19	19997
20	19998
21	19999
22	20000
23	20001
24	20002
25	20005
26	20006
27	20007
28	20008
29	20009
30	20010
31	20011
32	20012
33	20013
34	20014

35	20079
36	20080
37	20081
38	20082
39	20083
40	20085
41	20086
42	20087
43	20088
44	20089
45	20090
46	20091
47	20092
48	20093
49	20115
50	20116
51	20118
52	20119
53	20120
54	20121
55	20122
56	20123
57	20124
58	20125
59	20360
60	20361
61	20362
62	22180
63	22181
64	22182
65	22183
66	22185
67	22186
68	22188
69	22189
70	22190
71	22191
72	22192
73	22193
74	22194
75	22196
76	22197
77	22198
78	22199
79	22200
80	22201

81	22202
82	22204
83	22206
84	22207
85	22208
86	22209
87	22210
88	22211
89	22212
90	22213
91	22214
92	22215
93	22216
94	22217
95	22218
96	22219
97	22220
98	22221
99	22222
100	22223
101	22224
102	22225
103	22226
104	22227
105	22228
106	22229
107	22230
108	22231
109	22232
110	22233
111	22234
112	22235
113	22236
114	22237
115	22238
116	22239
117	22240
118	22241
119	22242
120	22243
121	22244
122	22245
123	22334
124	22335

The B1002C sequence (SEQ ID NO:4; 517 bp) was used in a BlastN search of the GenBank Human EST database to identify overlapping sequences that extended further toward the 5' end of the corresponding gene. Two human EST clones were identified in this manner and were purchased from Genome Systems (St. Louis, MO) and sequenced. The resulting sequence information gave rise to a partial predicted open reading frame of 214 amino acids, which is 88% identical over 180 amino acids to the mouse "iroquois homeobox protein 3" (GenBank protein accession #Y15001). The extended B1002C sequence, along with the predicted open reading frame, are provided herein as SEQ ID NOs:125 and 126, respectively. The alignment between B1002C and the mouse iroquois homeobox protein 3 is presented in Figure 1.

EXAMPLE 2

SYNTHESIS OF POLYPEPTIDES

15

Polypeptides may be synthesized on a Perkin Elmer/Applied Biosystems Division 430A peptide synthesizer using Fmoc chemistry with HPTU (O-Benzotriazole-N,N,N',N'-tetramethyluronium hexafluorophosphate) activation. A Gly-Cys-Gly sequence may be attached to the amino terminus of the peptide to provide a method of conjugation, binding to an immobilized surface, or labeling of the peptide. Cleavage of the peptides from the solid support may be carried out using the following cleavage mixture: trifluoroacetic acid:ethanedithiol:thioanisole:water:phenol (40:1:2:2:3). After cleaving for 2 hours, the peptides may be precipitated in cold methyl-t-butyl-ether. The peptide pellets may then be dissolved in water containing 0.1% trifluoroacetic acid (TFA) and lyophilized prior to purification by C18 reverse phase HPLC. A gradient of 0%-60% acetonitrile (containing 0.1% TFA) in water (containing 0.1% TFA) may be used to elute the peptides. Following lyophilization of the pure fractions, the peptides may be characterized using electrospray or other types of mass spectrometry and by amino acid analysis.

30

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

CLAIMS

What is claimed:

1. An isolated polypeptide, comprising at least an immunogenic portion of a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:
 - (a) sequences recited in SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125;
 - (b) sequences that hybridize to a sequence recited in any one of SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125, under moderately stringent conditions; and
 - (c) complements of sequences of (a) or (b).
2. An isolated polypeptide according to claim 1, wherein the polypeptide comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125, or a complement of any of the foregoing polynucleotide sequences.
3. An isolated polypeptide comprising a sequence recited in SEQ ID NO:126.
4. An isolated polynucleotide encoding at least 15 amino acid residues of a breast tumor protein, or a variant thereof that differs in one or more substitutions, deletions, additions and/or insertions such that the ability of the variant to react with antigen-specific antisera is not substantially diminished, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide comprising a sequence recited in any one of SEQ

ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125, or a complement of any of the foregoing sequences.

5. An isolated polynucleotide encoding a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide comprising a sequence recited in any one of SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125, or a complement of any of the foregoing sequences.

6. An isolated polynucleotide, comprising a sequence recited in any one of SEQ ID NOs: 1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125.

7. An isolated polynucleotide, comprising a sequence that hybridizes to a sequence recited in any one of SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125, under moderately stringent conditions.

8. An isolated polynucleotide complementary to a polynucleotide according to any one of claims 4-7.

9. An expression vector, comprising a polynucleotide according to any one of claims claim 4-8.

10. A host cell transformed or transfected with an expression vector according to claim 9.

11. An isolated antibody, or antigen-binding fragment thereof, that specifically binds to a breast tumor protein that comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125, or a complement of any of the foregoing polynucleotide sequences.

12. A fusion protein, comprising at least one polypeptide according to claim 1.

13. A fusion protein according to claim 12, wherein the fusion protein comprises an expression enhancer that increases expression of the fusion protein in a host cell transfected with a polynucleotide encoding the fusion protein.

14. A fusion protein according to claim 12, wherein the fusion protein comprises a T helper epitope that is not present within the polypeptide of claim 1.

15. A fusion protein according to claim 12, wherein the fusion protein comprises an affinity tag.

16. An isolated polynucleotide encoding a fusion protein according to claim 12.

17. A pharmaceutical composition, comprising a physiologically acceptable carrier and at least one component selected from the group consisting of:

- (a) a polypeptide according to claim 1;
- (b) a polynucleotide according to claim 4;
- (c) an antibody according to claim 11;
- (d) a fusion protein according to claim 12; and
- (e) a polynucleotide according to claim 16.

18. A vaccine comprising an immunostimulant and at least one component selected from the group consisting of:

- (a) a polypeptide according to claim 1;
- (b) a polynucleotide according to claim 4;
- (c) an antibody according to claim 11;
- (d) a fusion protein according to claim 12; and
- (e) a polynucleotide according to claim 16.

19. A vaccine according to claim 18, wherein the immunostimulant is an adjuvant.

20. A vaccine according to any claim 18, wherein the immunostimulant induces a predominantly Type I response.

21. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a pharmaceutical composition according to claim 17.

22. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a vaccine according to claim 18.

23. A pharmaceutical composition comprising an antigen-presenting cell that expresses a polypeptide according to claim 1, in combination with a pharmaceutically acceptable carrier or excipient.

24. A pharmaceutical composition according to claim 23, wherein the antigen presenting cell is a dendritic cell or a macrophage.

25. A vaccine comprising an antigen-presenting cell that expresses a polypeptide comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

- (a) sequences recited in SEQ ID NOs:1-125;
- (b) sequences that hybridize to a sequence recited in any one of SEQ ID NOs:1-125 under moderately stringent conditions; and
- (c) complements of sequences of (i) or (ii);
in combination with an immunostimulant.

26. A vaccine according to claim 25, wherein the immunostimulant is an adjuvant.

27. A vaccine according to claim 25, wherein the immunostimulant induces a predominantly Type I response.

28. A vaccine according to claim 25, wherein the antigen-presenting cell is a dendritic cell.

29. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of an antigen-presenting cell that expresses a polypeptide comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

- (a) sequences recited in SEQ ID NOs:1-125;
- (b) sequences that hybridize to a sequence recited in any one of SEQ ID NOs:1-125 under moderately stringent conditions; and
- (c) complements of sequences of (i) or (ii) encoded by a polynucleotide recited in any one of SEQ ID NOs:1-125;

and thereby inhibiting the development of a cancer in the patient.

30. A method according to claim 29, wherein the antigen-presenting cell is a dendritic cell.

31. A method according to any one of claims 21, 22 and 29, wherein the cancer is breast cancer.

32. A method for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

- (i) polynucleotides recited in any one of SEQ ID NOs:1-125; and
- (ii) complements of the foregoing polynucleotides;

wherein the step of contacting is performed under conditions and for a time sufficient to permit the removal of cells expressing the antigen from the sample.

33. A method according to claim 32, wherein the biological sample is blood or a fraction thereof.

34. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated according to the method of claim 32.

35. A method for stimulating and/or expanding T cells specific for a breast tumor protein, comprising contacting T cells with at least one component selected from the group consisting of:

(a) polypeptides comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

(i) sequences recited in SEQ ID NOs:1-125;
(ii) sequences that hybridize to a sequence recited in any one of SEQ ID NOs:1-125 under moderately stringent conditions; and

(iii) complements of sequences of (i) or (ii);

(b) polynucleotides encoding a polypeptide of (a); and

(c) antigen presenting cells that express a polypeptide of (a);

under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells.

36. An isolated T cell population, comprising T cells prepared according to the method of claim 35.

37. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a T cell population according to claim 36.

38. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

(a) incubating CD4⁺ and/or CD8⁺ T cells isolated from a patient with at least one component selected from the group consisting of:

(i) polypeptides comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

(1) sequences recited in SEQ ID NOs:1-125;

(2) sequences that hybridize to a sequence recited in any one of SEQ ID NOs:1-125 under moderately stringent conditions; and

- (3) complements of sequences of (1) or (2);
 - (ii) polynucleotides encoding a polypeptide of (i); and
 - (iii) antigen presenting cells that expresses a polypeptide of (i);
- such that T cells proliferate; and
- (b) administering to the patient an effective amount of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient.

39. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

- (a) incubating CD4⁺ and/or CD8⁺ T cells isolated from a patient with at least one component selected from the group consisting of:
 - (i) polypeptides comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:
 - (1) sequences recited in SEQ ID NOs:1-125;
 - (2) sequences that hybridize to a sequence recited in any one of SEQ ID NOs:1-125 under moderately stringent conditions; and
 - (3) complements of sequences of (1) or (2);
 - (ii) polynucleotides encoding a polypeptide of (i); and
 - (iii) antigen presenting cells that express a polypeptide of (i);
- such that T cells proliferate;
- (b) cloning at least one proliferated cell to provide cloned T cells; and
- (c) administering to the patient an effective amount of the cloned T cells, and thereby inhibiting the development of a cancer in the patient.

40. A method for determining the presence or absence of a cancer in a patient, comprising the steps of:

- (a) contacting a biological sample obtained from a patient with a binding agent that binds to a breast tumor protein, wherein the tumor protein comprises an amino acid

sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs:1-125 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of polypeptide that binds to the binding agent; and

(c) comparing the amount of polypeptide to a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient.

41. A method according to claim 40, wherein the binding agent is an antibody.

42. A method according to claim 43, wherein the antibody is a monoclonal antibody.

43. A method according to claim 40, wherein the cancer is breast cancer.

44. A method for monitoring the progression of a cancer in a patient, comprising the steps of:

(a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs:1-125 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of polypeptide that binds to the binding agent;

(c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and

(d) comparing the amount of polypeptide detected in step (c) to the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

45. A method according to claim 44, wherein the binding agent is an antibody.

46. A method according to claim 45, wherein the antibody is a monoclonal antibody.

47. A method according to claim 44, wherein the cancer is a breast cancer.

48. A method for determining the presence or absence of a cancer in a patient, comprising the steps of:

(a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NO:1-125 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; and

(c) comparing the amount of polynucleotide that hybridizes to the oligonucleotide to a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient.

49. A method according to claim 48, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a polymerase chain reaction.

50. A method according to claim 48, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a hybridization assay.

51. A method for monitoring the progression of a cancer in a patient, comprising the steps of:

(a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide

sequence recited in any one of SEQ ID NO:1-125 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide;

(c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and

(d) comparing the amount of polynucleotide detected in step (c) to the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

52. A method according to claim 51, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a polymerase chain reaction.

53. A method according to claim 51, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a hybridization assay.

54. A diagnostic kit, comprising:

(a) one or more antibodies according to claim 11; and

(b) a detection reagent comprising a reporter group.

55. A kit according to claim 54, wherein the antibodies are immobilized on a solid support.

56. A kit according to claim 54, wherein the detection reagent comprises an anti-immunoglobulin, protein G, protein A or lectin.

57. A kit according to claim 54, wherein the reporter group is selected from the group consisting of radioisotopes, fluorescent groups, luminescent groups, enzymes, biotin and dye particles.

58. An oligonucleotide comprising 10 to 40 contiguous nucleotides that hybridize under moderately stringent conditions to a polynucleotide that encodes a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125, or a complement of any of the foregoing polynucleotides.

59. A oligonucleotide according to claim 58, wherein the oligonucleotide comprises 10-40 contiguous nucleotides recited in any one of SEQ ID NOs:1, 3, 4, 8, 9, 11, 12, 14, 15, 17, 26, 30, 31, 35, 36, 47, 52, 55, 60, 62, 63, 66, 69, 71, 72, 74-78, 81, 83-87, 92, 94, 97, 99-104, 107, 108, 110, 112, 113, 117, 118, 120-122 or 125.

60. A diagnostic kit, comprising:
(a) an oligonucleotide according to claim 59; and
(b) a diagnostic reagent for use in a polymerase chain reaction or hybridization assay.

1
1 M S F P Q L G Y Q Y I R P L Y P P E R F G A A G G G S S A G G R S C F G A G A S E L A A S G S B1002Conf. PRO
iroquois. PRO

1
51 L S N V L S S V Y G A P Y A A A A A A A A A A Q G Y G A F L P Y A T E L P I F F Q L G A Q Y E L K D B1002Conf. PRO
iroquois. PRO

1
101 S P G Y Q H P A T A A A A F F H P H P A F Y P Y Q Q Y Q F G D P S R F K N A T R E S T S T L K A W L N B1002Conf. PRO
iroquois. PRO

1
151 E H R K N P Y P T R G E K I M L A I I T K H T L T Q V S T W F A H A R R R L K K E N K M T W A P R S B1002Conf. PRO
iroquois. PRO

1
201 R T D E E G N A Y G S E R E E E D E E E D F E E S K R E L E H E E E L A G R G G H G G R G A G R B1002Conf. PRO
iroquois. PRO

1
R R R D E E I D L E N L D S A A G S E L T L A G A A H R N G D F G L G P I S D C K T S D S D S S B1002Conf. PRO
iroquois. PRO

11 F E R G A G R G S S G G R E E B G A R R R E F F G L A F R R V V V O S L D P C A P A T A F A S A L O K F B1002Conf. PRO
iroquois. PRO

301 E G L E D R P L S V L S L A P P P P P P V A R A P A S P S F S F S S L D P C A P A F A P S S A L Q K E B1002Conf. PRO
iroquois. PRO

61 K I W S L A E T A T S P D N P R R S F P P G A G G S F F G A A V A F S A L U L S P A A A A A A A A A A B1002Conf. PRO
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351 K I W S L A E T A T S P D N P R R S F P P G A G G S F F G A A V A P F T L Q L S P A A A A A A A A A A B1002Conf. PRO
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109 P I V S A P L G K F P A W T N R P F P P G P F P G F R F H P L S L L G S A F P H L L G L F G A A G H F B1002Conf. PRO
iroquois. PRO

401 R I V S A P L G K F P A W T N R P F P P G P F A G F R F H P L S L L G S A P O H L L G L F G A A G H F B1002Conf. PRO
iroquois. PRO

179 A A A A A - F A R P A F F R G T D R C S A L E V E K K L L P T A F O P V P P P O H L L G A A A L V B1002Conf. PRO
iroquois. PRO

451 A A A A A Y A R P A E P E S G T D R C S A L E V E K K L L P T A F O P V P P P O H L L G A A A L V B1002Conf. PRO
iroquois. PRO

200 I S A L S S S B1002Conf. PRO
iroquois. PRO

501 L S A L S S S B1002Conf. PRO
iroquois. PRO

FIG 1

SEQUENCE LISTING

<110> Corixa Corporation
Mitcham, Jennifer L.
Jiang, Yuqiu

<120> COMPOSITIONS AND METHODS FOR THE THERAPY
AND DIAGNOSIS OF BREAST CANCER

<130> 210121.482PC

<140> PCT

<141> 2000-06-23

<160> 127

<170> FastSEQ for Windows Version 3.0

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<213> Homo sapien

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gccattccac tgttttctgg cctttatgat ttctgctgag aaatccactg ttagtctgat	300
ggggtctccc ttcatagcac caatgacctg aagagccttg ttgaaggaag actccatctg	360
atgactcaga gcaagtattt tttagtgtgt tattgttatt agcaagaaag agggccataa	420
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<213> Homo sapien

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tcttagcttc ctgttctaataaatgcacgg ctttaccttt cctgtcagaa ataaaccaag	360
gctctaaaag atgatttccc ttctgtaact ccctagagcc acaggttctc attccttttc	420
ccattatact tctcacaatt cagtttctat gagtttgatc acctgatttt ttttaacaaa	480
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gagtagcttc acaaagatag gagataaccg agaaactgtt tttattgcat ctctctgtgt	240
aggccacctt gtaagctgtg gcctcagact taccttttat cttgtgtaat gcttttttac	300
cactattgtg tctgctgctt atgtcttttc caggcatgta ctgttgcagg ggtgtccaat	360
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<213> Homo sapien

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<213> Homo sapien

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<211> 777

<212> DNA

<213> Homo sapien

<400> 7

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gaacctgcgg	gagtcggcca	ccatcacgtg	cctgggtgacg	ggcttctctc	ccgcggacgt	360
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<213> Homo sapien

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cattgttaac	gggccttatt	ctgatctcac	ctgtcgcgta	gctctaatat	tcacataaac	960
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<400> 9

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<400> 11

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<212> DNA

<213> Homo sapien

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<211> 500

<212> DNA

<213> Homo sapien

<400> 16

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cataaagaaa	gttttatattg	tattttacta	ctgtttggct	aattttgata	tataactggc	180
tacaaacaga	gccttactat	ttattagtgg	ggaaatgatt	ttaagaccgt	ccttttcagt	240
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gctgtctcat	ttaaaactgt	tggatatagc	tggatcctgg	ataggaaaat	gaaattattt	360
tttcattgtg	ttttttaatt	gggtgatcc	aaagctggca	ccttcaggca	cattgggtctc	420
atagccatta	ctgtttttat	tgcccttcta	agatcctgtc	ttcagctggg	tcagagaaaa	480
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<210> 17

<211> 732

<212> DNA

<213> Homo sapien

<400> 17

tactgatctc	tgttctagt	agtgggtcaa	aaagcatatg	catccttata	cgtcaactca	60
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aaatgacttg	ttcctgggga	atgccttcaa	ccaaaacaca	gctagtattt	ctatgccccca	180
aatccaatcc	cagtctttca	tgatccatgc	cggcggttgg	gtggggaggg	gaatcattgg	240
ttgggggaag	ggaggaaacc	ccacctccag	cccccgccac	cgggctccct	gggcacccag	300
caagatctgg	ggctgcagag	aacagaagag	ctgggtgcact	taatccagct	ctgcccttgg	360
ggggaggagg	acctgtgtgt	caggctctgc	catgggaacg	agtgtaaacc	gtggctgtct	420
cctgcagtga	gccaccgcgg	caggcacggt	gactgtttta	ctgacatcac	tcaaaagcta	480
aagcaataac	attctcctgc	gttgctgagt	cagctgttca	tttgtccgcc	agctcctgga	540
ctggatgtgt	gaaaggcatc	acatttccat	tttctccgt	gtaaatgttt	tatgtgttcg	600
cctactgata	ccattcgttg	cttctattgt	aaatatattg	catttgtatt	tattatctct	660
gtgttttccc	cctaaggcat	aaaatggttt	actgtgttca	tttgaacca	aaaaaaaaaa	720
aaaaaaaaaa	aa					732

<210> 18

<211> 103

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(103)

<223> n = A,T,C or G

<400> 18

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gatcggtttt	tcactgaccc	ggtaaggcgg	gggggcgagc	ccc		103

<210> 19

<211> 586

<212> DNA

<213> Homo sapien

<400> 19

acgcctgcag	gtaccgggtcc	ggaattcccg	ggtcgactac	tactactaaa	ttcgcgggccg	60
cgtcgactcc	cagcccctat	cgagaaaacc	atctccaaag	ccaaagggca	gccccgagaa	120
ccacaggtgt	acaccctgcc	ccatcccg	gaggagatga	ccaagaacca	ggtcagcctg	180
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cagccggaga	acaactacaa	gaccacgcct	cccgtgctgg	actccgacgg	ctccttcttc	300
ctctatagca	agctcacctg	ggacaagagc	aggtggcagc	aggggaacgt	cttctcatgc	360
tccgtgatgc	atgaggetct	gcacaaccac	tacacgcaga	agagcctctc	cctgtccccg	420
ggtaaatgag	tgccgacggc	ggcaagcccc	cgtccccgg	gctctcgcg	tcgcacgagg	480
atgcttgga	cgtaccccg	ctacatactt	cccaggcacc	cagcatggaa	ataaagcacc	540
caccactgcc	ctggaaaaaa	gtcgacgcgg	ccgcgaattt	agtagt		586

<210> 20

<211> 646

<212> DNA

<213> Homo sapien

<400> 20

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cttcgcgggc	gtagacttt	tgtttctcg	agtctgctt	gctcagcgtc	aggggtgctg	180
tgaggctgta	ggtgctgtcc	ttgctgtcct	gctctgtgac	actctcctgg	gagttaccgc	240

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gcaggcacac	aacagaggca	gttccagatt	tcaactgctc	atcagatggc	gggaagatga	360
agacagatgg	tgcagccaca	gttcgtttga	tctccagctt	ggccccctgg	ccaaaagtgt	420
acggaggtga	gctgtcgacg	cgcccgcgaa	tttagtagta	gtaggcggcc	gctctagagg	480
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caattcactg	gcggtcgttt	tacaacgtcg	tgactgggaa	aaccctggcg	ttacccaact	600
taatcgctt	gcagcacatc	cccccttcgc	cagctggcgt	aatagc		646

<210> 21
 <211> 625
 <212> DNA
 <213> Homo sapien

<400> 21						
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atacaaagta	acatttttagt	acagaaaatc	ccagtctgtc	agctcagtag	ctgtctgtgc	120
acactgtacc	atctcagtc	cactctgcct	gtaacttaaa	aaacagcccc	tacccccaga	180
gggtctgcga	gttaatacct	tgagaatagt	ctacagtttt	tcatagtttg	tctgagctag	240
aaaacttgta	cctgtaaaac	aaaggacagc	attgaggact	gaaacttgct	tytttttttg	300
aacaactgtg	caagaaaata	tatccctttt	taaaaaacat	cagttatggc	taaactacaa	360
tctagtgtct	agaattacaa	agaataaaat	gaaatcaaag	atttctcgct	agtaaaatga	420
aatgttagga	acagtattaa	aatataggtc	ctaccccaac	gacacttaca	cagagcccag	480
tacagtacct	attattaaca	ggacgcatag	cttaaggagg	aaccacatca	aatcttcagc	540
cagacatata	tagcctcaga	agtgcaaaaa	aaaaaaaaag	cccccaaacg	aagacacccc	600
actgagtagg	gtgcatgccc	tgagt				625

<210> 22
 <211> 576
 <212> DNA
 <213> Homo sapien

<400> 22						
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tgtggacctc	tgggaaggaa	agaatatggc	ctgtgtgcag	cggacgctga	tgaatctggg	120
tgggctggca	gtagcccgag	atgatgggct	cttctctggg	gatcccaact	ggttccctaa	180
gaaatccaag	gagaatcctc	ggaacttctc	ggataaccag	ctgcaagagg	gcaagaacgt	240
gatcgggtta	cagatgggca	ccaaccgcgg	ggcgtctcag	gcaggcatga	ctggctacgg	300
gatgccacgc	cagatcctct	gatcccaccc	caggccttgc	ccttgccttc	ccacgaatgg	360
ttaatata	tgtagatata	tatttttagca	gtgacattcc	cagagagccc	cagagctctc	420
aagctccttt	ctgtcagggg	gggggggttca	gcctgtcctg	tcacctctga	ggtgcctgct	480
ggcactcctc	cccccatgct	tactaataca	ttcccttccc	catagccatc	aaaactggac	540
caactggcct	cttcctttcc	cctgggacca	aaattt			576

<210> 23
 <211> 416
 <212> DNA
 <213> Homo sapien

<400> 23						
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cgggggactc	tcttccagcc	tccgaccgcc	ctccgatttc	ctctccgctt	gcaacctccg	120
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ctccttcttg	ccaaccaacc	atgagctccc	agattcgtca	gaattattcc	accgacgtgg	240
aggcagccgt	caacagcctg	gtcaatttgt	acctgcaggc	ctcctacacc	tacctctctc	300
tgggcttcta	tttcggaccg	cgatgatgtg	gctctggaag	gcgtgagcca	cttcttccgc	360

gaactggccg aggagaagcg cgagggctac gagegtctcc tgaagatgca aaacca 416

<210> 24

<211> 1037

<212> DNA

<213> Homo sapien

<400> 24

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tcgagtcttg	tcgcggcatc	tccattaagg	atgtgttttg	cctgaggcgg	gaaatcccca	120
ttcccagcac	aggctctgtg	gagatggctg	tcgcagtgtt	cctgggtcatc	tgcatgtgtg	180
tgggtggtagc	catcttgggt	tactgcttct	tcaagaacca	gagaaaggac	ttccacggac	240
accaccacca	cccaccacc	acccttgcca	gctccactgt	ctccactacc	gaggacacgg	300
agcacctggt	ctataaccac	accaccggc	ccctctgagc	ctgggtctca	ccggctctca	360
cctggccctg	cttctctgct	gccaaggcag	aggcctgggc	tgggaaaaac	tttggaaacca	420
gactcttgcc	tgtttcccag	gcccactgtg	cctcagagac	cagggctcca	gccccctctg	480
gagaagtctc	agctaagctc	acgtcctgag	aaagctcaaa	ggtttggaag	gagcagaaaa	540
cccttggggc	agaagtacca	gactagatgg	acctgcctgc	ataggagttt	ggaggaagtt	600
ggagttttgt	ttcctctgtt	caaagctgcc	tgtccctacc	ccatggtgct	aggaagagga	660
gtgggggtggt	gtcagaccct	ggaggcccca	accctgtcct	cccagctcc	tyttccatgc	720
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ggcctagacc	agcagacaga	gccaggagag	gstcagctgc	attccgcagc	ccccaccccc	960
aagggtttcc	aacatcacag	cccagcccac	ccactgggta	ataaaagtgg	tttgggaaaa	1020
aaaaaaaaaa	aaaaaaaa					1037

<210> 25

<211> 1144

<212> DNA

<213> Homo sapien

<400> 25

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gaccgtcagg	gagaaaatgg	ccakatgatg	agctgcacat	gtcttgaggaa	cggaaaagga	120
gaattcaagt	gtgaccctca	tgaggcaacg	tggtacgatg	atgggaagac	ataccacgta	180
ggagaacagt	ggcagaagga	atatctcggt	gccatttgct	cctgcacatg	ctttggaggc	240
cagcggggct	ggcgctgtga	caactgccgc	agacctgggg	gtgaacccag	tcccgaaggc	300
actactggcc	agtcctacaa	ccagtattct	cagagatacc	atcagagAAC	aaacactaat	360
gttaattgcc	caattgagtg	cttcattgct	ttagatgtac	aggctgacag	agaagattcc	420
cgagagtaaa	tcattctttcc	aatccagagg	aacaagcatg	tctctctgcc	aagatccatc	480
taaaactggag	tgatgttagc	agaccagct	tagagttctt	ctttctttct	taagcccttt	540
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gagtttcctg	agggttttct	cataaatgag	ggctgcacat	tgctgtttct	gcttcgaagt	660
attcaatacc	gctcagtatt	ttaaatgaag	tgattctaa	atgtgtttg	ggatcaatag	720
gaaagcatat	gcagccaacc	aagatgcaaa	tggttttgaaa	tgatatgacc	aaaatttttaa	780
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gaacaagcat	gatcttggtta	ctgtgatatt	ttaaatatcc	mcagtactca	ctttttccaa	900
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tttggtataa	ttatggtggg	tgaaaaat	ttatactgta	tgtgccaaa	cttttaaaact	1080
gtggaaagac	aactgtttta	ataaaagatt	tacattcccc	aacttgaaaa	aaaaaaaaaa	1140
aaaa						1144

<210> 26

<211> 488

<212> DNA

<213> Homo sapien

<400> 26

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gtactcttgg	gagtttccat	cttitytggtc	tctgcccaga	atccgacaac	agctgctcca	120
gctgacacgt	atccagctac	tggtcctgct	gatgatgaag	cccctgatgc	tgaaaccact	180
gctgctgcaa	ccactgcgac	cactgctgct	cctaccactg	caaccaccgc	tgcttctacc	240
actgctcgta	aagacattcc	agttttaccc	aaatgggttg	gggatctccc	gaatggtaga	300
gtgtgtccct	gagatggaat	cagcttgagt	cttctgcaat	tggtcacaac	tattcatgct	360
tcctgtgatt	tcattccaact	acttaccttg	cctacgatat	cccctttatc	tctaatacagt	420
ttatttitytt	tcaaataaaa	aataactatg	agcaacataa	aaaaaaaaaa	aaaaaaaaaa	480
aaaaaaaaa						488

<210> 27

<211> 764

<212> DNA

<213> Homo sapien

<400> 27

agccccaggt	gagattgcag	ctgggtgattt	attgagaaac	ttccaggttg	agtaggcttc	60
attcagggca	tgtctctccc	ccaggcccca	tcttcataca	cttccgctaa	gatgcccaca	120
attcatcccc	cttcatgtgc	agcttgaaaa	acctgacat	tggttgctcc	tgccggggcat	180
acctgccatt	gccactgccc	atctctgtga	tgaggaaatc	aacaccagat	ctctggcagc	240
tcttgagggga	ttggccagtg	gccattctca	ggggtgctgt	ctagttttcc	cagagccccg	300
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gagtcagtgg	atggacaggt	ggtttcttcc	cacaagagag	aaatytaagt	gtctattgca	480
ggagaaacgt	cccttgccac	tccccactct	catcaggcca	agtggaggac	tgggccagagg	540
gcctgcacat	gcaaactcca	gtccctgcct	tcagagagct	gaaaagggtc	cctcgggtctt	600
ttatttcagg	gctttgcatg	cgctctatcc	cccctctgcc	tctcccccmm	ttytttggag	660
caaggagatg	cagctgtatt	gtgtaacaag	ctcattttgta	cagtgtctgt	tcattgtaata	720
aagaattact	tttccttttg	caaaaaaaaa	aaaaaaaaaa	aaaa		764

<210> 28

<211> 802

<212> DNA

<213> Homo sapien

<400> 28

accatcactt	gccgggcaag	tcagagtatt	aacaactatt	taaattggta	tctgcagaaa	60
ccagggaaag	cccctaacct	cctgatctat	gctgcattcca	ctttgcaaag	tgggggtccca	120
tcaaggttca	stggcagtg	atctgggaca	gatttcactc	tcaccatcag	cagtctgcaa	180
cctgaagatt	ttgcaactta	ctactgycaa	cagagttaca	ctaccccatc	cactttcrgm	240
cctgggacca	aagtggatgt	caaacgaact	gtggctgcac	catctgtctt	catcttcccg	300
ccatctgatg	agcagwtgaa	atctggaaact	gcctctgttg	tgtgcctgct	gaataacttc	360
tatcccagag	aggccaaagt	acagtggaa	gtggataacg	ccctccaatc	gggtaactcc	420
caggagagt	tcacagagca	ggacagcaag	gacagcacct	acagcctcag	cagcaccctg	480
acgctgagca	aagcagacta	cgagaaacac	aaagtctacg	cctgsgaagt	cacccatcag	540
ggcctgagct	cgcccgtcac	aaagagcttc	aacaggggag	agtgttagag	ggagaagtgc	600
ccccacctgc	tcctcagttc	cagcctgacc	ccctcccatc	ctttggccty	tgaccctttt	660
tccacagggg	acctaccctt	attgcgggtcc	tccagctcat	ctttcacctc	acccccctcc	720
tcctccttgg	ctttaattat	gctaattgtg	gaggagaatg	aataaataaa	gtgaatcttt	780
gcaaaaaaaaa	aaaaaaaaaa	aa				802

<210> 29

<211> 620

<212> DNA

<213> Homo sapien

<400> 29

ctcgagccag aattcggtcg aggagaactg gtataggagc tccaaggaca agaaacacgt	60
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ttccaaggaa atggctaccc aacttgcctt catgcgcctg ctggccaact atgcctctca	180
gaacatcacc taccactgca agaacagcat tgcatacatg gatgaggaga ctggcaacct	240
gaaaaaggct gtcattctac agggctctaa tgatgttgaa cttgttgctg agggcaacag	300
caggttcact tacactgttc ttgtagatgg ctgctctaaa aagacaaatg aatggggaaa	360
gacaatcatt gaatacaaaa caaataagcc atcacgcctg cccttccttg atattgcacc	420
tttggacatc ggtggtgctg accaggaatt ctttgtggac attggcccag tctgtttcaa	480
ataaatgaac tcaatctaaa ttaaaaaaga aagaaatttg aaaaaacttt ctctttgcca	540
tttcttcttc ttctttttta actgaaagct gaatccttcc atttctctg cacatctact	600
tgcttaaatt gtgggcaaaa	620

<210> 30

<211> 644

<212> DNA

<213> Homo sapien

<400> 30

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aaccacaaaa tattttctct gaaatctaca cttagtttta aaacagagat gggattttgc	180
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tacagctttg cattgkcacc ccatcactgc cagggaacca gcccaggca tacacagatg	300
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gggctaaata tctttaaaag tataactctg gacaatgtac ttagggacct actacttact	480
caaaataggg tagtagcatt aataactaaa tagggcgtga gtcatgagaa agattccctg	540
catctcttgc caagtattaa cactagtttt actaataagc aacggggtag ataactttga	600
agattgcac aacttcaaca caaaaaaatg tgaatatccc tcgt	644

<210> 31

<211> 674

<212> DNA

<213> Homo sapien

<400> 31

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aaagggagag ctgtaagctt cactctgtcc tacaccggag aaaagcagga ataactttac	180
cgtggaaata atgttttagct ttatcagag aaaattgtcc ttctagagca tagagtccca	240
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cagcatatca ctttttcttt ttgtgectca ggttcctcag ctgtaaaatt gaaaaatata	360
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agctgtaaat atcaactagg cagccaatag tgttgactat gaagatgcaa aactattact	600
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caccaaattc cgta	674

<210> 32
 <211> 713
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1) ... (713)
 <223> n = A,T,C or G

<400> 32

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ccttgaaaag	aaactattaa	aaaggataga	cattttctgat	gtaagtaaaa	cccccaagca	180
acttaatatc	catctgtcag	tcacaaactt	ttcccctaga	tttttttttt	aactagttct	240
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tttaggaaat	aaatatgcta	taaacaaggt	acagtacagt	gagagttaac	aatcctagga	420
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ttccatttaa	ctattacaga	aagcagtaac	tattgcaact	atctaatagt	tcacacaaaa	540
gaattaaaag	cttaaaaaata	atcttttagga	aacacaatat	tcaaaatcta	aacacactga	600
taaattatta	agattaagat	tatwtatgtg	ataaatgaaa	tctcctacca	atccatccag	660
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<210> 33
 <211> 698
 <212> DNA
 <213> Homo sapien

<400> 33

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tgccagcccc	acacagctac	agctttcttg	ctcccctcag	ccccagccc	ctccccatc	180
tcccaccctg	tacctcatcc	catgagaccc	tggtgcctgg	ctctttctgc	acccttggac	240
aagacaaacc	aagtcggaac	agcagataac	aatgcagcaa	ggccctgctg	cccaatctcc	300
atctgtcaac	aggggcgtga	ggtcccagga	agtggccaaa	agctagacag	atccccgttc	360
ctgacatcac	agcagcctcc	aacacaaggc	tccaagacct	aggttcctgg	acgagatggg	420
aaggcacagg	gagaagggat	aaccctacac	ccagacccca	ggctggacat	gctgactgtc	480
ctctcccctc	cagccttttg	ccttggcttt	tctagcctat	ttacctgcag	gctgagccac	540
tctcttccct	ttccccagca	tcactcccca	aggaagagcc	aatgttttcc	accataatc	600
ctttctgccc	acccttagtt	ccctctgctc	agccaagcta	gwtatcagct	ttcagggcca	660
tggttcacat	tagaataaaa	ggtagtaatt	agaaaaaa			698

<210> 34
 <211> 605
 <212> DNA
 <213> Homo sapien

<400> 34

aaggtgccat	cattcttgag	gaggaagtag	cgtggccgcc	aggtcttgat	gtactcccct	60
cgtttgtgca	gccaaacctc	cttcacaata	gccacgtcgc	tcattggtgcc	cgaggctccc	120
gagacgetca	cgcgtcctc	tcaggctggc	gtccccagag	cccagctggc	ctggccacag	180
cctctgatgc	accagctgac	aggetgcctc	ctccaggcag	cccctttgac	ttcttttgacc	240
caggctggct	cggccttccc	taagccctg	gtgacagatg	gccccgtttg	ctctccctgt	300

ccatggtgtt	cctacccatg	gatcctggga	cagggcacag	ggctcctccc	tgctccccag	360
actaggaaag	caaagaaatt	caaacatgag	gaagacagga	ccaggatgca	ggccactggc	420
gcaaacggga	gtccagagcc	ctccagcgca	agcccaaaaa	cctcctggga	gaaaccccag	480
gcccctccta	aaccacagcg	cccctgccgg	tctgaatctg	gttcattcat	ttggccaaca	540
tgtaccgggc	gtctcaggtt	ttgccaggcc	cagtgtctgg	cgctggagac	aaagagcctc	600
gtrcc						605

<210> 35
<211> 753
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (753)
<223> n = A,T,C or G

<400> 35						
atggagacgg	ctctgcagca	gtacactcta	gaaccatcag	aaaaaccttt	tggttctcaag	60
tctgtgcccc	tggccacggc	gcccattggc	gagcagagaa	cagaaagtac	ccccatcaca	120
scagtcaaac	agcctgagaa	agtggcagct	accaggcagg	agatcttcca	ggagcagttg	180
gcagcagtg	cagagttccg	cggtcttggg	cccctcttca	agtcctcgcc	tgagcccgtg	240
gccctcaccg	agtcagagac	ggagtatgtc	atccgctgca	ccaaacacac	cttcaccaac	300
cacatggttt	ttcagtttga	ctgcacaaac	acactcaatg	accagacctt	ggagaatgtc	360
acagtgcaga	tggagcccac	tgaggcctat	gaggtgctct	gttacgtgcc	tgcccggagc	420
ctgccctaca	accagcccgg	gacctgctac	acactgggtg	cactgcccac	agaagacccc	480
acagctgtgg	cctgcacatt	cagctgcatg	atgaagttca	ctgtcaagga	ctgtgatccc	540
accactgggg	agactgatga	cgaaggctat	gaggatgagt	atgtgctgga	agatctggaa	600
gttactgtag	ctgatcacat	tcaaaaaggtc	atgaaactga	acttcgaagc	agcctgggat	660
gangtagggg	atgaatttga	gaaggaggaa	acgttcacct	tgtctaccat	caagacactt	720
gaagaggctg	ggggnaatat	tgggaagtct	tgg			753

<210> 36
<211> 433
<212> DNA
<213> Homo sapien

<400> 36						
ggggcttgca	atcatttgaa	ttgttctgtt	tcacaataaa	ggagattcac	tgggttctgc	60
attttcagga	ttcaatagaa	ctgcttcatt	aaaaaaataa	tccttagcaa	gcattcgaat	120
cctaactgct	ttgatgcact	tgccctcggg	cacctgtcat	ttccaatatg	gtaggtgtca	180
aagtcaaaa	tatttactgg	gagaaaaaag	agaggagtgg	ttgtagaagt	ctccctaaat	240
cagacatgtc	aagcaatcag	ccaacgtggg	gtattttctc	ttcaatatat	tagtgtgaat	300
tgagacactg	agataaagac	atcgtgcaga	gataaatggg	gatacagtta	aatgtagcaa	360
ctcttgagtt	cattttttcc	cactgtagca	aaattaatgc	tttctcttta	ttgaaataaa	420
ttgctcatte	ctc					433

<210> 37
<211> 601
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (601)

<223> n = A,T,C or G

<400> 37

aggagtgtccc	cggtgtcccc	tcaccctgtg	gcaagtacat	ctcctgcgcc	gagtgcctga	60
agttcgaaaa	gggccccttt	gggaagaact	gcagcgcggc	gtgtccgggc	ctgcagctgt	120
cgaacaaccc	cgtgaagggc	aggacctgca	aggagaggga	ctcagagggc	tgctgggtgg	180
cctacacgct	ggagcagcag	gacgggatgg	accgctacct	catctatgtg	gatgagagcc	240
gagagtgtgt	ggcaggcccc	aacatcgccg	ccatcgtcgg	gggcaccgtg	gcaggcatcg	300
tgctgatcgg	cattctcctg	ctgggtcatct	ggaaggctct	gatccacctg	agegacctcc	360
gggagtacag	gcgctttgag	aaggagaagc	tcaagtccca	gtggaacaat	gataatcccc	420
ttttcaagag	cgccaccacg	acgggtcatga	accccaagtt	tgctgagagt	taggagcact	480
tggtgaaaac	aanggccgct	aggaccacc	atgtctgccc	atacgcgggc	ganacatggc	540
ttggcacact	nttgangatg	taccatttac	caaaatcnan	tttttcccc	tnaatnanct	600
g						601

<210> 38

<211> 713

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1) ... (713)

<223> n = A,T,C or G

<400> 38

gcttccgcct	attgattctg	ctcattttata	caataatgag	gagcaggttg	gactggccat	60
ccgaagcaag	attgcagatg	gcaktgtgaa	gagagaagac	atattctaca	cttcaaagct	120
ttggtgcaat	tcccatcgac	cagagttggt	ccgaccagcc	ttggaaaggt	cactgaaaaa	180
tcttcaattg	gattatgttg	acctctacct	tattcatttt	ccagtgtctg	taaagccagg	240
tgaggaagtg	atcccaaaag	atgaaaatgg	aaaaatacta	tttgacacag	tggatctctg	300
tgccacgtgg	gaggccgtgg	agaagtgtaa	agatgcagga	ttggccaagt	ccatcggggt	360
gtccaacttc	aaccgcaggc	agctggagat	gatcctcaac	aagccagggc	tcaagtacaa	420
gcctgtctgc	aaccaggtgg	aatgtcatcc	ttacttcaac	cagagaaaaa	tgctggattt	480
ctgcaagtca	aaagacattg	ttctgggttg	ctatagtgtc	ctgggatccc	accgagaaga	540
accatgggtg	gacccgaact	ccccggtgct	cttggaggac	ccagtccttt	gtgccttggc	600
aaaaaagcac	aagcgaaccc	cagccctgat	tgccctgcgc	taccactaca	gcgtgggggt	660
gtggtcctgg	ccaagagcta	caatgagcag	cgcacacnac	agaacgtgca	ggg	713

<210> 39

<211> 451

<212> DNA

<213> Homo sapien

<400> 39

gaaagaactg	actgaaacgt	ttgagatgaa	gaaagttctc	ctcctgatca	cagccatctt	60
ggcagtggtc	gttggtttcc	cagtctctca	agaccaggaa	cgagaaaaaa	gaagtatcag	120
tcgacagcga	tgaattagct	tcagggtttt	ttgtgttccc	ttacccatat	ccatttcgcc	180
cacttccacc	aattccattt	ccaagatttc	catggtttag	acgtaatttt	cctattccaa	240
tacctgaatc	tgcccctaca	actccccttc	ctagcgaaaa	gtaaacaaga	aggaaaagtc	300
acgataaacc	tggtcacctg	aaattgaaat	tgagccactt	ccttgaagaa	tcaaaattcc	360
tgtaataaaa	agaaaaacaa	atgtaattga	aatagcacac	agcattctct	agtcaatatc	420
tttagtgatc	ttctttaata	aacttgaaag	c			451

<210> 40

<211> 778
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(778)
 <223> n = A,T,C or G

<400> 40

accgcaccc	agccgccc	gcacacaa	gaggtgggt	aggaaatcca	gagttgccat	60
ggagaaaatt	ccagtgtcag	cattcttgct	ccttggtggc	ctctccctaca	ctctggccag	120
asataccaca	gtcaaacctg	gagccaaaaa	ggacacaaag	gactctcgac	ccaaactgcc	180
ccagaccctc	tccagagggt	ggggtgacca	actcatctgg	actcagacat	atgaagaagc	240
tctatataaa	tccaagacaa	gcaacaaacc	cttgatgatt	attcatcact	tggatgagtg	300
cccacacagt	caagctttaa	agaaagtgtt	tgctgaaaat	aaagaaatcc	agaaattggc	360
agagcagttt	gtcctcctca	atctggttta	tgaaacaact	gacaaacacc	tttctcctga	420
tggccagtat	gtcccagga	ttatgtttgt	tgacccatct	ctgacagtta	gagccgatat	480
cactggaaga	tattcaaate	gtctctatgc	ttacgaacct	gcagatcagc	tctgttgctt	540
gacaacatga	agaaagctct	caagttgctg	aagactgaat	tgtaaagaaa	aaaaatctcc	600
aagcccttct	gtctgtcagg	ccttgagact	tgaaaccaga	anaagtgtga	naaagactgg	660
ctagtggggg	aagcattagt	ggaacacact	ggattanggt	tatgggtttt	aatggttacc	720
aacaacctat	ttttttttta	naaaaaaacc	aaagttttta	aaaaaaattt	ngggtttt	778

<210> 41
 <211> 696
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(696)
 <223> n = A,T,C or G

<400> 41

agtgatgccg	gcagatcatc	tggtttcagc	gggaacnagc	atcttgcacg	tggctctggca	60
tggaccncag	ggctgtgggg	acttggggta	cagtaatcaa	gtaatccccc	tttccagaat	120
gcattaaccc	actccctttg	acctcacgat	ggggcagggtc	cccaagtgtg	caagctcagt	180
attcatgatg	gtgggggatg	gagtgctctc	cgagggttctt	gggggaaaaa	atattgtagc	240
atattttaagg	gaggcaatga	accctctccc	ccnctctctc	cctgcccaca	tctgtctcct	300
agaatcttat	gtgctgtgaa	taatagacct	tcwctgcccc	tccagttttt	atagacstga	360
ggttccagtg	tctcctggta	actggaacct	ytctgagggg	ggaatcctgg	tgctcaaatt	420
accctccaaa	agcaagtagc	caaagccgtt	gccaaacccc	accataaat	caatggggccc	480
tttatttatg	mcgactttat	ttattctaat	atgattttat	agtatttata	tatattgggt	540
cgtctgcttc	ccttgatatt	ttmttccttt	ttttgtaata	ttgaaaasgg	sgatataatt	600
attataagta	gactataata	tatttagtaa	tatatattat	taccttaaaa	gtctattttt	660
gtgttttggg	cattttttaa	taaacartct	gagtggt			696

<210> 42
 <211> 509
 <212> DNA
 <213> Homo sapien

<400> 42

ctcaggtgga	aaaggaggga	gctactctca	ggctgcgtgc	agcgacagtg	cccagggctc	60
------------	------------	------------	------------	------------	------------	----

tgatgtgtct	ctcacagctt	gaaaagcctg	agacagctgt	cttgtgaggg	actgagatgc	120
acgattttct	cacgcctccc	ctttgtgact	tcaagagcct	ctggcatctc	tttctgcaaa	180
ggcacctgaa	tgtgtctgcg	tccctgttag	cataatgtga	ggaggtggag	agacagccca	240
cccttgtgtc	cactgtgacc	cctgttccca	tgctgacctg	tgtttcctcc	ccagtcatct	300
ttcttgttcc	acaagaggtg	gggctggatg	tctccatctc	tgtctcaact	ttacgtgcac	360
tgagctgcaa	cttcttactt	ccctactgaa	aataagaatc	tgaatataaa	tttgttttct	420
caaataattg	ctatgagagg	ttgatggatt	aattaaataa	gtcaattcct	ggaatttgga	480
gagagcaaat	aaagacctga	gaaccttcc				509

<210> 43

<211> 388

<212> DNA

<213> Homo sapien

<400> 43

gcaggatttc	ttcactcctc	ccctttgtga	cttcaagagc	ctctggcatc	tctttctgca	60
aaggcacctg	aatgtgtctg	cgtccctgtt	agcctaattg	gaggaggtgg	agagacagcc	120
cacccttgtg	tccactgtga	cccctgttcc	catgctgacc	tgtgtttcct	ccccagtcac	180
ctttcttgtt	ccagagaggt	gggctggatg	gtctccatct	ctgtctcaac	tttatgtgca	240
ctgagctgca	acttcttact	tccctactga	aaataagaat	ctgaatatac	atttgttttc	300
tcaaawattt	gctatgagag	gttgatggat	taattaaata	agtcaattcc	tggaatgtga	360
gagagcaaat	aaagacctga	gaaccttc				388

<210> 44

<211> 565

<212> DNA

<213> Homo sapien

<400> 44

ggaaaagagg	gttccctctc	ttgtgwtgtg	ctcttccccc	cacctctaata	tcttctgctc	60
tgtttgggaa	gacgtggagg	aaaaggtgac	ttctgcccc	accgtctcta	ccccaccaa	120
ctctcgcgtg	ttggagagaa	ggggccctcc	cagcacaag	ttgcattcct	ccccctaata	180
ttattctaat	ttattaactt	tgaccacccc	ttcttgagcc	tgcagccttc	ccgtgtggcc	240
tgagggctgt	cgagtgtgct	gccccagccc	ctcccagccc	ttgcccagcc	tgggggagtg	300
gggaaggctt	gggcatggcc	ccgttggagg	ttgatttgc	gttttgtttc	ttgtctttgt	360
gttctgtggt	acttgctgag	agaaaagaaa	agtgtgacaa	gcagaaggag	gtgggaaaac	420
ggacccaaac	cccagtgtgc	cctgccccga	tgcctttcct	ttagtgggtg	gaaacctta	480
tctygcaaag	tgaatgtgtc	cccttcccca	cccthtagt	tatttcacag	aaaacaaaac	540
ctcccaataa	aacggttgaa	gcctg				565

<210> 45

<211> 661

<212> DNA

<213> Homo sapien

<400> 45

gttgaggaga	tgggatgtcc	cagatgatag	ggctcctggg	atttcagacc	caataccagc	60
aggactccag	tcacctctac	cccakttctc	caggacacag	cgctcccaac	tctgagtgtg	120
gtcccacctc	tggctccttg	agcacaacca	acgtgggaat	cacaccctcc	agacctccca	180
cagctccacc	ccagactggg	cgcgggccc	gcctccattt	cagctgtgac	aacctcagag	240
ccgtgttggc	ccaagcatga	caaggacgta	tgaaaacttc	cagtacttgg	agaataaggt	300
gaaagtccag	gggttttaaaa	atgggcccact	tcctctccag	tcctcctctg	agcgtctccg	360
ctctgggccc	tgccatctcc	tgetgtccct	gggcctcggc	ctgtgtgtgc	tggteatcat	420
ctgtgtgggt	ggattccaaa	attccaaatt	tcagagggac	ctggtgaccc	tgagaacaga	480
ttttagcaac	ttcacctcaa	acactgtggc	ggagatccag	gcactgactt	cccagggcag	540

cagcttggaa gaaacgatag catctctgaa agctgagggtg gagggtttca agcaggaacg 600
gcaggcagtt cattctgaaa tgctcctgcg agtccagcag ctgggtgcaag acctgaagaa 660
a 661

<210> 46
<211> 453
<212> DNA
<213> Homo sapien

<400> 46
tgctctgtgt gtgtgcctgc tgaataactt ctatcccaga gaggccaaag tacagtggaa 60
ggtggataac gccctccaat cgggtatctc ccaggagagt gtcacagagc aggacagcaa 120
ggacagcacc tacagcctca gcagcaccct gacgctgagc aaagcagact acgagaaaca 180
caaagtctac gcctgcgaag tcacccatca gggcctgagc tcgcccgtca caaagagctt 240
caacagggga gagtggttaga gggagaagtg cccccacctg ctctcagtt ccagcctgac 300
cccctcccat cctttggcct ctgacccttt ttccacaggg gacctacccc tattgcggtc 360
ctccagctca tctttcacct cccccccctc ctctccttg gctttaatta tgctaattgt 420
ggaggagaat gaataaataa agtgaatctt tgc 453

<210> 47
<211> 687
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1) ... (687)
<223> n = A,T,C or G

<400> 47
cttcggctaa aatgcagaaa tgcattgctgt cagcgttggt atttcacatt caatggagct 60
gaatgttsag gacctcttc cattgaagct ataatttatt tggaccaagg aagccctgaa 120
atgaattcaa caattaatat tcatcgact tctctgtgg aaggactttg tgaaggaatt 180
ggtgctggat tagtggatgt tgctatctgg gttggcactt gttcagatta cccaaaagga 240
gatgcttcta ctggatggaa ttcagtttct cgcattatta ttgaagaact accaaaataa 300
atgctttaat tttcatttgc tacctctttt tttattatgc cttggaatgg ttcacttaaa 360
tgacatttta aataagttta tgtatacatc tgaatgaaaa gcaaagctaa atatgtttac 420
agaccaaagt gtgatttcac actgttttta aatctagcat tattcatttt gcttcaatca 480
aaagtgggtt caatattttt ttagttgggt tagaataact tcttcatagt cacattctct 540
caacctataa tttggaatat tgttgggtgt tttgtttttt ctcttagtat agcattttta 600
aaaaaatata aaagctacca atctttgtca atttgtaa atgttaanaat tttttttata 660
tctggtnaaa taaaaaatta ttttccc 687

<210> 48
<211> 944
<212> DNA
<213> Homo sapien

<400> 48
ggcagagga gcatggacat gaggggtccc gctcagctcc tggggctcct gctactctgg 60
ctccgaggtg ccagatgtga catecagatg acccagctct catcttccct gtctgcttct 120
gtaggagaca gagtcaccat cacttgccgg gcaagtcaga acattaacaa cttcttaatt 180
tggtatcagc aaaaaccagg gagacccctc cagctcctga tctatcttac atctaatttg 240
caaagtgggg tcccatctag gttcagtggt agtggatctt ggacagattt cactctcacc 300
atcacgggtc tgcagcctga agattttgca acttactact gtcaacagag tcacagtacc 360

ccgtacacct	tgggccaggg	gaccaagctg	gagatcaaac	gaactgtggc	tgcaccatct	420
gtcttcatct	tcccgccatc	tgatgagcag	ttgaaatctg	gaactgcctc	tgttgtgtgc	480
ctgctgaata	acttctatcc	cagagaggcc	aaagtacagt	ggaagggtga	taacgccctc	540
caatcgggta	actcccagga	gagtgtcaca	gagcaggaca	gcaaggacag	cacctacagc	600
ctcagcagca	ccctgacgct	gagcaaagca	gactacgaga	aacacaaact	ctacgcctgc	660
gaagtcaccc	atcagggcct	gagctctccc	gtcacaaaga	gcttcaacag	gggagagtgt	720
tagagggaga	agtgccecca	cctgctcctc	agttccagcc	tgacccctc	ccatcctttg	780
gcctctgacc	ctttttccac	aggggacctc	cccctattgc	ggtcctccag	ctcatcttcc	840
acctcacccc	cctcctcctc	cttggcttta	attatgctaa	tgttggagga	gaatgaataa	900
ataaagtga	tctttgcacc	tgtaaaaaaa	aaaaaaaaaa	aaaa		944

<210> 49

<211> 449

<212> DNA

<213> Homo sapien

<400> 49

ggcacgaggg	cttcgaggaa	ccagccctga	tacctgtgac	cctctctgta	cgctaccctc	60
ctgaagtgtc	catctccggc	tatgatgaca	actggtacct	cggccgtact	gatgccaccc	120
tgagctgtga	cgtccgcagc	aaccagagc	ccacgggcta	tgactggagc	acgacctcag	180
gcaccttccc	gacctccgca	gtggcccagg	gctcccagct	ggtcatccac	gcagtggaca	240
gtctgttcaa	taccaccttc	gtctgcacag	tcaccaatgc	cgtgggcatg	ggccgcgctg	300
agcaggtcat	ctttgtccga	gaaaccccc	gggcctcgcc	ccgagatgtg	ggcccgcctg	360
tgtggggggc	cgtggggggg	acactgctgg	tgctgctgct	tctggctggg	gggtccttgg	420
ccttcatect	gctgaggggtg	aggaggagg				449

<210> 50

<211> 703

<212> DNA

<213> Homo sapien

<400> 50

ggcacgaggg	tgagcccatg	ccccacccaa	ccctgccagc	tgagcaaagg	acagtcttac	60
agcgtcaatg	tcaccttcac	cagcaatatt	cagtctaaaa	gcagcaaggc	cgtggtgcat	120
ggcatcctga	tgggcgtccc	agttcccttt	ctcattcctg	agcctgatgg	ttgtaagagt	180
ggaattaaact	gccctatcca	aaaagacaag	acctatagct	acctgaataa	actaccagtg	240
aaaagcgaat	atccctctat	aaaactgggtg	gtggagtggc	aacttcagga	tgacaaaaac	300
caaagtctct	tctgctggga	aatcccagta	cagatcgttt	ctcatctcta	agtgcctcat	360
tgagtccggt	gcattctggc	aatgagtctg	ctgagactct	tgacagcacc	tccagctctg	420
ctgcttcaac	aacagtgact	tgctctccaa	tggtatccag	tgattcgttg	aagaggagggt	480
gctctgtagc	agaaactgag	ctccgggtgg	ctggttctca	gtggttgtct	catgtctctt	540
tttctgtctt	aggtgggttc	attaaatgca	gcacttgggt	agcagatgtt	taattttttt	600
tttaacaaca	ttaacttgtg	gcctctttct	acacctggaa	atttactctt	gaataaataa	660
aaactcgttt	gtcttgtcaa	aaaaaaaaaa	aaaaaaaaaa	aaa		703

<210> 51

<211> 963

<212> DNA

<213> Homo sapien

<400> 51

caatgaggct	ccctgctcag	ctcctggggc	tgctaattgt	ctgggtctct	ggatccagtg	60
gggatattgt	gatgactcag	tctccactct	cccagcccgt	cacccttggg	gagccggcct	120
ccatctcctg	caagtctagt	cagagcctcc	tacatagtaa	tggatacagt	tatttggatt	180
ggtacctgca	gaagccagga	cagtctccac	agctcctgat	ctatttggct	tctaagcggg	240

cctccgggggt	ccctgacagg	ttcagtgga	gtggatcagg	cacagatttt	acactgagaa	300
tcagtagagt	ggaggctgag	gatgttggt	tttattactg	catgcaaact	ctacagaccc	360
cgctcacttt	cggcggaggg	accaaggtgg	agatcaaacg	aactgtggct	gcaccatctg	420
tcttcactct	cccgccatct	gatgagcagt	tgaaatctgg	aactgcctct	gttgtgtgcc	480
tgctgaataa	cttctatccc	agagaggcca	aagtacagtg	gaagggtggat	aacgccctcc	540
aatcgggtaa	ctcccaggag	agtgtcacag	agcaggacag	caaggacagc	acctacagcc	600
tcagcagcac	cctgacgctg	agcaaagcag	actacgagaa	acacaaactc	tacgcctgcg	660
aagtcaccca	tcagggcctg	agctcgcccc	tcacaaagag	cttcaacagg	ggagagtgtt	720
agagggagaa	gtgccccccc	ctgctcctca	gttccagcct	gacccccctc	catcctttgg	780
cctctgaccc	tttttccaca	ggggacctac	ccctattggg	gtcctccagb	tcwtttttcc	840
cctccccccc	ctcctcctcc	ttggbttaa	ttatgytaak	gttggrggar	aatgaataaa	900
taaagtgawt	ttttgmaaaa	aaaaaaaaaa	aaaaaaaaaa	aaaaagaaaa	aaaaaaaaaa	960
aaa						963

<210> 52

<211> 628

<212> DNA

<213> Homo sapien

<400> 52

cctttctctc	tcctcatttc	ggtgcatgtc	ctttctgcag	ctgcctttca	gcacaggtgg	60
ttccactggg	ggcagctaac	gctgagtgac	aaggatggga	agccacaggt	gcattttact	120
caagtcttct	ctagtcaatg	aggggcaccc	agtgtctctg	gggcaggctg	ggtgggtggc	180
ccctaggtat	cagcctctct	tactgtactc	tcggggaatg	ttaacctttc	tattttcagc	240
ctgtgccacc	tgtctaggca	agctggcttc	cccattggcc	cctgtgggtc	cacagcagcg	300
tggtgcccc	ccagggccac	cgcttctttc	ttgatcctct	ttccttaaca	gtgacttggg	360
cttgagtctg	gcaaggaacc	ttgcttttag	cttcaccacc	aaggagagag	gttgacatga	420
cctccccgcc	ccctcaccaa	ggctgggaac	agaggggatg	tggtgagagc	caggttcctc	480
tggtccctct	caggtgtgtt	tccactagtc	actactgtct	tctccttgta	gctaataaat	540
caatattctt	cccttgctct	tgggcagtg	agagtgtctg	tggtgtacg	ctgcacctgc	600
ccactgagtt	ggggaaagag	gataatca				628

<210> 53

<211> 598

<212> DNA

<213> Homo sapien

<400> 53

ctccctccct	ttttcccacc	ctccacctct	cacggggcct	gccccagccc	ctaagccccg	60
tcccctaacc	cccagtcctc	actgtcctgt	tttcttaggc	actgagggat	ctgagtaggt	120
ctgggatgga	caggaaaggg	caaagtaggg	ogtgtgggtt	ccctgcccc	gtccggaccg	180
ccgateccag	gtgcgtgtgt	ctctgtctct	cctagccctt	ytytcacaca	tcacattccc	240
atggtggcct	caaraaaggc	ccggaagcgc	caggctggag	ataacagcct	yttgccccgc	300
ggccctgogt	cggccctggg	gtaccatgtg	gccacaactg	ctgtggcccc	ctgtccccaa	360
racacttccc	cttgtctccc	tggttgccty	tcttgcccc	tgtcctgaag	cccagcgaca	420
caraaggggg	tggggcggg	ytatggggar	aaagggagcg	aggtcaragg	agggcatggg	480
ttggcaggg	gggcgtttgg	ggccctytat	gctggctttt	caccccarag	gacacaggca	540
gcttccaaaa	wawatattat	ttcttcacgg	gaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	598

<210> 54

<211> 592

<212> DNA

<213> Homo sapien

<400> 54

gtgttttacac	agtaagatca	agagtcaggc	cctggaattt	ccagaaccag	cccgacatag	60
ttgaggactt	gaaggacctg	atcaccctgt	atgctggaca	agaaccccca	gtcgaggatc	120
gtggtgccgk	aatcaagat	mctggtraag	accatratat	gtaaacgctc	ctttgggaac	180
ccattcgagg	gcascggmg	ggaggaaacg	tcactgtcag	ygcctggaaa	cttgctcacr	240
aagcaaggca	gygaagacaa	cctccagggc	wccgacccgc	cccccggtgg	ggaggaggaa	300
gtgctmttgt	gagaggcagt	ccctgcgtgg	aaagtgtgtg	ggccccccag	ccccggctcc	360
ccgcacgcga	tgcattccat	gcggccggag	gaggccatgg	agccccagta	gctgcctgga	420
tcgctcgacc	tcgcatgcgc	gccgcgtcgc	ctctgggggg	atgctgcacc	gcgtttccat	480
agcagcatgt	cctacggaaa	cccagcacgt	gtgtagagcc	tcgatcgtea	tctgctgggt	540
atttggtttt	tcctttgttg	ttttaaaggg	gacaaaaaaa	aaaaaaaaaa	aa	592

<210> 55

<211> 504

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1) ... (504)

<223> n = A,T,C or G

<400> 55

agcgggacga	gcgckngygg	acttatcggg	gccccggggac	ccgggggcgcg	cggaggggtgc	60
acttcaccct	cctgcccgak	cgctacgagc	cactggagga	gccggcgccg	agcgagcagc	120
ccaggaagag	gtaccggagg	aagctgaaga	agtactggca	agaatgtcgg	gaaggtcate	180
atmaaakgat	gccgctacgt	ggctatcggc	ctgcaaggmt	tcgctgcags	ctactccgcc	240
ccgtttgcgg	tagccaccag	cgtgggtatct	ttcgtgcgct	aatgggagct	gctgtgycag	300
gtgccccag	agtgaacggg	agccccctgt	gtgggaactt	tgtgaatcct	ggagcatctc	360
atacttgaac	acacakcata	tttgggaagag	aaaacatgcc	tttctttgnt	gaatcacatt	420
agtatgatga	gtgagtcatn	cctgcccata	tgctgagctt	ctcacatctc	tcagtcacac	480
gtggacccag	tgggtcaatcc	tgca				504

<210> 56

<211> 749

<212> DNA

<213> Homo sapien

<400> 56

gtaagatata	agtttcaggc	agggtctgaa	ggactggatt	ggccaaacat	cagacctgtc	60
ttccaaggag	accaagtcc	ggctacatcc	cagcctgtgg	ttacagtgc	gacaggccat	120
gtgagccacc	gctgccagca	cagagcgctc	ttccccctgt	agactagtgc	cgtagggagt	180
acctgctgcc	ccagctgact	gtggccccct	ccgtgatcca	tccatctcca	gggagcaaga	240
cagagacgca	ggaatggaaa	gcggagtcc	taacaggatg	aaagtccccc	catcagttcc	300
cccagtacct	ccaagcaagt	agctttccac	atgtgtcaca	gaaatcagag	gagagatggg	360
gttggggagc	ctttggagaa	cgccagtctc	ccaggccccc	tgcattctat	gagtttgcaa	420
tgtcacaacc	tctctgatct	tgtgctcagc	atgattcttt	aatagaagtt	ttattttttc	480
gtgcactctg	ctaatacatg	gggtgagcca	gtggaacagc	gggagacctg	tgctagtttt	540
acagattgcc	tcctaatag	gcggctcaaa	aggaaaccaa	gtgttcagga	gttgtttctg	600
accactgat	ctctactacc	acaaggaaaa	tagtttagga	gaaaccagct	tttactgttt	660
ttgaaaaatt	acagcttcac	cctgtcaagt	taacaaggaa	tgctgtgcc	aataaaagg	720
ttctccaaaa	aaaaaaaaaa	aaaaaaaaaa				749

<210> 57

<211> 673

<212> DNA

<213> Homo sapien

<400> 57

agatgtcatg	acccatcaac	gtctcctctc	attgtccctc	tgctgggaca	aagtgccttg	60
tgtctggctg	ggggacaacc	aagagccccc	aagtgcactt	ccctaagggtc	ctccagtgtc	120
tgaacatcag	cgtgctaagt	cagaaaagg	gcgaggatgc	ttacccgaga	cagatagatg	180
acaccatgtt	ctgcgccggt	gacaaagcag	gtagagactc	ctgccagggt	gattctgggg	240
ggcctgtggt	ctgcaatggc	tccctgcagg	gactcgtgtc	ctggggagat	tacccttgtg	300
cccggcccaa	cagaccgggt	gtctacacga	acctctgcaa	gttcaccaag	tggatccagg	360
aaaccatcca	ggccaactcc	tgagtcattc	caggactcag	cacaccggca	tccccacctg	420
ctgcagggac	agccctgaca	ctccttttcag	acctcattc	cttcccagag	atgttgagaa	480
tgttcatctc	tccagcccct	gaccccatgt	ctcctggact	caggggtctgc	ttccccacac	540
ttgggctgac	cgtgtctctc	tagttgaacc	ctgggaacaa	tttccaaaac	tgtccagggc	600
gggggttgcg	tctcaatctc	cctggggcac	tttcatctc	aagctcaggg	cccatccctt	660
ctctgcagct	ctg					673

<210> 58

<211> 994

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(994)

<223> n = A,T,C or G

<400> 58

gacgggcccg	gtgctgaagg	gcagggaaca	acttgatggt	gctactttga	actgcttttc	60
ttttctcctt	tttgcacaaa	gagtctcatg	tctgatattt	agacatgatg	agctttgtgc	120
aaaaggggag	ctggctactt	ctcgtctctg	ttcatccac	tattattttg	gcacaacagg	180
aagctgttga	aggaggatgt	tcccattctg	gtcagtccta	tgccgataga	gatgtctgga	240
agccagaacc	atgccaaata	tgtgtctgtg	actcaggatc	cgttctctgc	gatgacataa	300
tatgtgacga	tcaagaatta	gactgcccc	accagaaat	tccattttga	gaatgttgtg	360
cagtttgccc	acagcctcca	actgctccta	ctcgcctcc	taatggtcaa	ggacctcaag	420
gcccccaagg	agatccaggc	cctcctggta	ttcctgggag	aaatggtgac	cctgggtattc	480
caggacaacc	agggtccctt	ggttctcctg	gcccccttg	aatctgtgaa	tcatgcccta	540
ctggctcctc	gaactattct	ccccagtatg	attcatatga	tgtcaagtct	ggagtagcag	600
taggaggact	cgcaggctat	cctggaccag	ctggccccc	aggccctccc	ggtccccctg	660
gtacatctgg	tcatectggt	tcccctggat	ctccaggata	ccaaggaccc	cctgggtgaac	720
ctgggcaagc	tggtccttca	ggccctccag	gacctcctgg	tgctataggt	ccatctggtc	780
ctgctggaaa	agatggagaa	tcaggtagac	ccggacgacc	tggagagcga	ggattgcctg	840
gacctccagg	tatcaaagg	ccagctggga	tacctggatt	ccctgggatg	aaaggacaca	900
gaggcttcga	tggacgaaat	ggagaaaagg	gtgaaacagg	tgctcctgga	ttaaaggggtg	960
aaaaggtctt	ccaggcgaaa	aggagcnaca	agtt			994

<210> 59

<211> 639

<212> DNA

<213> Homo sapien

<400> 59

tggtggtggt	ggaggacctg	cactctggcg	cccagcagca	ctgggtccggc	cactctgceg	60
agatctccac	gctggccctc	agccacagt	cccaggctct	ggcctctgcc	tcgggcccga	120
gcagcacgac	cgccattgt	cagatccgcg	tctgggacgt	gtctggcgcc	ctctgccagc	180
atctcatttt	ccccatagc	accaccgtgc	tggccctggc	cttctcacca	gatgacaggc	240

ttcttgtcac	actggggggac	cacgatggcc	gcaccctcgc	cctgtggggc	acggccacct	300
atgacctcgt	gtcctccacc	cgcctcccgg	agccggtgca	tggtgtggcc	ttcaaccctt	360
gggacgccgg	cgagctcacc	tgtgtggggc	agggcaactgt	caccttctgg	ctccttcagc	420
agcgtggggc	agacatcagc	cttcagggtgc	gtcgagagcc	agtcccagag	gcagtggggg	480
ctggagagct	gacctcgctc	tgctacgggg	cacctcccct	gctctattgt	ggcaccagct	540
ctggccaggt	ctgtgtctgg	gacacgcgtg	ccggccgctg	cttcttgtcc	tgggaggcgg	600
atgacggtgg	cattgggctg	ttgctgttct	cgggttctc			639

<210> 60

<211> 470

<212> DNA

<213> Homo sapien

<400> 60

ggcacgaggt	ttttattctt	tgcacaattg	tttcattggt	tgacacttaa	tgactcgtc	60
atgtgcatac	gacagtagca	ttctgaccac	acttgtaacgc	tgtaacctca	tctacttctg	120
atgtttttta	aaaatgactt	ttaacaagga	gagggaaaag	aaaccacta	aattttgctt	180
tgtttccctg	aagaatgtgg	caacactggt	ttgtgatttt	atgtgtgcag	gtcatgcaca	240
cagttttgat	aaagggcagt	aacaagtatt	ggggcctatt	tttttttttt	tccacaaggc	300
attctctaaa	gctatgtgaa	attttctctg	cacctctgta	cagagaatac	acctgcccct	360
gtatatectt	ttttcccctc	ccctcccctc	cagtgggtact	tctactaaat	tgttgtcttg	420
tttttttatt	tttaaataaa	ctgacaaatg	acaaaaaaaa	aaaaaaaaaa		470

<210> 61

<211> 535

<212> DNA

<213> Homo sapien

<400> 61

ggcacgaggg	gttcttgggc	caatttcttg	ctgactcagc	cccactctgt	gtcggagtct	60
ccgggggaaga	cggttaaccat	ctcctgcacc	ggcaacagag	gcagcattgc	cgccaactat	120
gtgcagtggg	accagcagcg	cccgggcagt	gccccacca	ctgtgatcta	tgaagataac	180
caaagaccct	ctgggggtccc	tgatcgggtc	tctggctcca	tcgacagctc	ctccaactca	240
gcctcccctca	ctatctctgg	actgaagcct	gaggacgagg	ctgactacta	ctgtcagtct	300
tatgagagca	gcaatcattg	ggtgttcggc	ggagggacca	agctgaccgt	cctaggtcag	360
cccaaggctg	ccccctcggt	cactctgttc	ccacctcctc	ctgaggagct	tcaagccaac	420
aaggccacac	tggtgtgtct	cataagtga	ttctaccggg	gagccgtgac	agtggcctgg	480
aaggcagata	gcagccccgt	caaggcggga	gtggagacca	ccacaccctc	caaac	535

<210> 62

<211> 696

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(696)

<223> n = A,T,C or G

<400> 62

gccagtatcc	aatgggggaat	cagctctaaa	aggaaccaga	ccaacgtttt	ccagcccntt	60
cattntgggtg	actgagggga	ggaaagaatg	ggagggggta	ttcttgtcta	gtggatggaa	120
aggaaacaca	ctgtcaaatt	actatatctc	cttgggttttc	tattacagta	gaattctcca	180
gccatatttt	tattgtctat	gggggaagtt	ggagatgggtg	accttgatta	gaagtgtctg	240
gaggggggata	aatggagggg	ataagattca	gttgggttttg	gaaaatgtta	aagtcttaaa	300

ataatgcgtc	catctgaaga	atTTTTtcta	aaaccagagt	ttataaaaat	atcactgata	360
cagcctgccc	cctcatttcc	ctgccacagg	agatgtcttg	gactagagac	acttgtttaa	420
taatagcttg	tctctgatat	tcccagtagc	ttccctctgt	gtgaggaaag	gatagaaatg	480
ttcaggacat	catcatacag	gctcctcatc	tacaaagttc	cagtagcagt	gacgcctaca	540
cggaagactt	ggaactgcaa	acaggctggg	gtcacctcag	tgacatctga	cgctgtccaa	600
ccagaagttc	gattttttgtt	ctgggggtga	aggaggaaac	agacgtgtac	taaaggacta	660
aaataatttg	tctatacaaa	aaaaaaaaaa	aaaaaa			696

<210> 63
 <211> 256
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(256)
 <223> n = A,T,C or G

<400> 63						
taatatTTtat	tgaagnctta	ctttgtcctc	tacatagggg	agcttctgtc	agggaaatgtt	60
ggttcttccc	aagaaacact	gattttcttt	caggagact	tcattgtgtc	atttatttcc	120
accacagcag	attttaagaa	attataatat	gtaatatTTg	atatctataa	agagtanatc	180
taacgtgaat	aaattatgaa	gcataactat	gagtacctat	gacccatnac	acatatatat	240
taaaacattt	taaata					256

<210> 64
 <211> 678
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(678)
 <223> n = A,T,C or G

<400> 64						
ggcacgaggg	ctanaagcca	tgcctctttt	ctccgtattc	aacaagccca	ttataacaag	60
tctttcacgc	tcaaccattc	aaacaatccc	aacaatccca	gcctaagaat	ttcctggcat	120
ttgatgaaaa	tggctgtggg	ttttgctatc	tttaagctcc	ctgggaagca	ggatataagc	180
ccagggtctg	caggctcttt	ctagctacct	gctcttgac	atagagtttg	cctcatgggt	240
gcaagaaggc	tgccatagct	ccagatggca	catagacatt	ccaggcagca	ggaaaaagga	300
aggagcactt	aaccgaagga	ggtcagggag	ttggttagtc	tccacctgaa	gaagagagcc	360
atgaaccagc	ttcagttgac	taacgggctc	ctgtgagtg	atctttggac	ttttctggag	420
ggtgaaatct	agatgtggta	tgtgtcttaa	agcagccacc	aactcctccc	attaccttcc	480
aagttagccc	aactacacga	tgagacctct	tccctgccct	tgatctggg	ctgagcctct	540
gacttgctgt	gaccaacaga	atgcagtga	agtgatgccg	atactacct	ccctgcccta	600
gacttgggat	acctgcagct	atattcttcc	attcctcagg	acttgcaaaa	acgggtctca	660
ncatgccttn	ccagagcc					678

<210> 65
 <211> 678
 <212> DNA
 <213> Homo sapien

<220>

<221> misc_feature
 <222> (1)...(678)
 <223> n = A,T,C or G

<400> 65

ggcacgagga	ggacacactc	aggatagaaa	ctttatgtgt	ataggggaatg	tggtacagcc	60
tttagccagg	agtcatactt	catcagacac	cagaggacac	acacagtgt	gtggcttttt	120
cagccattgc	tagataccaa	agtggagaca	ttctgtgtgt	gattatgcat	gagactgtac	180
tggttaagact	tgtatctcca	tccacctgaa	ggagaattgc	tggtctattt	tcaggagccc	240
tgccttctct	cactgtggat	ggtgggttgt	ggaaacccgg	tcaggtaatg	atagtggcag	300
gaggcagtca	aatgcccagg	cagatagggg	tgggtacctg	gtgaaacca	accttaaagc	360
tgaagacagt	ccgggctaaa	tcctcatact	gaattgagaa	cctgtcttcc	catttggtgt	420
gctttctctc	gattgatecc	aaccttcac	ctattttacg	tatacctgcc	ctttccta	480
tggtttttac	actgctgtgc	ccaccttttg	agtgggtgct	ttgcatactt	acaaatcagt	540
caacgtgtat	tcccctattc	tgagcccata	aaagacccan	actcagctgc	agtgaggaga	600
gaaatccct	gctgnngggg	gtgggggacca	ctccttgc	ccctctncac	tganagctgt	660
ctttttgctc	aataaaat					678

<210> 66
 <211> 606
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(606)
 <223> n = A,T,C or G

<400> 66

gcatcaactg	ccgccagccc	ttcatnttnt	ccgcctcttc	ctagacgtgc	tacacctggt	60
tgagttctac	ctggaggaag	ggatcactga	tgaagaagcc	atctccctca	tcgacctgga	120
ggtgctgaga	cccaagcggg	atgacagaca	gctagagatt	gcaaacaaca	gctcccagat	180
tctgcggcta	gtggagacca	aggactccat	cggagatgag	gacccgttca	cagctaagct	240
gagctttgag	caaggtggct	cagagttcgt	gccagtgggt	gtgagccggc	tggtgctgcg	300
ctccatgagc	cgccgggatg	tcctcatcaa	gcgatggccc	ccacccctga	ggtggcaata	360
cttccgctca	ctgctgcttg	acgcctccat	taccatgtgc	ccctcctgct	tccagatgtt	420
ccattntgag	gactatgagt	tgctgggtgct	tcagcatggc	tgctgcccct	actgccgcag	480
gtgcaaggat	gacctggcc	catgaccagc	atcctgggga	cggcctgcac	cctctgcccg	540
ccttggggtc	tgctgggctg	tgaaggagaa	taaagagtta	aactgtcaaa	aaaaaaaaaa	600
aaaaaa						606

<210> 67
 <211> 579
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(579)
 <223> n = A,T,C or G

<400> 67

ggcactnaggc	ccaattatat	gttcatttttg	tatatTTTTT	ggtcgggggga	aaaattgacc	60
tgcagtanaa	aaacctttga	ccattttttat	gtccattgga	tactttcctt	tttatcatct	120
taaaaaaaga	taactagtac	taatcattgt	agtggcctaa	gtgtgattta	actcttgaag	180

tcacaccctc	cgaaagatga	gtagaaacca	gcaccagcac	agcccagatc	ttctctttcc	240
tctccttttc	ctcattttatt	cctaaaggaa	tctgaccatt	ttacgtctct	acggcccaaa	300
aaaagacaaa	aataaaaatt	ccttttttatt	cctgtcaact	ggatggaaac	acaaatttca	360
tggagctgtg	taccatcgaa	gaaacctggt	gtctggcatg	aaattactgt	aaagaacttc	420
ctgtaaaaca	cgttcttttaa	caaactgaaa	tgaaaagcat	tggagcgtct	gaatgaaaga	480
cgtgacctcc	tgctgggact	ctgatgggtct	tcagcattca	ccttcgtgtg	tcttcagtgt	540
ctcattgtca	tccctgcttc	tgtttggtct	tagagtgtt			579

<210> 68
 <211> 258
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(258)
 <223> n = A,T,C or G

<400> 68						
ggcatntggc	cctgaccana	tccttaaccc	tagcgatggc	ttgggagatg	ggggggttga	60
tagcatntc	tttcttggcc	cttccttata	ctaggaaaan	atggttcctc	tccttgtgtg	120
tgtctcttcc	ccccaccct	aattcttctg	ctctgttttg	gaagacgtgg	aggaaaaggt	180
gacttctgcc	cccaccgctc	ttacccccac	tgtantggcc	tttggagatg	ccccacctc	240
ccccccacca	actctcgc					258

<210> 69
 <211> 628
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(628)
 <223> n = A,T,C or G

<400> 69						
ggcacnagga	gaattntttg	gaataaaaact	gactgataaa	actggaaatg	ttgctcgggc	60
tgtaactgg	caggaaagat	ttcagcatct	gaatgagtc	cngcacaact	atttaagaat	120
cactcgtatt	cttaaaaagcc	ttggtgagct	tggatatgaa	agtttttaaat	ctcctcttgt	180
aaaattttatt	cttcatgaag	ctcttgtgga	gaatactatt	cccaatatta	agcagagtgc	240
tctagagtat	tttgtttata	caattagaga	cagaagagaa	aggagaaagc	tcctgcggtt	300
cgcccagaaa	cactacacgc	cttcagagaa	ctttatctgg	ggaccgcctc	gaaaagaaca	360
gtcggaggga	agcaaagccc	agaaaatgtc	ttccccctctc	gcctccagtc	ataacagtca	420
aacttctatg	cacaaaaaag	ccaaggactc	caaaaattcc	tcctcagctg	ttcattttaa	480
tagcaaaaca	gctgaagaca	aaaaagtggc	accaaaagag	cctgtggaag	agacagacag	540
gccacccan	agcccagcaa	tgaagctgcc	aagccaagaa	atacagaaga	aggacagtaa	600
tgctganaac	atgaattctc	aacctgag				628

<210> 70
 <211> 439
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature

<222> (1)...(439)

<223> n = A,T,C or G

<400> 70

ggcagcagggc	gcacacccgc	tgggcggccc	ctgtggagac	cctggaaaac	atcatcgcca	60
ctgtagacac	gaggctgcct	gagttctcag	agctgcaggg	ctgtttccgg	gaggagctca	120
tggaggcctt	gcacctgcac	ctggtgaagg	agtacatcat	ccaactcagc	aaggggccc	180
tggtcctcaa	gacggccgag	cagcagcagc	agctggctgg	gtacatcctg	gccaatgctg	240
acaccatcca	gcacttctgc	acccagcagc	gctccccggc	gacctggctg	cagcctgctc	300
tccctacgct	ggccgagatc	attcgctgc	aggaccccag	tgccatcaag	attgaggtgg	360
ccacttatgc	cacctgctac	cctgacttca	gcaaaggcca	cctgancgct	atcctggcca	420
tcaaggggaa	cctatccaa					439

<210> 71

<211> 328

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(328)

<223> n = A,T,C or G

<400> 71

tnggaactca	ggtttacnca	aactgagggg	gccccagccc	tngtaccncc	cctggttacc	60
caggatccat	ntgccctcan	aaaagngttc	aggtacagca	gctgaggctg	ccctgaggaa	120
tcaagggggc	attaccaagg	ggcaggaaaa	ggatatgtaa	naggnggcct	tcatggtana	180
gcttgaccca	anaactactc	cncattngga	tggcccagac	tgactccatc	ccctgacttt	240
ccctttgact	tcnccctgtt	tgtaaataaa	acaataaaat	ggaaggtgct	gtggacggga	300
aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa				328

<210> 72

<211> 721

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(721)

<223> n = A,T,C or G

<400> 72

ggtttctggt	gctattctgc	caagttatcg	agctcctcct	catgtttcaa	cattccatct	60
tcccgtttct	atcctcgact	ccaaagtaag	ccttcttagc	tccaatcagg	gatgaggggc	120
tcaacctctt	ctgtcctcaa	agaggccaaa	cgcagtgcca	cagtcggtag	ccttcacttt	180
tagatgtcct	attcatgtaa	aaaagaaggt	gccccacca	ggcttacatc	agcaataagc	240
aattctaattg	caacgatggg	gtccacattt	taccccagtg	tgtgcccattg	tatgcctttg	300
tgcccgtgta	attattgtta	gcgccccctt	cacttagagg	ggtgatgata	aactgtggcc	360
accttgatta	caaccacat	ttcctgcttt	ggggagcttc	caagtaacag	gccatttctt	420
acctccctcc	aggaacagtg	ggcactgccc	accacctcgt	gtctgctcat	aggatgacgc	480
tggagatccc	cacacttact	ctacccctct	ggcaaattgg	cattccgggtg	gtgggtttttg	540
tttcttttaa	cacattaaat	aaatgagtat	ataggatgtg	aggggagggg	tgagaacaac	600
tagctgtagc	atgtgtaggc	tatatacttt	accattcgac	ttcttttctt	tttttttttt	660
aaaaaaaaaa	aagtgcctga	ctgggttcaa	gcttcatcat	gaaaaaaaaa	aaaaaaaaaa	720
a						721

<210> 73
 <211> 596
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(596)
 <223> n = A,T,C or G

<400> 73

attgtacaga acagcccagg tagggaggac agntgcccga ggteccatag gantgcatgc	60
ctcaagccca cgtcatgcag agccactcag ctcaccctgn tcagggcacg tggtttacct	120
gcattcccct nttgcaggtc ctacgtgtgg aggacgtacc atntgacctn tgcaggagag	180
aaactcacgg aagacagaaa gaagctccga gactacggca tccggaatcg anacgagggt	240
tccttcatca aaaagctgag gcaaaagtga gcctccagac aggacaaccc tnttcatcac	300
tgggtggctga gctttttccc agcaggaatg ggtcctcgaa tcatcgtgcc tntttcacan	360
aaaggacgtt gtggtggcct caccaccaggc atgcccacaa gtaactgtca gcataaacct	420
gggggcccctc aggactagga caggggtgagc cagtgtctcc tcctttcatg tacttggcct	480
gagactgacc tctccctagg tccaaatgcc ctagtccatc ggcagaccca cggcctggcc	540
cactgtataa aataaacctg tttgcttntt agtttgaaaa aaaaaaaaaa aaaaaa	596

<210> 74
 <211> 302
 <212> DNA
 <213> Homo sapien

<400> 74

ggcagcagggc ttaaaccaag tttctctgca gctctttcgg ttctgcttac agtgtgtggg	60
aaatctgatt tttttcccct agtaatatgt tgataagaaa tttagtgtat tgactgcctc	120
agtgacacaa tttatcttta aagggtgtgga agctgggtggg gaccaaagtgt tacctgtgtt	180
tttgctgttg attgctatctt tcagaagcaa accatgtttt tcacttacag taggagtcaa	240
caaatttggg attttagaag ggggaggagg gagctatctt tgtaagactg ctgtcatatt	300
tg	302

<210> 75
 <211> 635
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(635)
 <223> n = A,T,C or G

<400> 75

ggcagcaggt ttaatgtgac tttcctgagt taacactgaa tagtatttct aaaatttttt	60
gtactttatt ttttaatgta acttggttcta tctatctata tatatatttg atagtgtgtg	120
gaataatatc cccaggtatt ttccatatta aatgctaatt atcttttgat ttctttttca	180
taagcagatc tggcatttat tacagggtctg ccgcttaaga gaactcatta taatgaacgt	240
ttattatatt ttgcagttcc atgcctgttg tccattgatt gacatgagca cccctgtttt	300
ctctggagaa atacctcccc tctctggggt gcttctctgtg gtagtgtctt tcaggatatc	360
gttccactag ctacaggtga gcattttacc cattgttgga taatggtaat ctctttttca	420
gaattttgag tctgtaattc atttgtagat gaaccagaaa atgtgggaac tcattcattc	480

ttgtcccaga	attctgttga	gaacatccat	tcattctggc	taattgatta	caagaataac	540
tgnggatacg	atccctttan	aacctgcttc	tctgatctgn	gtgtttcctc	acttctcaat	600
aaaaatgtct	tttgctaaaa	aaaaaaaaaa	aaaaa			635

<210> 76
<211> 678
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(678)
<223> n = A,T,C or G

<400> 76						
ggcacgaggc	tcttgattct	tggcttgcc	ctcctccaat	tccaaactta	gtgaaatggc	60
cttaagcatt	ttaaactgta	tgtatacatt	agcgcattca	tgcctttcta	aacgcatttc	120
aaatgtcaac	caggaaggca	caccactgta	ttagttttat	actgccgctg	taaaatttac	180
cacaaactta	gtgacttaac	acaaatttat	tgcaattctg	taggetggaa	gtctgactat	240
gggtctcact	ggactagaat	caaggctggc	aggctgcctt	ccttcctgga	ggttctaggg	300
gagactctgt	ctcctgctcc	ttcaggctgc	tggcagaatc	cacatccttt	cggtggcagg	360
gccaaaggcc	ccactttctt	gctgactgta	aactaaggcc	acttcagct	tgtagaggct	420
gcctacattc	cttggtctct	ggccccctcc	tccatcttca	gagctagcag	gttcagtctg	480
tgtcacgaac	catttctctg	gttccctgca	gacaggaaag	gttgtcccta	aggactcatg	540
agattaggtt	gggcccagcc	agataataca	tgataatctc	cctcctcaag	gnttttaata	600
ttaaacacat	ctgcaggaca	cattttgcc	tgtaactaac	attcactgg	ccaggggatt	660
aaggaatgaa	ccctcttt					678

<210> 77
<211> 669
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(669)
<223> n = A,T,C or G

<400> 77						
ggcacgaggg	agaatcttaa	aaaaaaaaaa	acgtttctca	ctgtcttaaa	tagaattttt	60
aaatagtata	tattcagtgg	catttttggag	aacaaagtga	atttacttcg	acttcttaaa	120
tttttgtaaa	agactataag	tttagacatc	tttctcattc	aaatttaaag	atatctttct	180
cctcttgatc	aatctatcaa	tattgataga	agtcacacta	gtatatacca	tttaatacat	240
ttacactttc	ttatttaaga	agatattgaa	tgcaaaataa	ttgacatata	gaactttaca	300
aacatatgtc	caaggactct	aaattgagac	tcttccacat	gtacaatctc	atcatcctga	360
agcctataat	gaagaaaaag	atctagaaac	tgagttgtgg	agctgactct	aatcaaatgt	420
gatgattgga	attagaccat	ttggcctttg	aactttcata	ggaaaaatga	cccaacattt	480
cttagcatga	gtacctcat	ctctagaagc	tgggatggac	ttactattct	tggttatatt	540
ttanatactg	aangngcta	tgtttctgtt	attattccaa	gactggagat	aggcagggct	600
aaaaagggat	tattattttc	ctttaatgat	ggggctaaaa	ttcttcctat	aaaattcctt	660
aaaataagg						669

<210> 78
<211> 134
<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(134)

<223> n = A,T,C or G

<400> 78

ggcacgaggg gtcgatttta atagcgaatc ctttttcttg tagaggtaag taaaatcttc	60
ctgacaaggt tgtcctctnt tcacggcaca gacaatgggc ggncctgttta tgaggggtga	120
gaagngacnc ccgc	134

<210> 79

<211> 396

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(396)

<223> n = A,T,C or G

<400> 79

tanatgcaag tntnttgtng gatatacgta ttgagatatt acncctagtc tgtggcttga	60
ctgttttctt tatgtctttt gatgaatana agttttaaat tttgacaagg tcaaatttat	120
ttttttcttt tgtttgatat tttttntctc caatttaacc ccaagatttc anatattntg	180
ctctattana taaactttat atttttatat ttgtgatctn ccttgaattg atatgtatgt	240
tgtgaattat ggatcagggt tntttttttc ccccatataa gtatccagtc attgtaacnc	300
tgtttattga aanaattatc ctttcctcat taaattncct tgccaattag taaaaaatca	360
attaccata aaaaaaaaaa aaaaaaaaaa aaaaaa	396

<210> 80

<211> 731

<212> DNA

<213> Homo sapien

<400> 80

tctacatcat cctgagagcg ccttcagact ggacagaact gtaggaactg ccgtctctta	60
tggtagatgt ggcccatcac aatatacctg caggtttaaa ctctggacac ggcttattgc	120
agctggagga gtccagggtc aacatgtgga ctcggaagaa gaagccgtcg ggggtgaggt	180
acaggcggtt catcttgtac agcccgctcc gatccaccag caggggtcacc agccggatgc	240
ccgcgcccag cacgtccaca tcgtgcacga tcccgttcac cgcgaattcg ctctcgcaga	300
acgcctcgcg ttctgtccaga tcggaccggc aggcgcgcgc gcagggtca gcgccgggct	360
ccgcgtccct ctgcggcggg ctgaggccga agctgaggct gaagcgcggc ggcggcgcgg	420
gcgcgggtcc cggggccagc gcggcggcgc gcggccgggt ggggtgggcg tggccggcgg	480
agccctcggg ggcgagcgcg ggcggccggg cggcggggaa gcgcagggcc cgtgagcgcg	540
cgcgtcgcgg ggcgtcctcc gacgcgggct cgcggcgcgg cgacgcgcgg ttctcggctt	600
gcgccaggcg cgggcccggc gggccagcgg gcgcgcgtgc ggcctcccgg aagctggcct	660
ggttgtcggc cggcggtggg ggcttcgggc ggcgcgggtc aggccaggcg cggacggcgg	720
gagcgacctc g	731

<210> 81

<211> 396

<212> DNA

<213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(396)
 <223> n = A,T,C or G

<400> 81

cccttcaccc	caatccntga	gaagttcttt	ttctgcttcc	tgcccaacat	natcaacagg	60
acctatttcc	cattttcctg	ntcttnctn	anccagntat	tggnngnggt	tttgaaatgg	120
ctnatnatga	ggaanagtnt	gatoenacac	ttggagganc	naggggtgca	ggtggtnttt	180
tggtgcctta	atgaanagtc	ggattttgaa	gcagccttna	gcgtgggagc	cacnggcgtc	240
ataacggatt	atcccnagc	cctnnggcan	tacctggaca	accatggnc	agctgcccgg	300
acctcctaag	tccagaance	tngaggtntc	ctgtttntnt	tccatgaaaa	ataaatattt	360
gcctttcgat	caaaaaaaaa	aaaaaaaaaa	aaaaaa			396

<210> 82
 <211> 502
 <212> DNA
 <213> Homo sapien

<400> 82

ctcagaggaa	agaagaaaag	ggccaggagt	cagacgtcac	acccgggggc	tcccctttcc	60
catgtagaag	tggtggcatg	cgtcagtctc	ctttacagag	gggtggatgt	atccgtggag	120
gaggggcctt	ctctctttct	aattgcacta	tacttggtct	agcttcagtc	tgagataact	180
taagacctcc	atggggctgt	gatccataga	cttagcaagt	cttgcccttat	ctatggagct	240
cgatgggtgag	aattgtgacc	attgtctgat	gtccatagtt	ccttccccct	agattgtttc	300
tttccacgga	ttgtgttcat	ctgaaccatt	ttatttttta	tttaccaaag	tactgtactt	360
ggctatttgc	agtgttttca	aaaccaaag	tttctttttt	tgtgttttta	atcttcgata	420
cttggtgcaa	tagaagctgc	aaagatgtgc	cactttatct	atgaaatgga	gttttgtata	480
ccaataaatt	ctagtttaaa	aa				502

<210> 83
 <211> 666
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(666)
 <223> n = A,T,C or G

<400> 83

gatttttggt	atttgtactt	atttatgaac	tttacttgag	acagaatatg	gttaaaatta	60
ggaacatcta	ctttgaatga	ggtttatttt	tctattttga	atttgcctta	tgtatattca	120
aaggcttatg	gaaatactgt	aaaggaacat	taggaaaagg	acaaataggc	tataaccatc	180
tatcttaaaa	tcagaccctt	agtataagca	cctctttttc	ttttcctctt	tgacaattta	240
gtctcttatt	taggtccatg	taattaattt	cattccatta	tttttttagct	gtttattcta	300
aaaaacaaaa	attttcagcc	actcccat	attctcccat	gacatggtec	tatatagcag	360
tacttaacac	agtgccttgc	acatagtagg	cattcagtaa	atacttacat	gcatgaatga	420
ataatgtatt	ttcagtgtaa	caaatttatt	ataaaagtgt	agttcgactc	ttcttgggtc	480
tgagatttga	gagtacagaa	ttacagggaa	tgaagagagg	tataagtaga	tatttttaag	540
gaaatgaagt	ataaattaat	aacttgactt	acccctccat	aagttactgc	taactggaga	600
tacccttgta	tgcccaacct	gtaaaggaaa	aagntctatt	tctatattta	taggacattc	660
ttctca						666

<210> 84
 <211> 199
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(199)
 <223> n = A,T,C or G

<400> 84
 aanntactta agttattcaa agaatggtat ctttcttgca agagtaattt aagcacatgg 60
 gaaagattct agactttttg tttcttgcaa canacagtgc cctctgctgc tagaaacctt 120
 tttctactta ctatcatttt tattgtggct tgagctcanc tcaatctggt gcagatgacg 180
 ctggacaact actaaccaa 199

<210> 85
 <211> 670
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(670)
 <223> n = A,T,C or G

<400> 85
 cacttcagga gagaacttca tagcacaatg tctttctata agatattttt aatgatttag 60
 tattttacaa catttggttta ccatattttg atataaccatt tttttctatc tgcccagttt 120
 tattaaaaaa actatatatt attttctaaa gaaacaatca tatttttata caaaattatg 180
 ttttcaggta acgaaataga tgtagggtac agtggaacat aagcagtgtt acccctggct 240
 gggagtcagt attatacaac aaatgggtgag ctggaacatg cctgtctctg gctgtccctc 300
 ctgtgctggg tcgcggatgt gtaggcaaca ttgccttate acgctagggt cacctgacac 360
 tttaaaagga aaaaaagttc catagagtgc tgtggtcaca aaattgtttt gcttttatca 420
 aatactttaa tagaaccaaa gttgcagata ttggaatgta tgggaagtac tcagtctctg 480
 cataagagga ttaaagtatg aaaggatcat ttaatgactg ttttacttat aagtcattaa 540
 gtaatccacc atttcttatg gatgatgctt aagcctgggt aggtttgtac tctaaggagc 600
 ccagatcata atgcagngca tttccttanc ccttagagtt tcttgcaaac atttaaaaaa 660
 agacntattt 670

<210> 86
 <211> 401
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(401)
 <223> n = A,T,C or G

<400> 86
 atcttccac gagtgggatt ctggccttca gagaccagga gggagtgtct gggccgcang 60
 tgtggcactg tggtagagagt gtgtgtcttt gcacacacag tgcagcggga acggtggggc 120
 tggctgggtgc tgaagacaga cacactcctg agccaaggte ttgtcttcaa cctccccgtc 180
 ccgttgtecc attttgctct gtgaagggtgc aaatcccttt cttcccttcc catctcaggc 240

tctcctgttt	tccctcaggg	tccagtatgc	ctttgagctt	tagctgttaa	aaaggaaccc	300
ccgtgacttg	acacagcttt	cacancctggc	tgctangacc	ggcgggctgg	gtgttcacgt	360
gtgtctgtgt	catggatgca	atgcangccc	tnangactg	t		401

<210> 87
 <211> 373
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(373)
 <223> n = A,T,C or G

<400> 87						
cttttttttt	ctttttttta	tttgaaaact	actttancaa	taattaattc	catgattatc	60
acattctgcc	attaagggat	attagtaccg	taatactgaa	gaaattttat	taagtctgaa	120
cttctggggg	aggcagcttc	tttgtttctt	ttctatccac	ccttgctcgg	tgaggatatt	180
gtttcttgac	taataaaccc	tttgatactt	taaccagaaa	tcagctcata	aagctatttt	240
tgagtatagt	tnnggnaaaa	aaaaaagggt	aacttgggna	ataccttcca	nnctgacctc	300
cntntaccaa	gatatttttc	agggctttta	tttactatgc	nctaanacta	tgcncttttt	360
ctgaaatatt	ttt					373

<210> 88
 <211> 507
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(507)
 <223> n = A,T,C or G

<400> 88						
ccaggagcag	cccattcctg	atgctttctc	agagactcct	gcaggcagcc	aggccacang	60
acccttgtgg	tcccacccca	cacacgccag	attctttcct	gaggctgggc	tcccttccca	120
cctctctcac	tccttgaaaa	cactgttctc	tgccctccaa	gaccttctcc	ttcacctttg	180
tccccaccgc	agacaggacc	agggatttcc	atgatgtttt	ccatgagtcc	cctgtttgtt	240
tctgaaaggg	acgctacccg	ggaagggggc	tgggacatgg	gaaaggggaa	gttgtaggca	300
taaagtcagg	ggttcccttt	tttggtctgt	gaaggctcga	gcatgcctgg	atggggctgc	360
accggctggc	ctggcccttc	agggtccctg	gtggcagctc	acctctccct	tggattgtcc	420
ccgaccttg	ccgtctacct	gaggggcctc	ttatgggctg	ggttctaccc	aggtgctagg	480
aacactcctt	cacagatggg	tgcttgg				507

<210> 89
 <211> 796
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(796)
 <223> n = A,T,C or G

<400> 89

gactttaatg	ggnatatggg	ctctacactc	atagggaccc	acatgttcca	agcctccagc	60
tgtgtgttct	gcttcctcaa	gcagcagctg	actctgatat	tccccatctc	accttagcag	120
tctatatcaa	gcaaagggca	tgagaaatcc	catgcaccca	tcatgtatgt	aataaaaggc	180
agatattatg	tgggtctcttg	aaccagtctt	aggcatggag	gttgaggatc	aggagtgact	240
tgaggggtact	gactggcaga	gcaggagccc	cgttatcttg	gacaaacacc	gccactttaa	300
gtttcagctt	cattttttagc	cttctggatt	taaggaaatt	acttttttaa	aaactataag	360
cagccaaaaa	aagcagacag	taaaatgcag	ataaaacagc	tcgggcacag	aggaagggtgg	420
agggaaagtc	tcttgggtaa	ctgccaaact	tcacctcat	acaatgggcc	ccagtaaaac	480
agtgggcctt	aataagcaca	ttcctttccc	tccaggtgca	ctaaaatagg	gaagctaaaa	540
gcagacttgg	ggggatatgcc	tacagctgca	gaaaaatgta	taaaagcaaa	cacaccactc	600
tccctcccat	ataagcacia	caaaaaaaca	cagaagcagt	ccaagcctnt	aagaaactct	660
cccaccctaa	atccttaaac	actcttagtc	tgtagaaaag	actctaacct	aattcagcca	720
gcagccccctc	tcaggtgtgt	ttntntntaaa	ataaacctgt	nttaaccatc	aagccaaaaa	780
aaaaaaaaaa	aaaaaa					796

<210> 90

<211> 462

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1) ... (462)

<223> n = A,T,C or G

<400> 90

tagaacatnc	tgaatttttt	ttgtactgnt	ggactntatt	cagtgtcatg	tcctatatct	60
gatcaagtn	tcaagnagat	aattttanaa	tgaaaaagaa	aatcctcttg	tnggaaacaa	120
aagacgtttt	atatgtgcag	tatgacaaan	aggagtttca	nagacaactt	tgaatccttg	180
tcagcctgga	gaccagcncc	agaggaatnn	ccaaggcaaa	ctcccatata	tttgcttccc	240
ccaaattgct	gcccctacag	actcaaagct	ctttttcttt	gttttggtgt	ttntctaaaa	300
atttactgtt	ntttgtcgat	gctatataag	ccagggagtt	ttaagacgcc	agctntttga	360
natttgntca	ttcccctgta	tttcccacat	anatattaca	tatacccgng	taataaattt	420
atgtttgtta	aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	aa		462

<210> 91

<211> 591

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1) ... (591)

<223> n = A,T,C or G

<400> 91

cgggccttgg	aagcctttga	cctggatcct	gcacagtggg	gagtcaatgt	ccagccctac	60
tccgggtccc	cagccaacct	ggccgtctac	acagcccttc	tgcaacctca	cgaccggatc	120
atggggctgg	acctgcccga	tggggggccat	ctcaccacag	gctacatgtc	tgacgtcaag	180
cggatatcag	ccacgtccat	cttcttcgag	tctatgccct	ataagctcaa	ccccaaaact	240
ggcctcattg	actacaacca	gctggcactg	actgctcgac	ttttccggcc	acggctcact	300
atagctggca	ccagcgccct	tgctcgccct	attgactacg	cccgcacatg	agaggtgtgt	360
gatgaagtca	aagcacacct	gctggcagac	atggcccaca	tcagtggcct	ggtggctgcc	420
aaggtgattc	cctcgccctt	caagcacgca	gacatcgta	ccaccactac	tcacaagact	480
cttcgagggg	ccaggtcagg	gctcatcttc	taccggaaag	ggnggaaggc	tgtggacccc	540

aagactggcc gggagatccc ttacacattt gaggaccgaa tcaactttgc c 591

<210> 92
 <211> 647
 <212> DNA
 <213> Homo sapien
 <220>
 <221> misc_feature
 <222> (1)...(647)
 <223> n = A,T,C or G

<400> 92
 gaataactag aaattttattg gatcaggttt cacatttgca tttttgaaaa ctactaccaa 60
 aaagatttca ccaattttaca actccatcat tagtaagaat gcctgtttgc ctatagtctg 120
 ccaaccctga atccttaaaa attttttgcca atctggtagg caaaatttct ttcttttctt 180
 tgaatattaa tgaggaggaa catcttttca tgtttcttgg ccatttgcac ttcttattat 240
 gaattgcttt tgcccatttt ccttttttta attatgaaag tctaatgact accttctcat 300
 tgtataaaaa acacagttct ttgaatagag agaccctttt ctccaatgct accaatcaca 360
 ttccacttac cacagtttaa catacatcct ctagtcacct ttccgtacga atatacatat 420
 acataaaaaac acttttttaca taaataggat ctcatattct gtagcttttt aaaatttttg 480
 nctcaaaaaa aganaacang gcttttaaatt tctttaatgg gtgaatatga ataaatacta 540
 tgaaaatgcc attattttatt cccttaattt ttttcctctc gctattacat tgccaaagna 600
 aacatcctat tcanatgtct ttgngcatgt gngngaatat ttcttta 647

<210> 93
 <211> 740
 <212> DNA
 <213> Homo sapien
 <220>
 <221> misc_feature
 <222> (1)...(740)
 <223> n = A,T,C or G

<400> 93
 gtcattgatag aggaaattaa tgcaaggaaa gaaaacaagc ccagttgggt ttgcttgtgc 60
 tactgcaagg cttttacaatc agatttatatc acatacatag atgaactcct gacctatata 120
 aatgcaaaac ccaacctgtt ctctatgtct ctaacggatc cacatctggc tctgaccgtc 180
 ttcttttgcc catgtctacc ataccagttc cgcttgactg gccagggaaa atgggaagga 240
 gccagaaatg ccatcatgac ccagtgggac cgaacattca aggtcatcaa agctcgagtt 300
 gtacaagagt ctccatctcc ctttgaaagt tttcttaaag tcttttagct tctggctttg 360
 cttgtggcta tttttctgat tttcctataa gtaaaagatc tcctaaatgg aagatgcaca 420
 gagtagattt acaatgctcc aattcctctc ttacagcaat attgccttca cagttataaa 480
 ctgtattcaa atagtaaagg ccacctctc gcttccttgg ctggccccag ggctaccact 540
 ggtattcctg agcctctccc agctccactt ctaatgctag agaatgataa ctaagacttc 600
 tgtgcatttg aaggttgttg gaaagttaca ggttcatttt anaaagaaag ctgttcttga 660
 cagcaactct gagccatcat acctctttcc ntataaacta ttttccagaa ctcnactaaa 720
 accccttact tcacaaatga 740

<210> 94
 <211> 608
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(608)
 <223> n = A,T,C or G

<400> 94
 ttttacaatc ctaggaaggc ccaccaatth catttcacgc gccagggcgg ctgcagttgg 60
 aggccgaggg cagccctctg ctcactgaat gtcttgcatg tgctgactgc tgcccgcagt 120
 gctgaacatg cccacccgcc caggcccagc actgcttggt gggtcagcat ctagtgctgc 180
 tgtcacatct ttgtctgcac agccagtagg attgcctcag ccaggggggt tatcagaagg 240
 tgtgcaaggc ctttggggga actgagcccc tatagtgggc agtctccttt accttcccac 300
 ctccctgaaa agcacagaag acagtgcctt ggtttgtggt ttgaagcaaa caagtcagct 360
 ttctggcttt gcccacaaac tgtgatggaa cataataaaa ctggagatat ggtttttaac 420
 actgcaaaaa ggaaaaagca tcaagtttct acttctggct ggaaagcaaa accaatctca 480
 gctgacaagg ctgggcaaac taagttttcc tgagcccatt ttcctttgag ccctgaccta 540
 nctgcctta cctcattaag gtttggttaa agcantggaa aggagganga ngcanggggtg 600
 gatggggg 608

<210> 95
 <211> 706
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(706)
 <223> n = A,T,C or G

<400> 95
 agaataatga gtacacaagt ttttgccat ccttttcctt aaaatataag aaataaagga 60
 gctcttacga aaaaccaggg ttagaccgca taaaaaataa agtgaacagt gaggtggtag 120
 caagacttct ttttagaaaa gaaagcattt acctgcctgt ctgtaagggt gaaatttcat 180
 cagtttgcaa acgataagaa atgcagactt gctcttgata gaaatgctta gaaacactct 240
 ggggggggaa aaagctcctt ccatatactg tgagacattt gttaagtga atctattggt 300
 tatcagcttt taaggataaa aaaggatatt taaaagttgg atatttagga tatttgagga 360
 tattccttta tgagctctcc atatccttct tgagaaactg gttaaaaaag gaataggggt 420
 tgagtgttac agagagtagt ctgaagattc ctgtgtaaaa gcaaagctaa caagcaatga 480
 agacatgaag caaaatacta atctaattgt gtaaagaaag gatattttta taagttcttt 540
 ctgcttgctg ctaagagttt gctaaagtgt catgaattat tctggttatt actaaagttt 600
 ctatgaaaca agtagatttt aagaataaat gtttctggaa aagaactatg ttatgatttt 660
 gtanaaatgt aaagattact tgagggtgtt aaaataaatt tttcat 706

<210> 96
 <211> 719
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(719)
 <223> n = A,T,C or G

<400> 96
 ccagggcctg tccctgcccc agtggcggct gcgctgggtt caggcccagg gggccctgna 60
 gaagctgtgc agctgaagag agggttcaaa cggaagccga gaacttgaca ctgttcaccc 120

caacacctca	cctccccccag	gacattttgga	agaaagcagc	gccaggattc	ctcggcagtc	180
gtccccaccc	gcacctgcag	tcccctcatg	tgctgttctg	ctgccccact	cagctcctgg	240
accctgtcct	ttcatcccgc	taaagcacc	cctaaaaccc	cttcatcact	ttcatttctca	300
gcaaaaagta	attgagcacc	tcctctaggg	gctggggagt	ccacactgaa	caaaagaaac	360
agaaaaccct	gtcttccagc	agttgagttc	tagggcaggg	agacagagtt	tacaagataa	420
ggaaaatata	tatgtagtat	gctgcaagtt	aactgctgtg	tgggaaatcc	agcagggggg	480
gggatgtgtg	atttgaattg	agggccacac	tgcccagggtc	gtgctccgtc	aaggggtgag	540
caggagcaac	aggggtgggt	gagtaanggc	ttgcagctgg	aggcaacagc	acatgcaaag	600
gccctgagcc	aggatgtgct	gcaataangg	ccactgaggg	ggacagtgtg	ngttgggggg	660
tgangaatgt	attaaaaaga	ngaaaaattg	ccttttataa	aaaaaaaaaa	aaaaaaaaaa	719

<210> 97

<211> 572

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1) ... (572)

<223> n = A,T,C or G

<400> 97

gcttggctaa	tttccaaatt	attgcataat	atgttctacc	ttaagaaaac	aggtttatgt	60
aacaaagtaa	tgggtgtgaa	tggatgatgt	cagttcatgg	gccttttagca	tagttttaag	120
catecttttt	tttttttttt	ttttgaaagn	gtgttagcat	cttgttactc	aaaggataag	180
acagacaata	atacttcact	gaatattaat	aatctttact	agtttacctc	ctctgctctt	240
tgccacccga	taactggata	tcttttcctt	caaaggaccc	taaactgatt	gaaatttaag	300
atatgtatca	aaaacattat	ttcattttaat	gcacatctgt	tttgctgttt	ttgagcagtg	360
tgcagtttag	ggttcatgat	aaatcattga	accacatgtg	taacaactga	atgccaaatc	420
ttaaactcat	tanaaaaata	acaaattagg	ttttgacacc	cattcttaat	tggataaatg	480
gatcaaaaat	agnggntcat	gaccttacca	aacacccttg	ctactaataa	aatcaaataa	540
cccttagaag	ggtatgtatt	tttagttagg	gg			572

<210> 98

<211> 520

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1) ... (520)

<223> n = A,T,C or G

<400> 98

tcagcctggg	gtcctccact	tctggaggcc	tctaggctct	atcagcaact	gggggacana	60
acagcagagc	tggagagtct	ggagctgcta	gttgaggcct	tgaatgtccc	atgcagttcc	120
aaagccccgc	agtttctcat	tgaggtagaa	ttactactgc	caccacctga	cctagcctca	180
ccccttcatt	gtggcactca	gagccagacc	aagcacatac	tagcaagcag	ggcaggagac	240
gctgcagagc	attacttgga	cctgctggcc	ctgttgctgg	atagctcgga	gccaagggtc	300
tccccacccc	cctccccctc	agggccctgt	atgcctgagg	tgtttttgga	ggcagcggtg	360
gcaactgatc	aggcaggcag	agcccaagat	gccttgactc	tatgtgagga	gttgctcanc	420
cgcacatcat	ctctgctacc	caagatgtcc	cggctgtggn	aagatgccan	aaaangaacc	480
aangaactgc	catactgncc	actctgggtc	tctgccaccc			520

<210> 99

<211> 470
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (470)
<223> n = A,T,C or G

<400> 99

tttttttcaa	agcaattgtg	acacctacct	gtggaccaag	gaaaaaacca	aatcatccnc	60
aatctttcag	ttccacaaca	acaccattat	aaaaacgagt	tgatctgaag	tggttccaaa	120
tctttcttat	cttcaagatc	tatgattagt	aattctgact	cgttgcaaag	caacatattt	180
tttgagatac	tggtgtgagc	cccgggagaa	tgtcagagtc	ctgtccactt	ggacatgggt	240
ggagggggagg	ttgggcaggg	ctgataaaaag	actagtacgc	ctcgtgtctc	ttcagtctta	300
ttttgcaaac	atcatctata	aagggtttttg	ctactccaag	ttttggtagc	ccaagctcat	360
caaagcatgt	gtctattatg	tgntctancat	agtaaaaatg	gctctaaatt	gcatataaat	420
gcccagattt	taataatcta	ttgnttcana	agaaaacaac	agnannngnt		470

<210> 100
<211> 570
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (570)
<223> n = A,T,C or G

<400> 100

agttttttatt	ttttttctgta	acgcccatec	tgccctgcagt	agaccctttg	ggcttaagan	60
gtccctacaga	ttctaatagt	ttcctctttg	tagatgtggt	aacatattta	aaatacttag	120
gacatggcct	ggtacgtgac	agatgggtgt	taacatcatt	gtcatatgaa	gaacactctc	180
tgtggggccaa	tggaggtgtg	ccttaaaaatt	cttcatcttg	cctgattttg	tttgcataac	240
ttctggagag	tctgtgtcct	cctcatctag	gccacctttc	cattttttgt	gaaagatgac	300
cttgteccctg	cagtctccgc	caggagtcac	acagcttttc	acagcagctg	ccatctttaa	360
ttctttttcc	tgagattcca	gcaagggttg	gacattgtca	ctttttttgt	ctagactctt	420
ttaaattttc	tgcatttgcc	tgaaaagcac	ccctgtaaga	atagatttct	catggctcta	480
aaaattatc	ccaagaatac	cttacttggg	tcaaaagcag	actgtttctc	ttcatttcat	540
ctcaaatacan	acttctgggc	aagatgttct				570

<210> 101
<211> 365
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (365)
<223> n = A,T,C or G

<400> 101

atcaataaac	caaggtaatg	cctnagggtt	ccctcccagt	cctcncatca	gctctggcct	60
catcaccaag	gtcacanagn	acacagggga	gggggaanac	ccacncacac	tccttggaat	120
gggtcctgtt	atttatgctt	ggngcncagn	catattanaa	gaaaaaaaaa	agctttgtat	180

tntttttccn catatnatgg ntgctgttta cacaccctgc caatgcntta gcnntggaga 240
gcttttttgca atatgnnggg gaaaggggag ggaggggaatg aaagtgccaa agaaaacatg 300
tttttaanaa ctnggggttt atacaataga atgttttcta gcagaaaaaa aaaaaaaaaa 360
aaaaa 365

<210> 102
<211> 546
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (546)
<223> n = A,T,C or G

<400> 102
antgacacac tagccccctg cgcctcggc atgaacagca tgggcaccat ggatggcacg 60
ctgatggtgg taggaagcan aagggtcct gggcagaaag ggcttggtnt gaggtgaaac 120
ccnccttca agcctgggat ggcctgaggg ctggggggccg ggctgccagc tntgggtagt 180
ntgtgggtgg agtganaatt tatggtgctt tttccgggcc tgcctcatgga ccaatcagca 240
tgcacttcct cccttntgag cccataaaaa ccctggattc agccagactt ggacanatgt 300
caatactacc aactgtggga agaagctacc cacttcagga ctcccttgact cctcaggatg 360
acctgcctac anaaagaagc taccactat gggtnntntn tntgctgana gctggatact 420
ngtcaggatg acctgcctgc anaaanagct acccnccatg ggtntcctnt gaactgttct 480
gttgcccaat aaagctcctn tcttttttgc tcnccctcca aaaaaaaaaa aaaaaaaaaa 540
aaaaaa 546

<210> 103
<211> 376
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (376)
<223> n = A,T,C or G

<400> 103
ctcaactccc ncttcacctt atgtaatgtc tgggcctgag atttctctct ggctttantt 60
tcttgatgtt cctgtgatat ggcttctgcc agcaatgaaa acaaggctcct tatagtagtn 120
ctccatggag tccagttcat cctggtggcg tctcctttgt tcatctcgt tttctttggc 180
atagtttctt aggtctcgta atctttgctt ttgaatgttt aaaccttctt caaacagttt 240
cttaaatatc atttcttccc gggtcctcat tctcatcatt tttgcacaca actgaactct 300
ataatcatca taatattttc gagcacgaac gatttgctgn cgatnttctt ttatctttga 360
ttgggncaac ctttgc 376

<210> 104
<211> 700
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)... (700)
<223> n = A,T,C or G

<400> 104

tttgaaaata	cagattttta	ccaacttttg	attctttttt	agttatatgt	ttgtctttcc	60
tttttaaatt	gttcaaaact	attttttaat	ggtcaagtta	ctaacacttg	aaaatcagat	120
actgcaccaa	atacagtgtt	tttccgtagt	gtttttaatg	agtgcaccta	ttactactgt	180
gcgagaattc	atgtttttacc	agtcattgtt	atattacaaa	cagacttgca	tgattaacca	240
gttgttacac	ttactttttc	aagttggagt	atatatgact	cagtgcagac	tggtctctct	300
tatgtgaatg	cacacatgca	gaaatgcaga	gtcaatttta	catgcccata	aagacatttg	360
taaagaattc	agctcttatg	gtctgttgta	taaatgtgta	tctaggcact	ttggaatttg	420
acctcacaga	tgttacaact	tgatcagtcg	tttgacctaa	tttgtggnag	ctatctgtat	480
gttttgcaat	cttaatacag	acatgccttc	caaaaagatt	aatacagaac	catcctgccg	540
ttttggataa	gtctatccag	ctgtggaaag	ggcaacctgt	ggtttctctg	tactgggtgt	600
taatggggga	agaatatgaa	cagctttaaa	gagctgtgta	ttgggggtac	tactattaaa	660
aaataagatc	tgacagagtc	tgactggcct	ttgggtggcc			700

<210> 105

<211> 729

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(729)

<223> n = A,T,C or G

<400> 105

taaacccctc	acactgagaa	ctgcttgtgt	gggagagagc	tggttggggt	gatcttttcc	60
gagtgtgact	tacctccttc	aaggggatgt	ttaagcttct	cgggcagaag	tggtgtgtct	120
attcctgaca	ccaaacaccg	tggtatatgt	ggttgtcaca	ctcagctagt	gatgataaag	180
gtgttcttaa	atatgttagc	tttcagtttt	cctgaggaag	caattttatg	gatacttccc	240
cctccttctc	aagtgaggaa	tagcagagca	aattttatct	ggaacttaaa	ccaatagtta	300
taaccaatag	tttcaacctc	ctgcctcacc	actgcttctc	tcttgagctc	tttccccaca	360
cctcaaaaag	agtacaaagt	gattccatct	gcagaggtaa	attcttttgt	taaaaaagta	420
ctgtttttct	tatcttttct	ggttctccta	ggtatcagaa	caaggtttaa	tagaaatcct	480
taaaaaagta	agccaacaaa	cagaaaagac	aacaacagtg	aaagtaagtg	tccccanatg	540
cttgtggcaa	atgaaaagat	ggatacttta	aagattaatg	ttgagtatac	atctaccaca	600
catatttttc	agcccanaga	cattttcctt	ttgtcaaaca	cgtgaaagtt	tggggagaaa	660
ngctgaatct	gttggggggag	ggttctaatt	tttataggct	cttgactcca	ttccccctt	720
ttaattcac						729

<210> 106

<211> 481

<212> DNA

<213> Homo sapien

<400> 106

gaaaagcaga	gtcagctact	gagagctctg	gaaagaagga	tgtcaagaag	gtgaaatcct	60
aaagcctaga	aataaagttt	taaatgggaa	actgctatct	tcttgttccc	atcttcaa	120
gctaattgcc	agttccagtg	tattcatggg	actctaagaa	aaatctcttt	ggttttgatt	180
tcttgcatat	tttatatatt	ttacaatgct	ttctacctga	aatgtgtagc	tttatatttt	240
atggcattct	agtatttttg	tgtactgtat	tttgtgcatt	tcatgtcttc	atcaaaatcc	300
tctcagtcct	tgttcttttg	aagcttgtgc	tgagggttta	gcttttctat	gttttatatg	360
ccgctgcttt	gaaagagAAC	ctagattcta	tagttgtatt	attgttggtt	catacttta	420
atttatatgg	ctgtggaaaa	acgaattaaa	atgttttgag	gagaaaaaaa	aaaaaaaaa	480
a						481

<210> 107
 <211> 519
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(519)
 <223> n = A,T,C or G

<400> 107
 cagctcctct cctgccagag ctaggcaggc gccgaagtag ccgcatggcc ccgtcagant 60
 accccagggg ctggagagcc aacctcaaag gcaccatccg tgagacaggc ctggagacca 120
 gctccgggtgg gaagctggct ggccatcaga agaccgtccc cacggctcac ctgacttttg 180
 ttattgactg caccacaggg aagcagctct ccctggcagc aaccgcatca ccacccaag 240
 cccccagtcc caatcgaggg cttgtcacc caccaatgaa gacctacatc gtgttctgtg 300
 gggaaaactg gcccctctg actcgggtga ccccatggg tgggggatgc cttgccagg 360
 ccagggccac cctgccgctc tgcagagggt ctgtggcctc agcttccttc ccagtcagcc 420
 cgctctgccc ccaggagggt cccgaggcta aggggaaacc cgtgaaggct gcgcctgtga 480
 ggtcttcaac ttggggaaca gtcaaggact cactgaaag 519

<210> 108
 <211> 669
 <212> DNA
 <213> Homo sapien

<400> 108
 ggacaatgaa gactgaacta tcgcacatta cctaagaaag atgggaattg acatgcacat 60
 cacaattgta tacacaacag aaattattga atcatgagat atacattccg gtgtgtgaca 120
 gattggcaca tgacataatc tgggttcttt atagactcag ttgttttggg gcgatctaga 180
 ttatcaagag aagagccttc atgcttagct ttatcttgta ccaaccacca gggcccttgc 240
 ttctgagcag gaagcagctg gggaatagc tctttctctt aatgacttcc aacatagttc 300
 tctcaaacct tactcctcca gaaggccacc ctacctggc tatggctact tcagaaaaaa 360
 cttggcctct ggtataatag agcagaatca tcacctcaca ttctatttca agccaaagtc 420
 aatatctcaa aggctgggtc tgtgatttat ttggctcttg ggagctccta ctgaaagtgc 480
 tgaaatgtcg tactgacact tcagacttat agctacctag actccaagta agatttatct 540
 ctgactggag ggtttctcct attaaaaacc aaagagtgtg ggggtgccttc acctgctagg 600
 taatcttcta tgccctaata ggaagaatgg gagcagcaga caagtaagtg caggaaggag 660
 aaccaaagc 669

<210> 109
 <211> 349
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(349)
 <223> n = A,T,C or G

<400> 109
 tttcaagccg gatthttgggc ctgcttaaacc cacttaaagt tanttaatga cagatgggtt 60
 gaggttttaa agtcttctgg agaaagcccg ccagagaaca ttccctttga agcccatgt 120
 aaaaatacgt gtgggagaga aagtgttttc tctgacttct gctgacagtg gctaaaactc 180

tgaactgtca	ggagtattca	aaataagact	gccttgtagg	taagcctgtg	gtagcttttt	240
tgagcacagg	ataaaatact	tgagtctttg	cttaaagtgt	actttctcaa	tgaggctttg	300
tatgactaaa	taaaatctgt	ataatcccca	aaaaaaaaaa	aaaaaaaaaa		349

<210> 110
 <211> 337
 <212> DNA
 <213> Homo sapien

<400> 110						
ggcctttccc	actgggtccat	ctggttttct	ctccaggggc	ttgcaaaatt	cctgacgaga	60
taagcagtta	tgtgacctca	cgtgcaaagc	caccaacagc	cactcagaaa	agacgcacca	120
gccagaagt	gcagaactgc	agtcactgca	cgttttcatc	tctagggacc	agaaccaaac	180
ccacccttcc	tacttccaag	acttattttc	acatgtgggg	aggttaatct	aggaatgact	240
cgtttaaggc	ctattttcat	gatttctttg	tagcatttgg	tgcttgacgt	attattgtcc	300
tttgattcca	aataatatgt	ttccttccct	caaaaaa			337

<210> 111
 <211> 552
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(552)
 <223> n = A,T,C or G

<400> 111						
atthttgtcaa	gtttcttttaa	tggctgaaca	gaaagaagct	tcaagtaatg	gagaaggcat	60
tgtctgagtg	cagctgcttt	cctggacgcc	tgtgccgttc	ctgtcttcca	aatcctatgc	120
ttggaggccc	ctggagggtac	atthtttgcca	ggaaccaacc	tgaccttaaa	aagatgagtg	180
tgacacagcc	ggctgggcag	gaggatggag	gtgccacagg	acaccacctg	cccacgccct	240
ggccagcctg	gccatgctgc	cgagtgcggg	gaggccaccc	caccagagg	gcacagggca	300
aaccctaagc	acgggggtat	tgcccttgaa	gccccagggg	atgccctgtg	ccggatcctc	360
atgccctcatt	gactagcctg	cttgctgaag	gagcccaggg	gcctgagcct	gcaacactgc	420
aaggggtgag	aagggcatgc	tgctgtgggc	gccactggac	tcaaacctca	cattanaagc	480
tacaaagaac	cccaaagtgc	cttcaanagc	cccaccaacc	cccaagccan	gctcatncct	540
gacacantga	at					552

<210> 112
 <211> 115
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(115)
 <223> n = A,T,C or G

<400> 112						
gtcctaggta	canaaacttc	ccaaacaana	actatgcncg	anonctaccc	cttcgagtg	60
tctnctttct	gctccgggtc	ccatggnttg	ccccattta	taaggtaacg	tgga	115

<210> 113
 <211> 649

<212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1) ... (649)
 <223> n = A,T,C or G

<400> 113

gcctgattaa	aaactaagca	gaagtagttt	taacanaaat	actcatgaaa	atgtttngaa	60
actgaaattt	aaacaactgt	aatattaagg	aaaccagaat	caataaatca	ctgtcttgcc	120
agcacagcta	cagagtaaca	tgattcaggg	gaggaaagtt	ccttagagtt	acttttataa	180
ttcttttttt	tttcctctta	ggtttanaaa	tcttaciaat	ttaaacttta	tcctttttaa	240
attatttgaa	cataatttag	atattgtaag	cttaaaatac	aaatgtttat	agataacctc	300
tttaccataa	actaatccct	ggcaagccat	ggctctcttt	tttttttggt	gtttaaagcc	360
tgtaaacagt	ttttctgaat	gatcatgaac	ttttcttggt	tttagcacta	ggatttagct	420
atgaagagag	ctcataggct	ttcaggtgct	aattgagatc	tgccctgtta	gagtcttggt	480
gtgctagatt	ggtcacattg	acaccagtgg	cagggaaggc	atctatgagt	ttgatgcttt	540
ttatcacaca	cttcagntgt	ttagaaagtt	attaccaata	cttttaaaca	acactccaag	600
aaaatttgct	atatttcttt	ctcatcacta	cagagagagt	agatttccc		649

<210> 114
 <211> 650
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1) ... (650)
 <223> n = A,T,C or G

<400> 114

tggcgattgg	tggtggcggt	ctggctcagc	tgggcagggg	gtaactttac	tgatttgggg	60
gtggttttta	gtttaatttt	tcttttctag	cttcccacgc	acggtcagtg	cgcacgttgt	120
aatcagctga	ggccatgtca	ggagacggag	ccacggagca	ggcagctgag	tatgtcccag	180
agaaggtgaa	gaaagcggaa	aagaaattag	aagagaatcc	atatgacctt	gatgcttgga	240
gcattctcat	tcgagaggca	caggttttagt	gatataggat	tacatttcct	tctctatggg	300
tccaatcaca	ctacttggtt	ctgcagtga	taatattttc	ataatcctaa	cattgtaaat	360
gctgtttatt	ggttttcaat	tttagaatca	acctatagac	aaagcacgga	agacttatga	420
acgccttggt	gcccagttcc	ccagttctgg	cagattctgg	aaactgtaca	ttgaagcaga	480
ggttactatt	ttattttatt	ttttcttata	tcagtatttg	cagcattcac	tgtagtgata	540
gaaaacaaag	ttangaacat	agccaattan	gacaaggagg	atttaaagt	gtcttacctt	600
tattttgtaa	aataggata	aaggagtaat	taaaatgaat	ttttgaaatt		650

<210> 115
 <211> 403
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1) ... (403)
 <223> n = A,T,C or G

<400> 115


```

gtattatgac tttaaaaacc ccattattga aaagtacctg acaaggcagc tcacgaancc      60
caggcctgtg atcctggacc cggcggaccc tacnngaaac ttgggtggng gagacccaan    120
gggnnggagg canctggcac angaggctga ggctgnctg aattacccat gctttaagaa    180
tngggatggg nccccantga gctcctgnat tctgctgggtg anacctcctg ctctctccct    240
gccattcatc cctgcccctc tccatgaagc ttganacata tanctggaga ccattctttc    300
caaanaactt acctnttgcc aaaggccatt tatattcata tagtgacang ctgtgctcca    360
tattttacag tcattttggg cacaatcgag ggttnctgga att                      403

```

```

<210> 116
<211> 397
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1) ... (397)
<223> n = A,T,C or G

```

```

<400> 116
cgaaagaaaa aaagtgatat tggaccctgg aaagattttg aaacttgagt ggtttgataa      60
cccttctatg tattgtaggg agaaaaaaaa aagttttattt tattccactg tcctccctta    120
aaagcatcat ttgancaata aatgaatatt gtcttttaaac caagggttag ggaattttcc    180
tctctctctc tctctcctct ctctttctgt tcaaagaact tcaaacattt gggaccacct    240
ggtattctgt attttcactg gccatattgg aagcagttct agttgcattg tattgagttg    300
tgctggcagt agtttccatg cctgtcaatg tatcatagtc ctttgttgcc cagataaata    360
aatatttgat acgcttttaa aaaaaaaaaa aaaaaaaa                      397

```

```

<210> 117
<211> 59
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1) ... (59)
<223> n = A,T,C or G

```

```

<400> 117
cacttatggg gacaatggga agactcttct tttncactgg actgtacctg gacntnaa      59

```

```

<210> 118
<211> 751
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1) ... (751)
<223> n = A,T,C or G

```

```

<400> 118
catcaacata cccgctttat tgctgactca tgacaactaa tgggaagaca tggctcagat      60
gtgcagccac agtgagcttc tgaacatttc ttctcagact aagctcttac acacagttgc    120
agttgaaaga aagaattgct tgacatggcc acaggagcag gcagcttcct gcagacatga    180
cagtcaacgc aaactcatgt cactgtgggc agacacatgt ttgcaaagag actcagagcc    240

```

```

aaacaagcac actcaatgtg ctttgcccaa atttaccocat taggtaaate tccctcctc 300
ccaagaagaa agtggagaga gcatgagtcc tcacatggaa acttgaagtc agggaaatga 360
aggctcacca attatttgtg catgggttta agttttcctt gaaattaagt tcaggtttgt 420
ctttgtgtgt accaattaat gacaagaggt tagatagaag tatgctagat ggcaaagaga 480
aatatgtttt gtgtcttcaa ttttgctaaa aataaccag aacatggata attcatttat 540
taattgattt tggtaagcca agtcctattt ggagaaaatt aatagttttt ctaaaaaaga 600
attttctcaa tatcacctgg cttgataaca tttttctcct tcgagttcct ttttctggag 660
tttaacaaac ttgttcttta caaatagatt atattgacta cctctcactg atgntatgat 720
attaagttct attgcttact ttggatttct a 751

```

<210> 119

<211> 591

<212> DNA

<213> Homo sapien

<400> 119

```

aggcttcttg tcacactgaa cacatccagc cacaggcacc agctgggttg gaccagcagc 60
ccccagcatc ctcttgcaact ggctggcaca aaaagaaacc tgctgtatac ccccaaagt 120
gtccctttcc caattacctc tggggctctc tgctgcttgc ctctgctgct ctggactggg 180
agagcttctg tcctgtgctg catgggtatt tagactgtgg gggagatgcc cttcttata 240
gcaactggagg aggaaaacaa attcttgtcc ccctcagaat gagagtggct cttcttgatt 300
tgcaagggca ctatggtcag ggcaaaggca tggcccaggt gtttaagtac agggtgacgt 360
gtgcctatgc aatgggggtg taaggcaggc acgaagagtc caaaaaatct aggtggcctc 420
tcagctctgc cacctctagc tgcattgacct tgggcaagct atgtaacccc aattgcctgc 480
tccattaaag actgtgaagg tagaatgttt gtaaagctct taacagtatg taagccttca 540
ataaatttca gttttccctc tgttttcttg atcaaaaaaa aaaaaaaaaa a 591

```

<210> 120

<211> 652

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1) ... (652)

<223> n = A,T,C or G

<400> 120

```

attttgcagt acattaaaac tgaggcccag agatgtgatt tgcttgaggc cacacagcta 60
gatttttggg ggaagtgggc cttgaacaca gtgtactttc tgcagtttct gactgtaaaa 120
cccagtgtct gctctctgag ttccatttcc aagccccctt ccatcttgga cctatgtggg 180
ctccaccata ttcacacacc accgccacca cttgccaatg cctctcttaa agcaatatac 240
ccattcgttc tcttattggg aactggatgg atgaagcccc aaattcagcc ccaccacag 300
agaagccttc ctacactcag cctctgtcca cccttggcaa atctttcaag ctctctcctc 360
caggaaagtg gggccccaac tcagtcactc caccctctc caggtccctg aggtgggttc 420
tactgtatcc ccatcacctc cacaactcca ctcacctctg acggctccat ccacctcacc 480
agttggaagg cttgtgggtt cagagaggag caatgctggg cagcgctgcc cagactccag 540
tgtttacaga tcaccagcat ttacaaccaa tccaatggcc agaagcctcc tctaaccana 600
aggagtctct aaggggcaga tgggggtgtg agtagtcggg gagtccggat tg 652

```

<210> 121

<211> 407

<212> DNA

<213> Homo sapien

<400> 121

ctcttttagta	tgacactggc	aactgacggc	actgcgata	tgtttggaat	aagggtcaaa	60
agaagaggct	gttctaagaa	agactaccga	gtaatcaatc	acctcctcag	agaaagtctg	120
ctcaggaaac	tctcctccca	ccagccctg	cacttggttc	cttgcggtgc	ctctggctgg	180
aagctgttgt	ccccaaacttc	agaaagtgtc	tgacagatttc	acaggetctc	tcttggttca	240
agggaccagc	tctgtgaaca	cggcaagtaa	caacagagag	aaaacgtgag	gagaaaagag	300
agaccgggat	tttgaaatcc	tgtttctctga	atgccatctc	accaggcaca	cgaaagaata	360
aaattaagga	atcaagaaaa	aaaaaaaaaa	aaaaaaaaaa	aaaaaaa		407

<210> 122

<211> 752

<212> DNA

<213> Homo sapien

<400> 122

caaateccctc	ctatccaggc	ttttctgtct	ctaatacccc	aagcgttacc	cctgctcttc	60
cctcattccc	ggggctgcag	gcgcccctcta	cagtcgcagc	tgtcacacca	ctacctgtgg	120
ctgccacagc	cccatcccca	gctccagtcc	tcccaggatt	cgcctcagca	ttcagttcca	180
atttcaactc	cgctcttggt	gcacaagccg	gtttatcatc	tggacttcaa	gctgcaggca	240
gttctgtttt	tccaggcett	ttgtccctcc	cgggtatccc	tgggtttcct	cagaatcctt	300
cacaatcatc	cttgcaagaa	ttacagcata	atgcggctgc	acagtcagca	ttgttacagc	360
aggtccattc	agcttcggct	ctggaaagct	atccagctca	gcctgatggg	tttcctagtt	420
atccttcagc	gccaggaaca	ccattttctt	tgcaaccaag	cctgtcccag	agtgggtggc	480
agtgaatact	tttaactttt	attctccttc	agagcaacat	cagaattgcc	tgagaactgc	540
aatgaacaat	ctgacaaatg	tgaagctggc	caaaagtcgg	aaaatgagaa	tgagggtaat	600
cctggagaaa	ttgtgacaac	aatttgaaaa	ttgtgggtgc	attttaaagt	gtgaacactc	660
ccctatgtaa	atatgctgac	aataaattgg	atggagaaatg	gtatttaaaa	agtgtttgga	720
gacttttcac	ctgtcctata	aaaatttgaa	tt			752

<210> 123

<211> 401

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(401)

<223> n = A,T,C or G

<400> 123

atgaaccaaa	tagaaataag	catgcaacat	gaacagctgg	aagagagttt	tcaggaacta	60
gtggaagatt	accggcgtgt	tattgaacga	cttgctcaag	agtaaagatt	atactgctct	120
gtacaggaag	cttgcaaatt	ttctgtacaa	tgtgctgtga	aaaatctgat	gactttaatt	180
ttaaaatctt	gtgacatttt	gcttatacta	aaagttatct	atcttttagtt	gaatattttc	240
ttttggagag	attgtatatt	ttaaaatact	gttttagagtt	tatgagcata	tattgcattt	300
aaagaaagat	aaagcttctg	aaatactact	gcaattgctt	cccttcttaa	acagtataat	360
aaatgcttag	ttgtgatnaa	aaaaaaaaaa	aaaaaaaaaa	a		401

<210> 124

<211> 103

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(103)

<223> n = A,T,C or G

<400> 124

aagataactt gctgggacca catgcctgat tgccacagct gtcacacgt tattttaatga 60
acctagtgtg gaagacagtg aaaaggggtcc attgacngtg gcn 103

<210> 125

<211> 1024

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(1024)

<223> n = A,T,C or G

<400> 125

gagggcagtg aggagcgagg agcgggcaga ggcagctccg gcggccgaga ggagggagcg 60
cggcgcagag aggagggggt tgcgccccgt agaaatgtca atcagagcct ggacccctgc 120
gctcccgcac cagccccgcg ctccgccctg cagaagccca agatctgggc cctcgcggag 180
actgccacaa gcccggaaca cccgcgcgcg tcgcctcccg gcgcgggggg gtctccaccg 240
ggggcagcgg tcgcgccttc cgccctgcag ctctctcccg ccgcgcgcgc cgccgcgcgt 300
cacagactgg tctcagcgcc gctgggcaag ttcccggtt ggaccaaccg gccgtttcca 360
ggcccaccgc ccggcccccg cccgcacccg ctctccctgc tgggetctgc ccctccgcac 420
ctgctgggac ttcccgagc cgcgggccac ccggctgccc ccgcgcctt cgctcggcca 480
gcggagcccg aaggcggaac agatcgctgt agtgccttgg aagtggagaa aaagttactc 540
aagacagctt tccagcccgt gccagggcgg cccagaacc atctggagc cgccctggtc 600
ttatcggtc tctcctcatc ctagttcttt aaaaaaaca aaaaaacaaa aaaaactttt 660
tttaatcggt gtaataattg tataaaaaaa atcgctctgt atagttacaa cttgtaagca 720
tgtccgtgta taaataccta aaagcaaaac taaacaaaga aagtaagaaa aagaaataaa 780
accagtcctc ctcagccctc cccaagtgcg ttctgtggca cccgcattc gctgtgaggt 840
ttgtttgtcc gggtgatttt ggggggtgga gtttcagtga gaataaacgt gtctgccttt 900
gtgtgtgtgt atatatacag agaaatgtac atatgtgtga accaaattgt acgagaaagt 960
atctatTTTT ggctaaataa atgagctgcc tgccactttg nctataaaaa aaaaaaaaaa 1020
aaaa 1024

<210> 126

<211> 214

<212> PRT

<213> Homo sapien

<400> 126

Arg Pro Arg Ile Arg His Glu Glu Gly Ser Glu Glu Arg Gly Ala Gly
1 5 10 15
Arg Gly Ser Ser Gly Gly Arg Glu Glu Gly Ala Arg Arg Arg Glu Glu
20 25 30
Gly Leu Ala Pro Arg Arg Asn Val Asn Gln Ser Leu Asp Pro Cys Ala
35 40 45
Pro Ala Pro Ala Pro Ala Ser Ala Leu Gln Lys Pro Lys Ile Trp Ser
50 55 60
Leu Ala Glu Thr Ala Thr Ser Pro Asp Asn Pro Arg Arg Ser Pro Pro
65 70 75 80
Gly Ala Gly Gly Ser Pro Pro Gly Ala Ala Val Ala Pro Ser Ala Leu
85 90 95

Gln Leu Ser Pro Ala Ala Ala Ala Ala Ala His Arg Leu Val Ser
 100 105 110
 Ala Pro Leu Gly Lys Phe Pro Ala Trp Thr Asn Arg Pro Phe Pro Gly
 115 120 125
 Pro Pro Pro Gly Pro Arg Pro His Pro Leu Ser Leu Leu Gly Ser Ala
 130 135 140
 Pro Pro His Leu Leu Gly Leu Pro Gly Ala Ala Gly His Pro Ala Ala
 145 150 155 160
 Ala Ala Ala Phe Ala Arg Pro Ala Glu Pro Glu Gly Gly Thr Asp Arg
 165 170 175
 Cys Ser Ala Leu Glu Val Glu Lys Lys Leu Leu Lys Thr Ala Phe Gln
 180 185 190
 Pro Val Pro Arg Arg Pro Gln Asn His Leu Asp Ala Ala Leu Val Leu
 195 200 205
 Ser Ala Leu Ser Ser Ser
 210

<210> 127

<211> 507

<212> PRT

<213> Homo sapien

<400> 127

Met Ser Phe Pro Gln Leu Gly Tyr Gln Tyr Ile Arg Pro Leu Tyr Pro
 1 5 10 15
 Pro Glu Arg Pro Gly Ala Ala Gly Gly Gly Gly Gly Gly Ser Ser Ala
 20 25 30
 Gly Gly Arg Ser Gly Pro Gly Ala Gly Ala Ser Glu Leu Ala Ala Ser
 35 40 45
 Gly Ser Leu Ser Asn Val Leu Ser Ser Val Tyr Gly Ala Pro Tyr Ala
 50 55 60
 Ala Ala Ala Ala Ala Ala Ala Ala Ala Gln Gly Tyr Gly Ala Phe Leu
 65 70 75 80
 Pro Tyr Ala Thr Glu Leu Pro Ile Phe Pro Gln Leu Gly Ala Gln Tyr
 85 90 95
 Glu Leu Lys Asp Ser Pro Gly Val Gln His Pro Ala Thr Ala Ala Ala
 100 105 110
 Phe Pro His Pro His Pro Ala Phe Tyr Pro Tyr Gly Gln Tyr Gln Phe
 115 120 125
 Gly Asp Pro Ser Arg Pro Lys Asn Ala Thr Arg Glu Ser Thr Ser Thr
 130 135 140
 Leu Lys Ala Trp Leu Asn Glu His Arg Lys Asn Pro Tyr Pro Thr Lys
 145 150 155 160
 Gly Glu Lys Ile Met Leu Ala Ile Ile Thr Lys Met Thr Leu Thr Gln
 165 170 175
 Val Ser Thr Trp Phe Ala Asn Ala Arg Arg Arg Leu Lys Lys Glu Asn
 180 185 190
 Lys Met Thr Trp Ala Pro Arg Ser Arg Thr Asp Glu Glu Gly Asn Ala
 195 200 205
 Tyr Gly Ser Glu Arg Glu Glu Glu Asp Glu Glu Glu Asp Glu Glu Glu
 210 215 220
 Ser Lys Arg Glu Leu Glu Met Glu Glu Glu Glu Leu Ala Gly Arg Gly
 225 230 235 240
 Gly Gly His Gly Gly Arg Gly Ala Gly Arg Arg Arg Arg Asp Glu Glu

505